GDC
Genetic Diversity Centre Zurich

Genetic Diversity: Analysis

Next Generation Sequencing

Monday, June 17, 2024

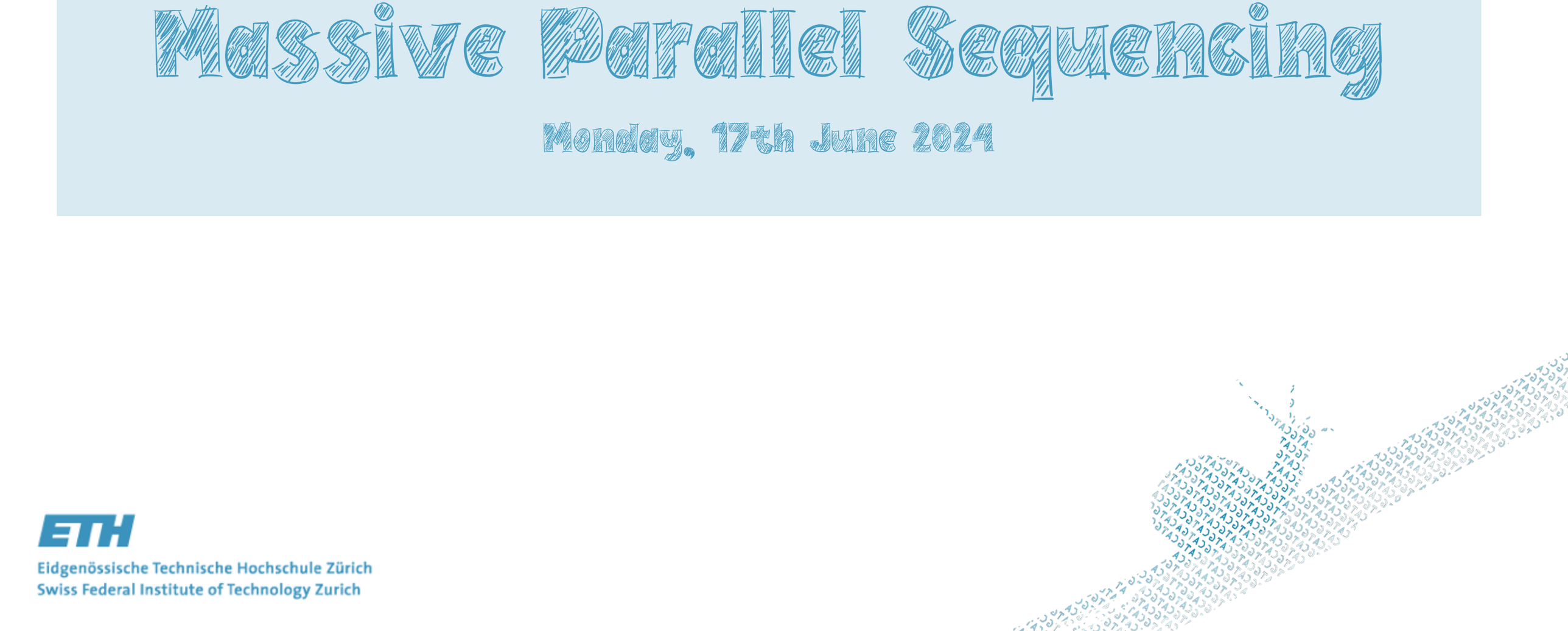
ETH
Eidgenössische Technische Hochschule Zürich
Swiss Federal Institute of Technology Zurich

GDC Genetic Diversity Centre Zurich

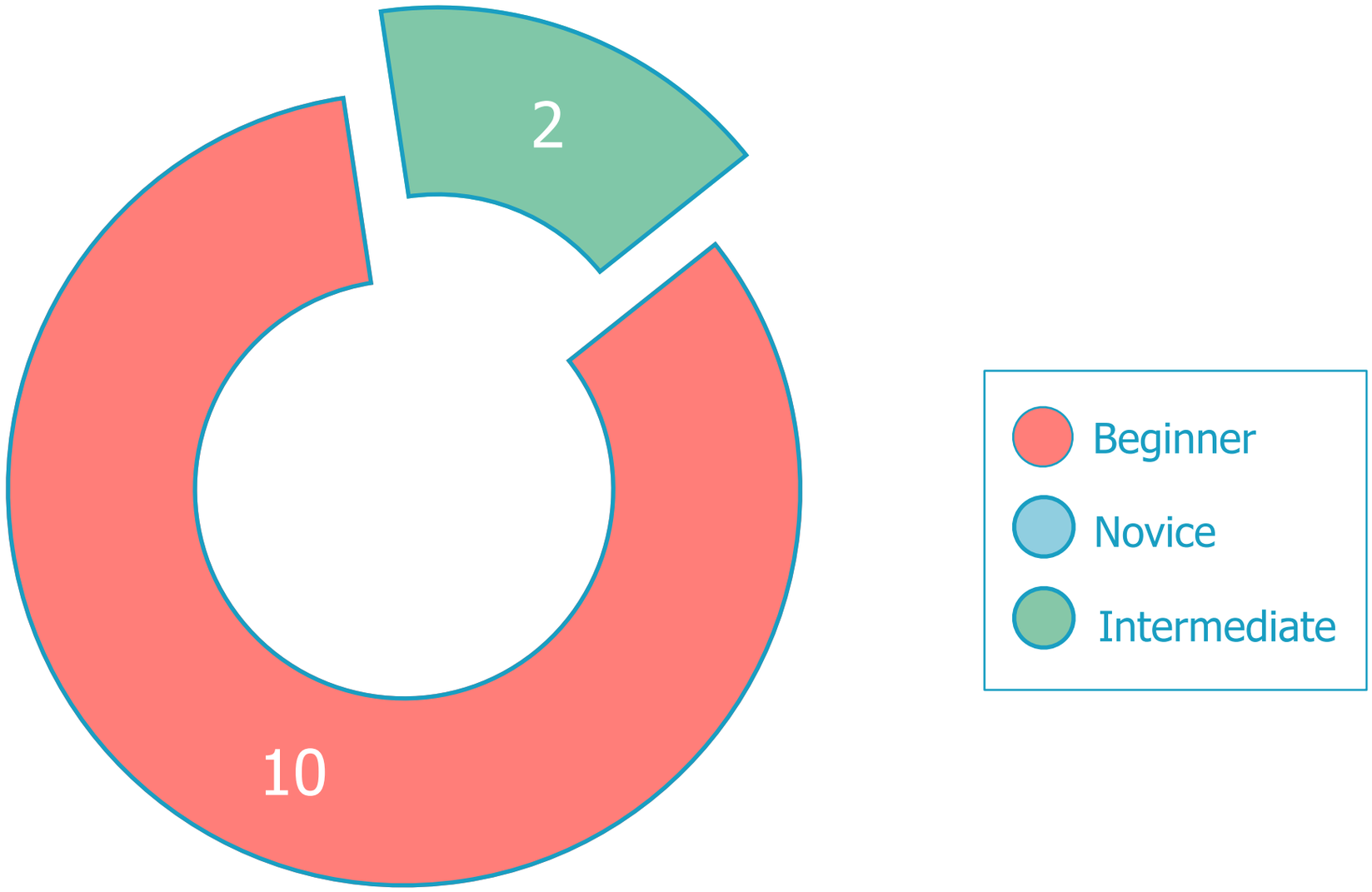
Genetic Diversity: Analysis

Massive Parallel Sequencing

Monday, 17th June 2024



ETH
Eidgenössische Technische Hochschule Zürich
Swiss Federal Institute of Technology Zurich


2
10
Beginner
Novice
Intermediate

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First Generation Sequencing

Sanger Sequencing

Second Generation Sequencing

Next Generation Sequencing (NGS)

Third Generation Sequencing

Single Molecule Sequencing

Fourth Generation Sequencing

??? Sequencing

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First Generation Sequencing
Sanger Sequencing

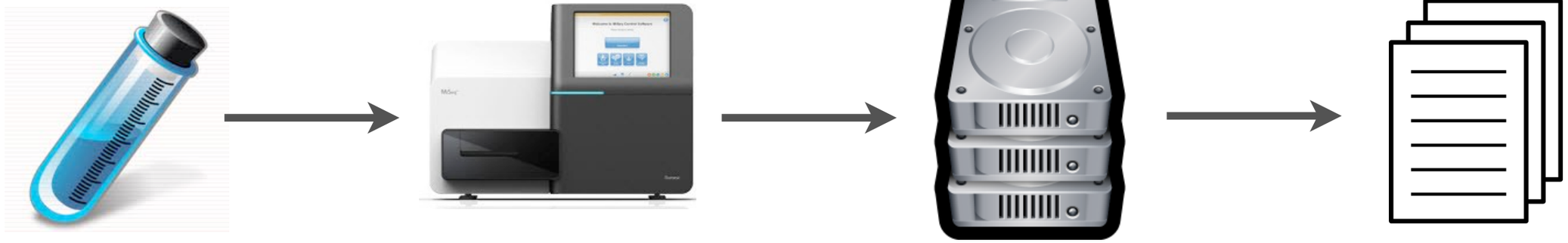
Second Generation sequencing
Next Generation Sequencing (NGS)

Third Generation Sequencing
Single Molecule Sequencing

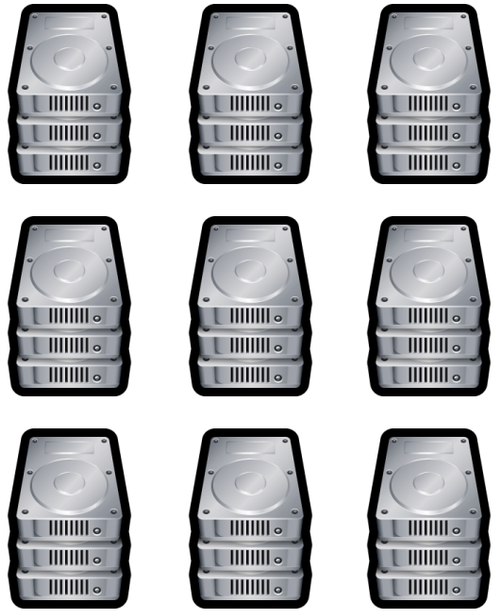
Fourth Generation Sequencing

Massive Parallel Sequencing

Next Generation Sequencing **Hype**



Next Generation Sequencing **Reality**




BREAKTHROUGH
AT THE HUMAN GENOME PROJECT
EUREKA! THE NEXT ONE'S A "G"!
SNAK-O
HIMELBLAU


nature
the human genome
Science
THE HUMAN GENOME

Actually, that's the coffee machine...this is the next-gen sequencer.

The **First Law of Technology** says we invariably **overestimate** the **short-term impact** of a truly transformational discovery, while **underestimating** its **longer-term effects**.

https://www.scientificamerican.com/

"The Human Genome Project has had a considerable effect on research and society more generally, but questions about what a human genome reference is today and how it can benefit human health remain to be resolved."

Source: Rood and Regev (2021) The legacy of the Human Genome Project. Science Vol 373, Issue 6562, pp. 1442-1443.

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(A-)Typical Workflow

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??? Question

Design

DNA or RNA (or Protein)

Library Prep

Sequencing

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MPS data (fastq, fastq.gz, fast5, bam)

Data Set(s)

Data Conversion

Data Storage (Archive)

Quality Check (QC)

Filtering / Trimming

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Error Correction

Merge Read Pairs

(High(er)-Quality) Reads

Assembly

Contig

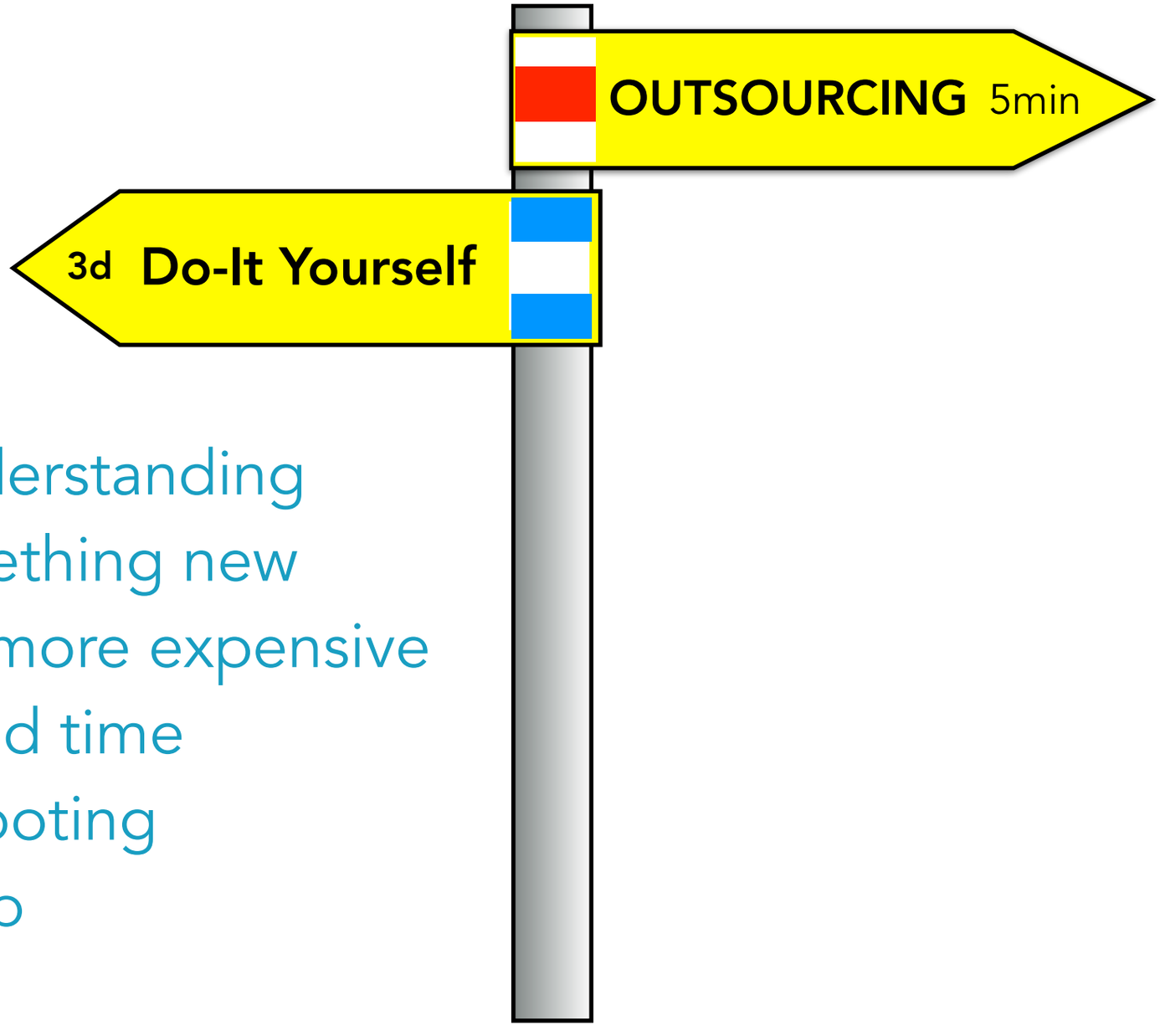
Mapping

Reference

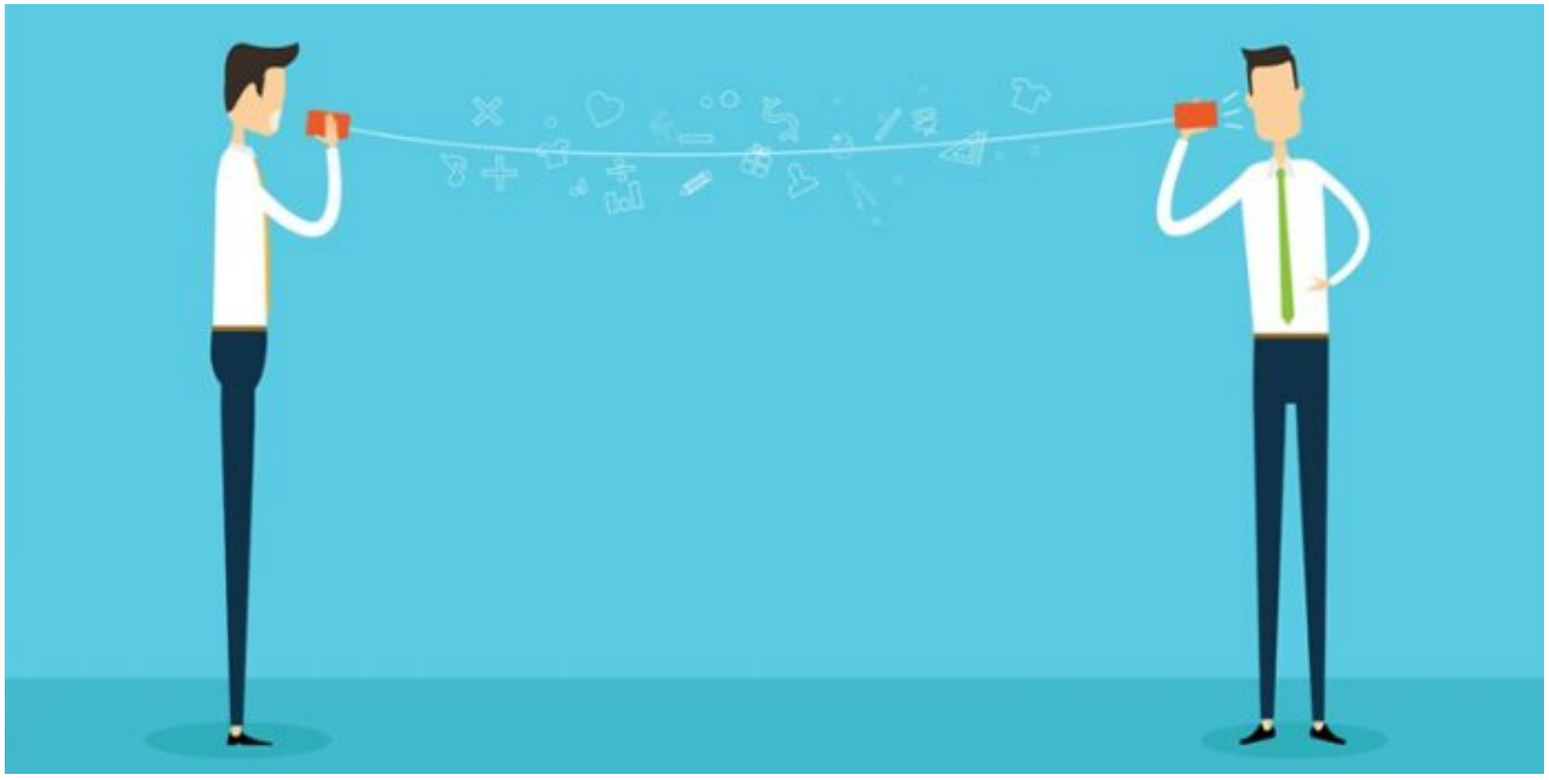
Clustering

Scaffolding / Gap-closing

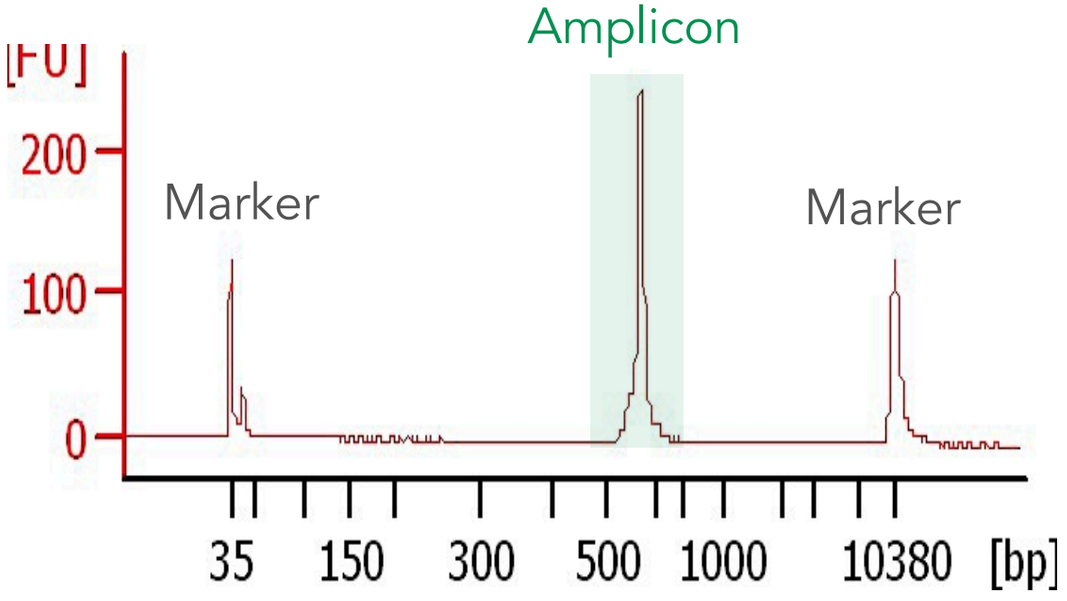
Data Analysis

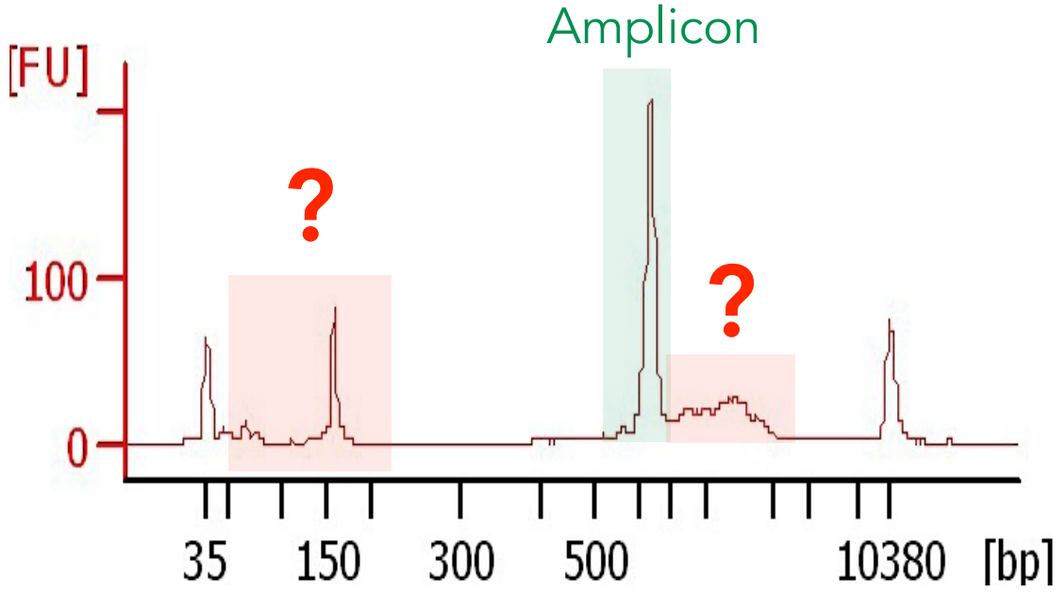


- better understanding
- learn something new
- often not more expensive
- turn-around time
- troubleshooting
- data mixup



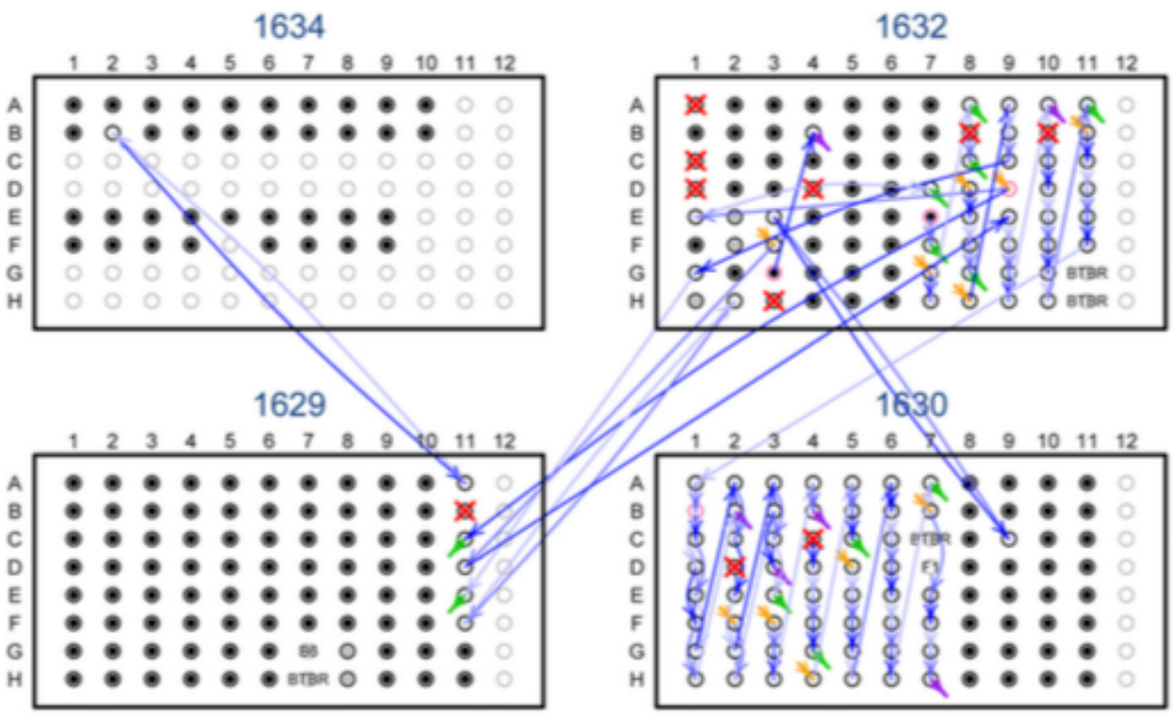
Example: Fragment Length Analysis





Identification and Correction of Sample Mix-Ups in Expression Genetic Data: A Case Study

Karl W. Broman,*,[2] Mark P. Keller,† Aimee Teo Broman,* Christina Kendziorski,* Brian S. Yandell,‡,§ Śaunak Sen,**,[1] and Alan D. Attie†
*Department of Biostatistics and Medical Informatics, †Department of Biochemistry, ‡Department of Statistics, and §Department of Horticulture, University of Wisconsin, Madison, Wisconsin 53706, and **Department of Epidemiology and Biostatistics, University of California, San Francisco, California 94107



"To map the genetic loci influencing a complex phenotype, one seeks to establish an **association between genotype and phenotype**. In such an effort, the maintenance of the concordance between genotyped and phenotyped samples and data is critical. **Sample mislabeling and other sample mix-ups will weaken associations**, resulting in reduced power and biased estimates of locus effects."

Sequencing Technologies

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1 Sanger (chain termination)

2 Roche 454 Pyrosequencing (pyrophosphate)

Ion Torrent (semiconductor technology)

Illumina (fluorescent - sequencing by synthesis (SBS))

Singular Genomics (fluorescent - rapid SBS)

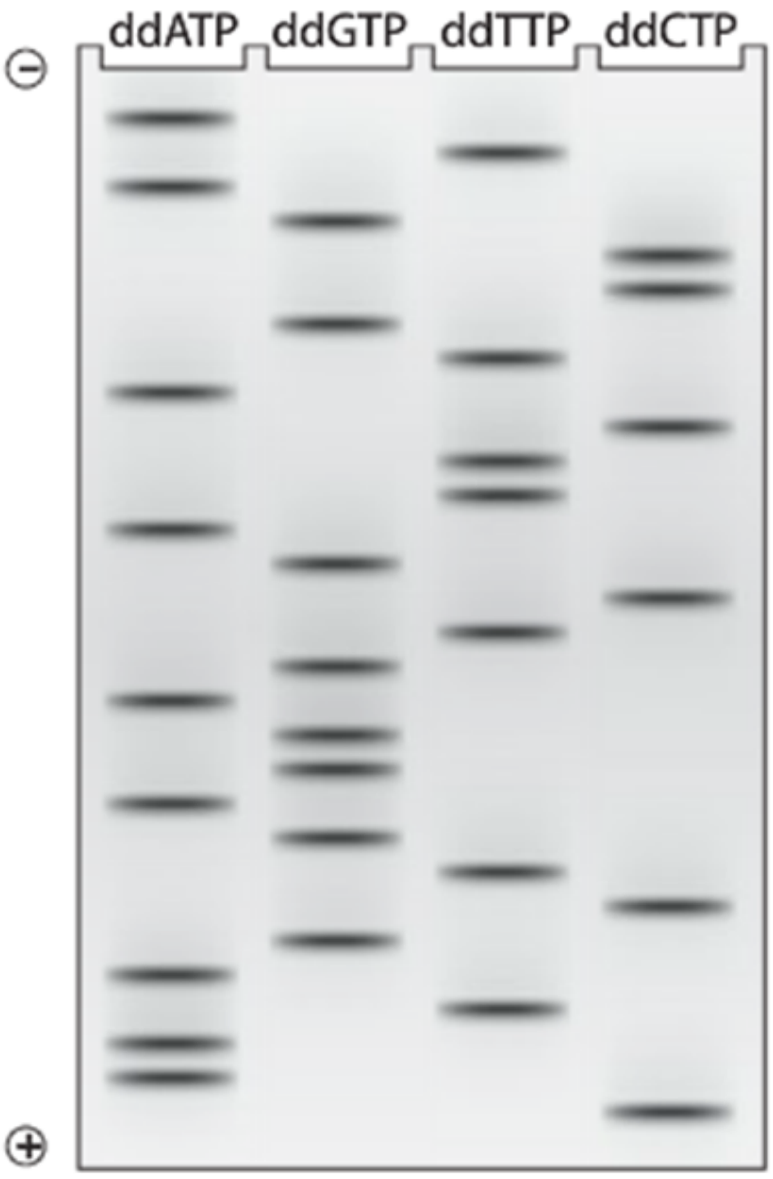
Aviti (fluorescent - sequencing by avidity)

3 **PacBio** (fluorophore)

Nanopore (ionic current)

Helicos - SeqLL (fluorescent)

4 Bionano - Saphyr (third-generation optical mapping)



The Nobel Prize in Chemistry 1980

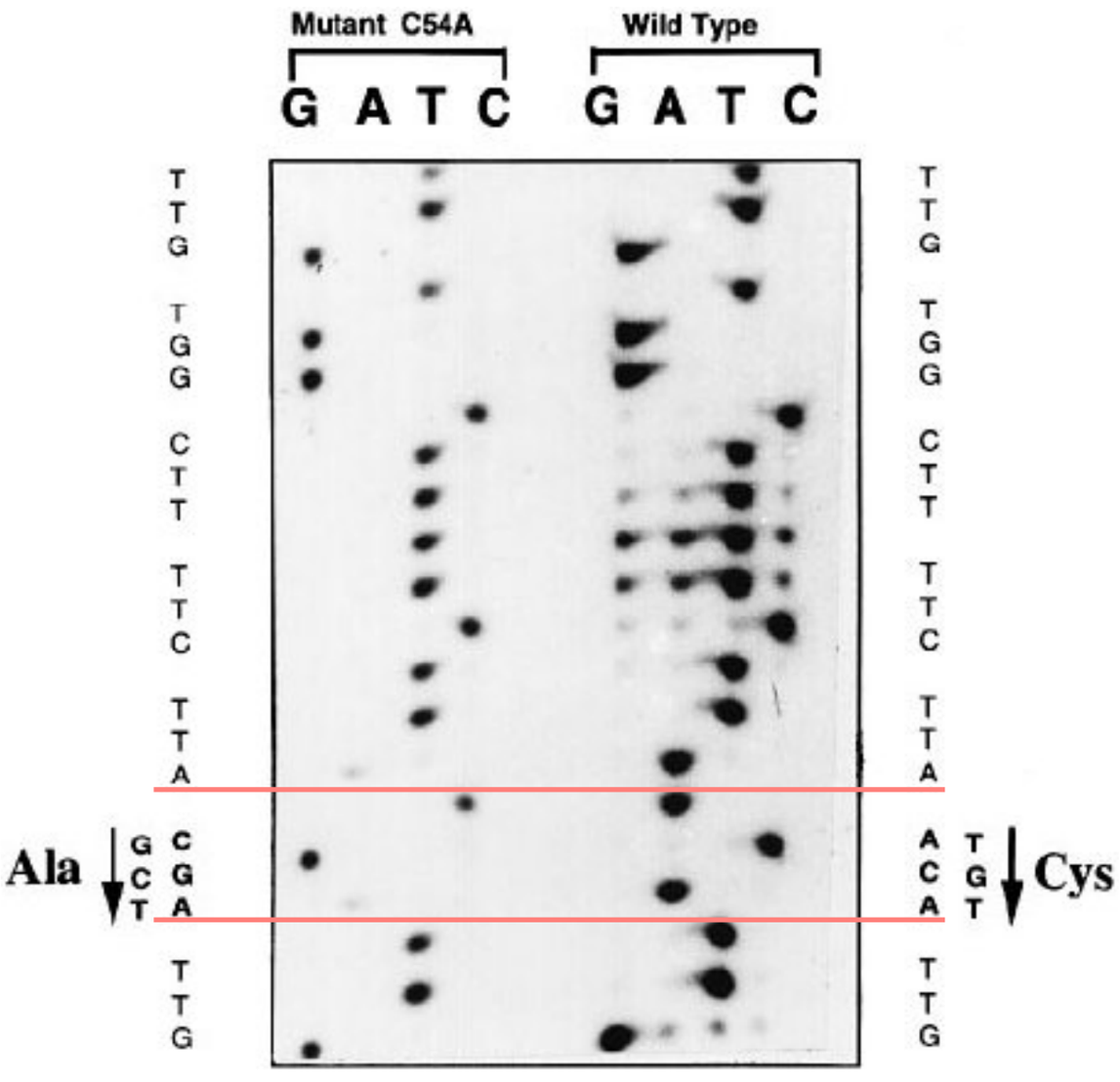
Paul Berg, Walter Gilbert, Frederick Sanger

Paul Berg

Walter Gilbert

Frederick Sanger

The Nobel Prize in Chemistry 1980 was divided, one half awarded to Paul Berg *"for his fundamental studies of the biochemistry of nucleic acids, with particular regard to recombinant-DNA"*, the other half jointly to Walter Gilbert and Frederick Sanger *"for their contributions concerning the determination of base sequences in nucleic acids"*.



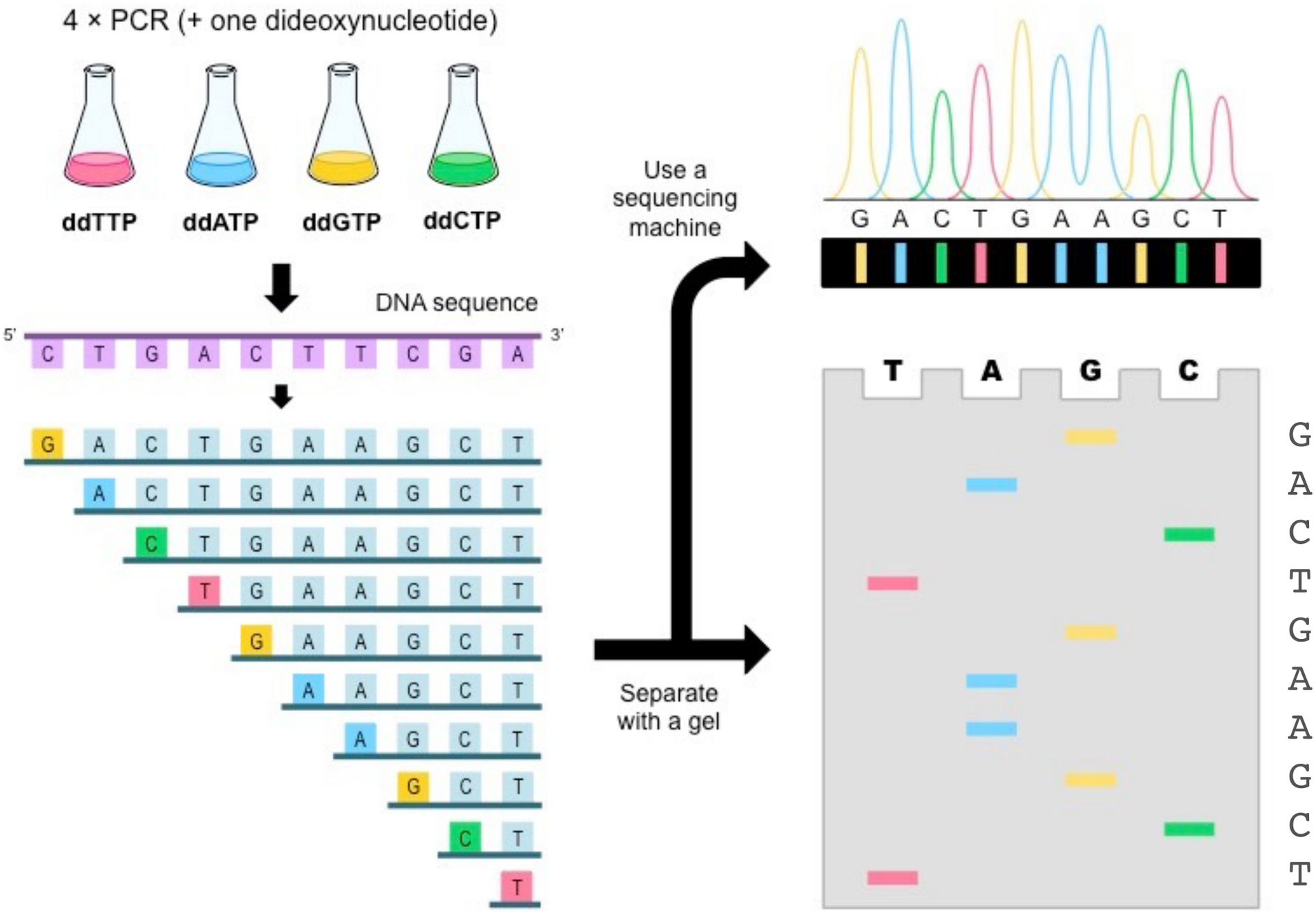
Protein engineering of BamHI restriction endonuclease: replacement of Cys54 by Ala enhances catalytic activity

November 1998 · Protein Engineering 11(10):931-5

DOI: 10.1093/protein/11.10.931

Source · PubMed

Autoradiogram of DNA sequencing gel covering the mutation region. The mutation was confirmed by full sequencing of the gene.


4 × PCR (+ one dideoxynucleotide)
ddTTP
ddATP
ddGTP
ddCTP
DNA sequence
5'
3'
C T G A C T T C G A
G A C T G A A G C T
A C T G A A G C T
C T G A A G C T
T G A A G C T
G A A G C T
A A G C T
A G C T
G C T
C T
T
Use a sequencing machine
Separate with a gel
G A C T G A A G C T
T A G C
G A C T G A A G C T

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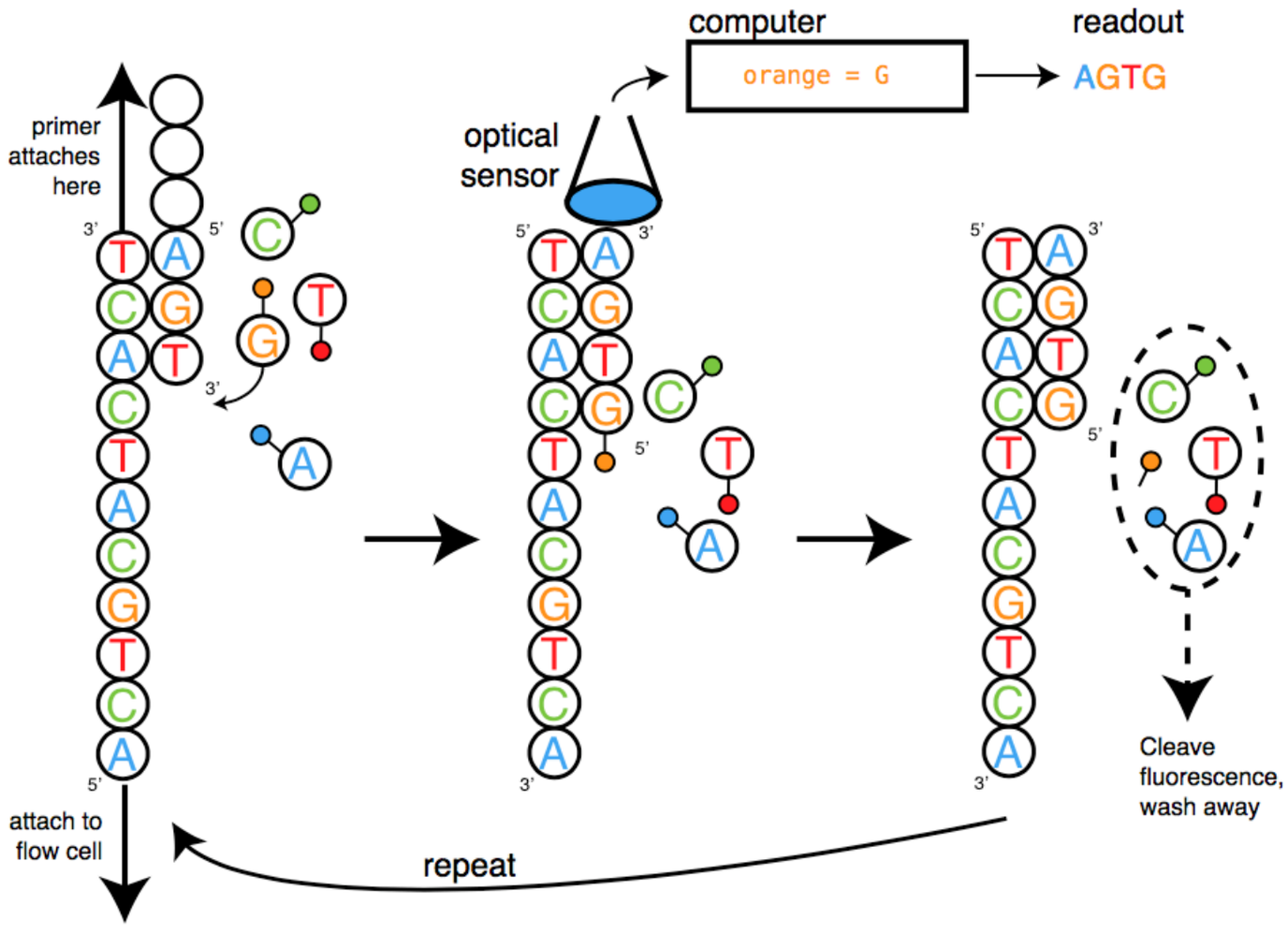
Sequencing by Synthesis (fluorescent)



Sequencing by Synthesis. dNTP fluorescence is translated to a base call.

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Illumina Systems

MiniSeq System

1.8-7.5 Gb
8-25 million
2 x 150 bp
50

MiSeq Series

0.3-15 Gb
1-25 million
2 x 300 bp
384

NextSeq Series

20-120 Gb
130-400 million
2 x 150 bp
96

HiSeq Series

125-1500 Gb
2.5-5 billion
2 x 150 bp
12

HiSeq X Series

900-1800 Gb
3-6 billion
2 x 150 bp
16

NovaSeq Series

134-6000 Gb
Up to 20 billion
2 x 150 bp
48

http://www.illumina.com

MGI	 Sequencers +	 Sequencers +	 Sequencers +	 Sequencers +
Product Model	DNBSEQ-T7	DNBSEQ-G400	DNBSEQ-G400* For HotMPS Only	DNBSEQ-G50
Features	Ultra-high Throughput	Adaptive	Adaptive	Effective
Applications	Whole Genome Sequencing, Deep Exome Sequencing, Transcriptome Sequencing, and Targeted Panel Projects.	WGS, WES, Transcriptome sequencing, etc.	WGS, WES, Transcriptome sequencing, etc.	Small whole genome sequencing, targeted DNA/RNA panels, low-pass whole genome sequencing
Flow Cell Type	FC	FCL & FCS	FCL	FCL & FCS
Lane/Flow Cell++	1 lane	2 or 4 lanes	4 lanes	1 lane
Operation Mode	Ultra-high Throughput	High Throughput	High Throughput	Medium Throughput
Max. Throughput / RUN	6TB	1440GB	720GB	150GB
Effective Reads / Flow Cell	5000M	1500-1800M	1500-1800M	500M / 100M
Average run time	24~30 hours for PE150 sequencing	FCS: 13~37 hours FCL: 14~109 hours	15.5-50.5 hours	9~40 hours
Min. Read Length	PE100	SE50	SE50	SE50
Max. Read Length	PE150	SE400/PE200	PE100	PE150

MGI Tech is the manufacturing sister of China's largest genome sequencing company, BGI Genomics.

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Singular Genomics (fluorescent - rapid SBS)

G4

A highly versatile benchtop sequencer targeting applications where accuracy, speed, flexibility, and scale matter most

Key milestones

- Launched 2021
- First units expected ship in Q2 2022

PX

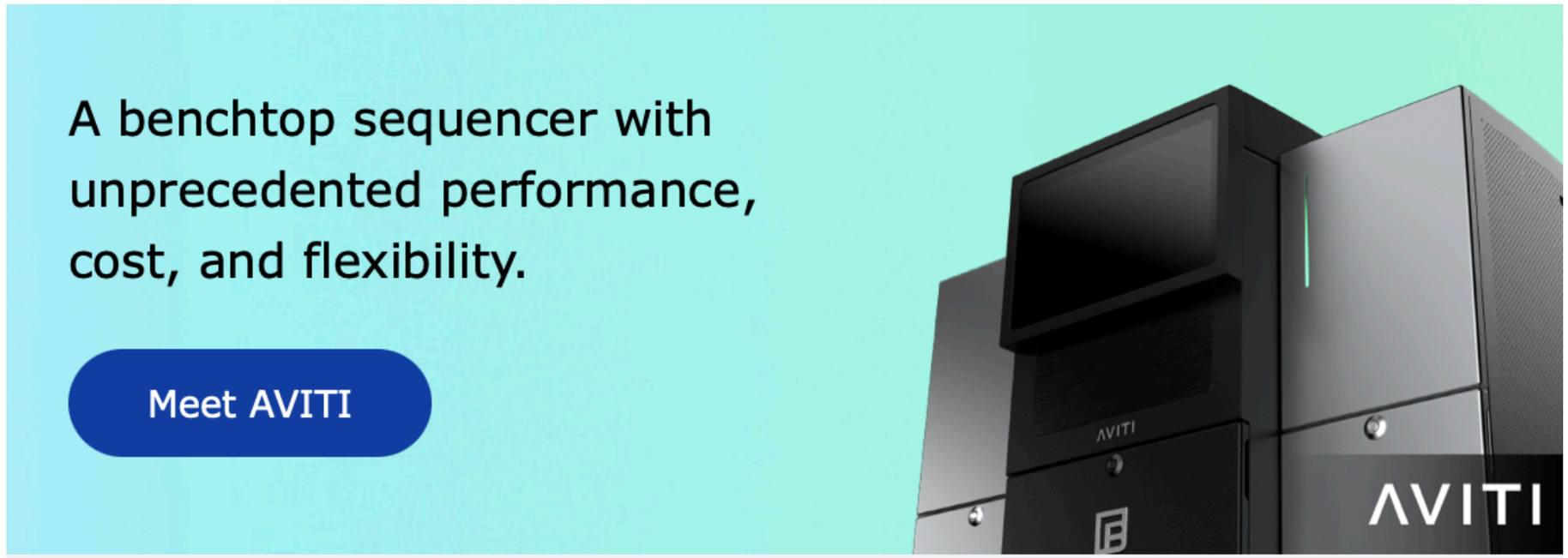
Integrated in situ platform for multiomic analysis in single cells and tissues

PX image is for illustrative purposes only

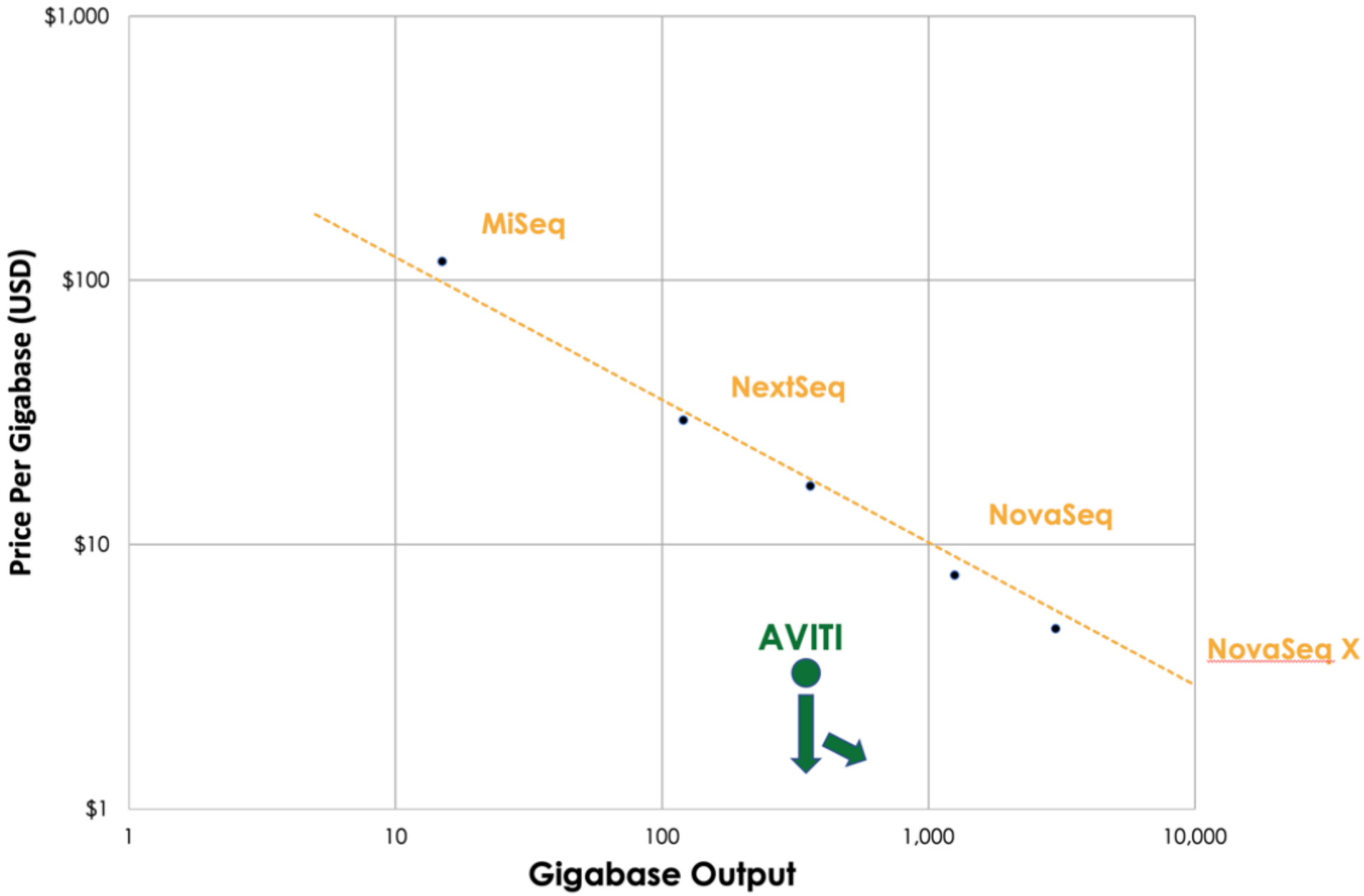
Key milestones

- Early access program to begin 2022
- Commercial launch expected in 2023

Element Biosciences (fluorescent - sequencing by avidity)



Performance	Flexibility	Cost
%Q30 > 90 at 2x150	Dual flow cells	$289K/instrument
1B reads/flow cell*	Flexible start	$1680/300 cycles $1080/150 cycles
600 Gb+ output/run	Tunable read throughput	Leasing/financing options available


$1,000
$100
$10
$1
Price Per Gigabase (USD)
1
10
100
1,000
10,000
Gigabase Output
MiSeq
NextSeq
NovaSeq
NovaSeq X
AVITI

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Pacific Biosciences

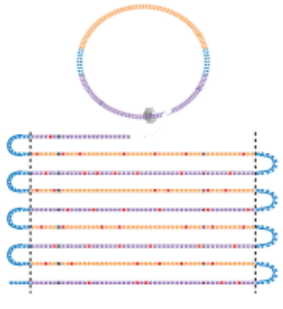
Over a decade of on market technological innovation

RS I /RS II
75k/150k ZMW

2011-2013: PacBio launches groundbreaking single-molecule sequencing platform; named top innovation by *The Scientist* magazine

Sequel
1M ZMW

2015: PacBio launches the Sequel system



HiFi

2018: HiFi enables ultra-high accuracy at long read lengths and establishes PacBio as leader in sequencing accuracy

Sequel II/IIe
8M ZMW

2019 | 2020: PacBio launches Sequel II and Sequel IIe

Revio
25M x4 ZMW

2023: PacBio expected to launch Revio enabling the sub-$1k long-read genome at scale

Increased throughput **>10,000-fold**

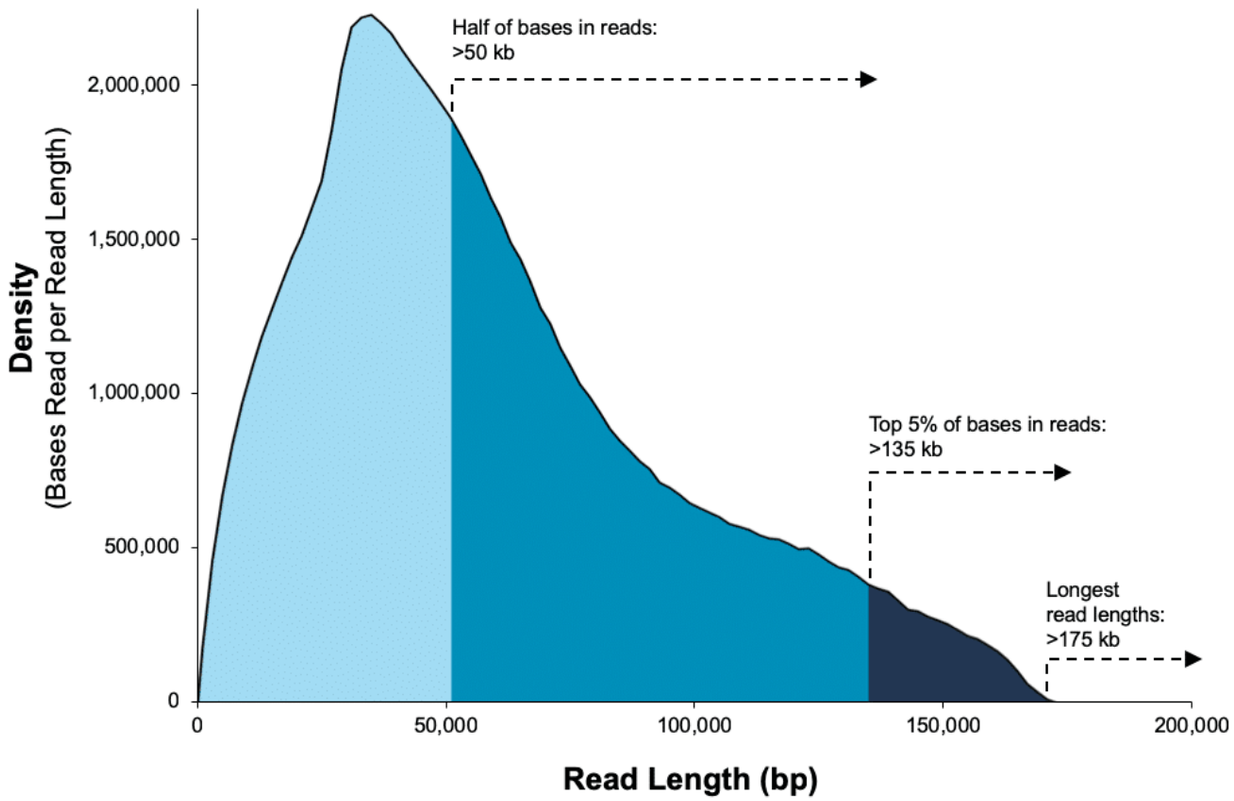
Increased read length **>100-fold**

PACBIO®

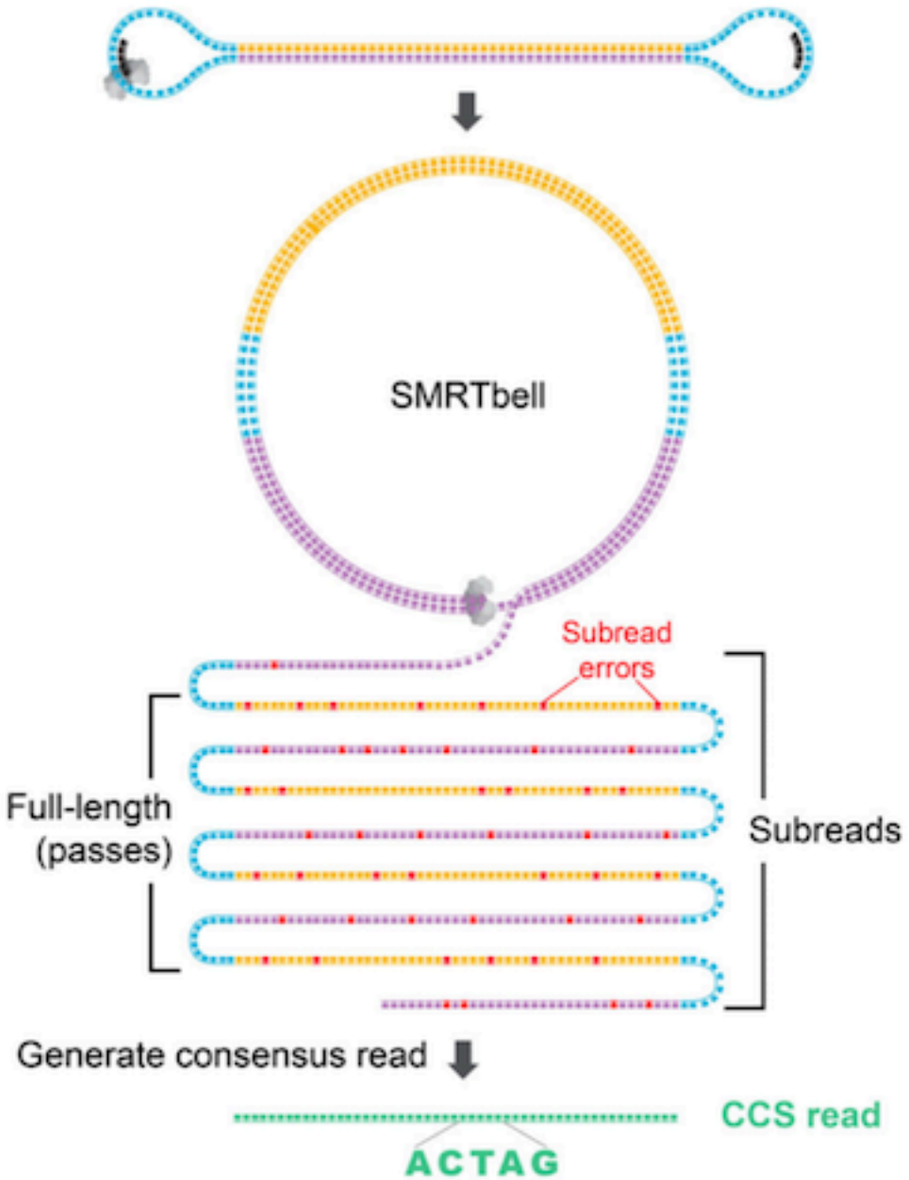
(Pacific Biosciences)

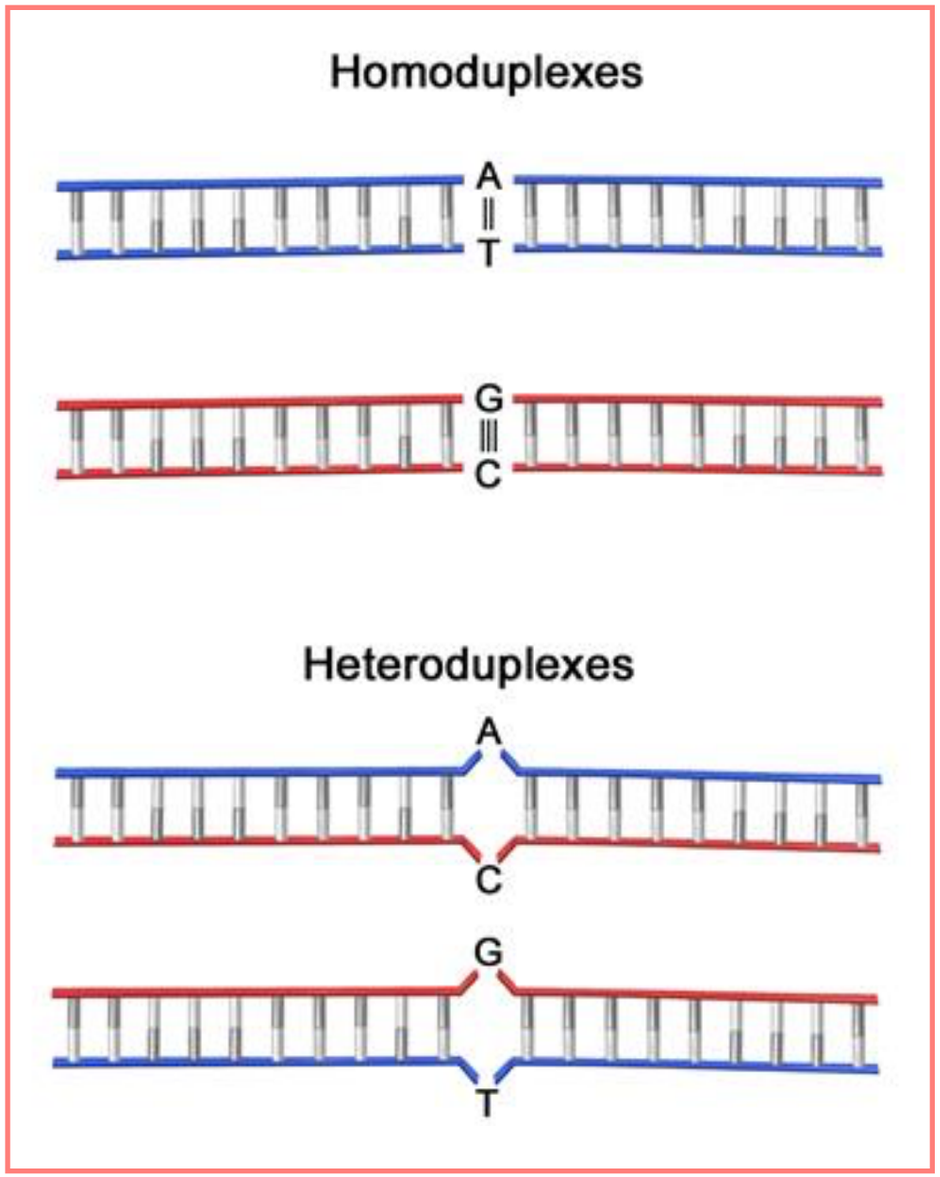
https://www.pacb.com

Sequel



Data from a 35 kb size-selected *E. coli* library using the SMRTbell Express Template Prep Kit 2.0 on a Sequel II System (1.0 Chemistry, Sequel II System Software v7.0, 15-hour movie)*.


SMRTbell
Subread errors
Full-length (passes)
Subreads
Generate consensus read
CCS read
ACTAG


Homoduplexes
A
T
G
C
Heteroduplexes
A
C
G
T

Oxford NANOPORE Technologies

SmidgION | Flongle | MinION | GridION | PromethION

https://www.nanoporetech.com


The Economist
Open-source medical devices
Dealing with nuclear waste
Burt Rutan, a maverick in flight
Technology Quarterly
December 1st 2012
The dream of the medical tricorder
the hand held devices inspiring medical add-ons for smartphones


Running
Stop Sequencing

MinION Mk1C

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Bionano

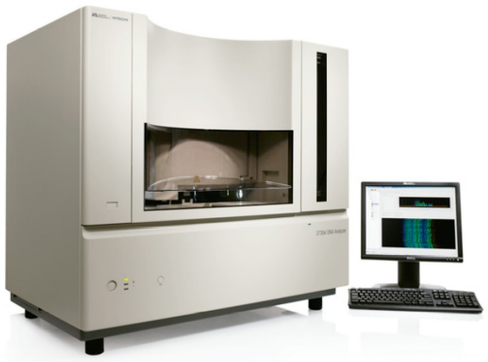
(optical mapping)

Optical mapping is a technique for constructing ordered, genome-wide, high-resolution restriction maps from single, stained molecules of DNA, called "optical maps".

Optical Sequencing

Optical sequencing is a single molecule DNA sequencing technique that follows sequence-by-synthesis and uses optical mapping technology. During synthesis, fluorochrome-labeled nucleotides are incorporated through the use of DNA polymerases and tracked by fluorescence microscopy.


PromethION




AVITI


NovaSeq™ 6000


DNBSEQ-T7

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What is the best NGS platform?

The best platform for a particular application depends on several factors, such as project goals, sample type, sequencing depth, budget and bioinformatics support. Here are some common NGS platforms:

1. Illumina (e.g., NovaSeq, NextSeq, MiSeq): Illumina is currently the most widely used NGS platform. HiSeq instruments offer high-throughput sequencing, making them suitable for large-scale projects. MiSeq is a smaller benchtop sequencer that is more cost-effective for smaller projects or labs with lower sequencing needs.

2. Element Biosciences (Aviti System). Aviti is an advanced DNA sequencing platform. It is designed to provide high-quality, accurate, and cost-effective sequencing for various genomic applications.

3. BGI Genomics (MGISEQ/T7, DNBSEQ): BGI Genomics is a cost-effective and therefore attractive option for large-scale projects.

4. Pacific Biosciences (PacBio Sequel II): PacBio uses Single Molecule Real-Time (SMRT) sequencing and is advantageous for long read sequencing. It enables the sequencing of longer DNA fragments, facilitating the assembly of complex genomes and the detection of structural variation.

5. Oxford Nanopore Technologies (MinION, GridION, PromethION): ONT sequencing uses nanopore-based technology to provide long reads and real-time data analysis. It is portable and has been used for field applications such as rapid pathogen identification and monitoring.

It is important to evaluate the specific requirements of your project and consider factors such as read length, sequencing depth, accuracy, cost and data analysis needs when determining the best NGS platform to use. As the technology is advancing rapidly, it is advisable to consult the latest information and experts in the field to make an informed decision based on the most up-to-date information available.

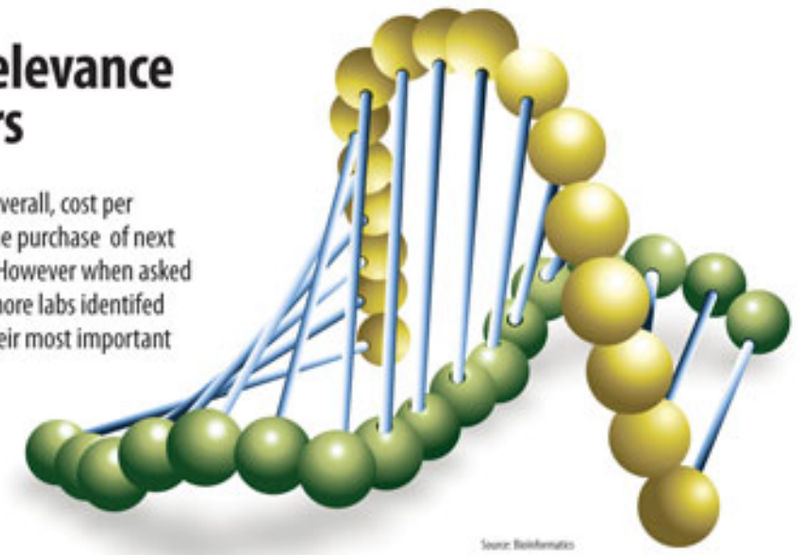
The all-in-one MPS platform does not exist (yet)!

- Research question
- Budget (including storage and analysis)
- Read / sequence length
- Number of reads / coverage
- Possible contaminants
- Quality and quantity of template
- Number of samples
- Availability

GEN Genetic Engineering & Biotechnology News

NGS: Cost and Relevance Are Key to Buyers

Recent market research shows that, overall, cost per base was the most cited concern in the purchase of next generation sequencing instruments. However when asked to identify their top three concerns, more labs identifed "Appropriate to My Application" as their most important criteria.



The 10 Most Critical Platform Attributes as Defined by Purchasers

1. Cost per base	43%	6. Read length	24%
2. Sequencing data quality	34%	7. Instrument cost	18%
3. Appropriate for my application	32%	8. Number of reads	17%
4. Reproducibility/accuracy	31%	9. Available software analysis tools	16%
5. Amount of DNA/RNA needed per experiment	25%	10. Instrument reliability	16%

Sequencing Data

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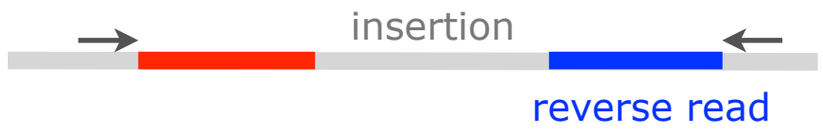
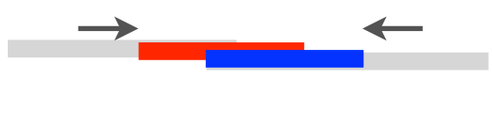
Sequence Read Data

Single Reads (SR)



Paired-End (PE) Reads



Overlapping Paired-End (PE) Reads

Single Reads (SR) with Index

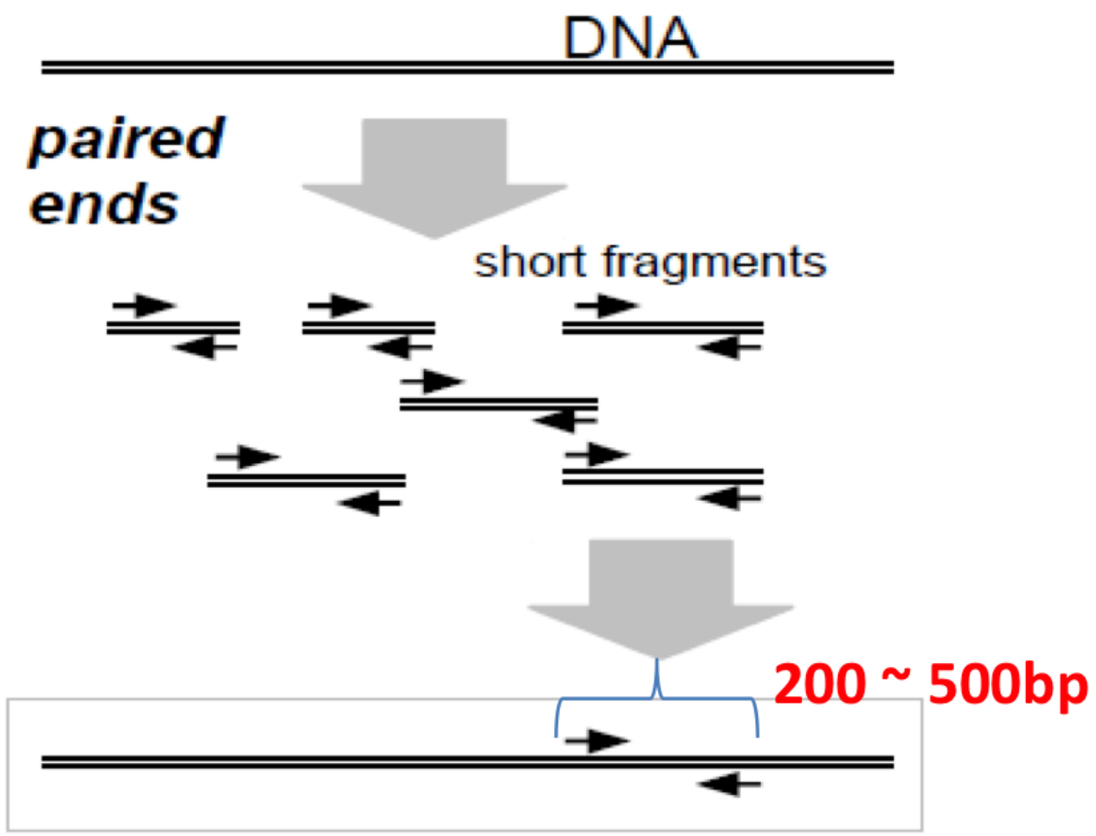
Paired-End (PE) Reads with Index

Paired-End (PE) Reads with Dual Indexing

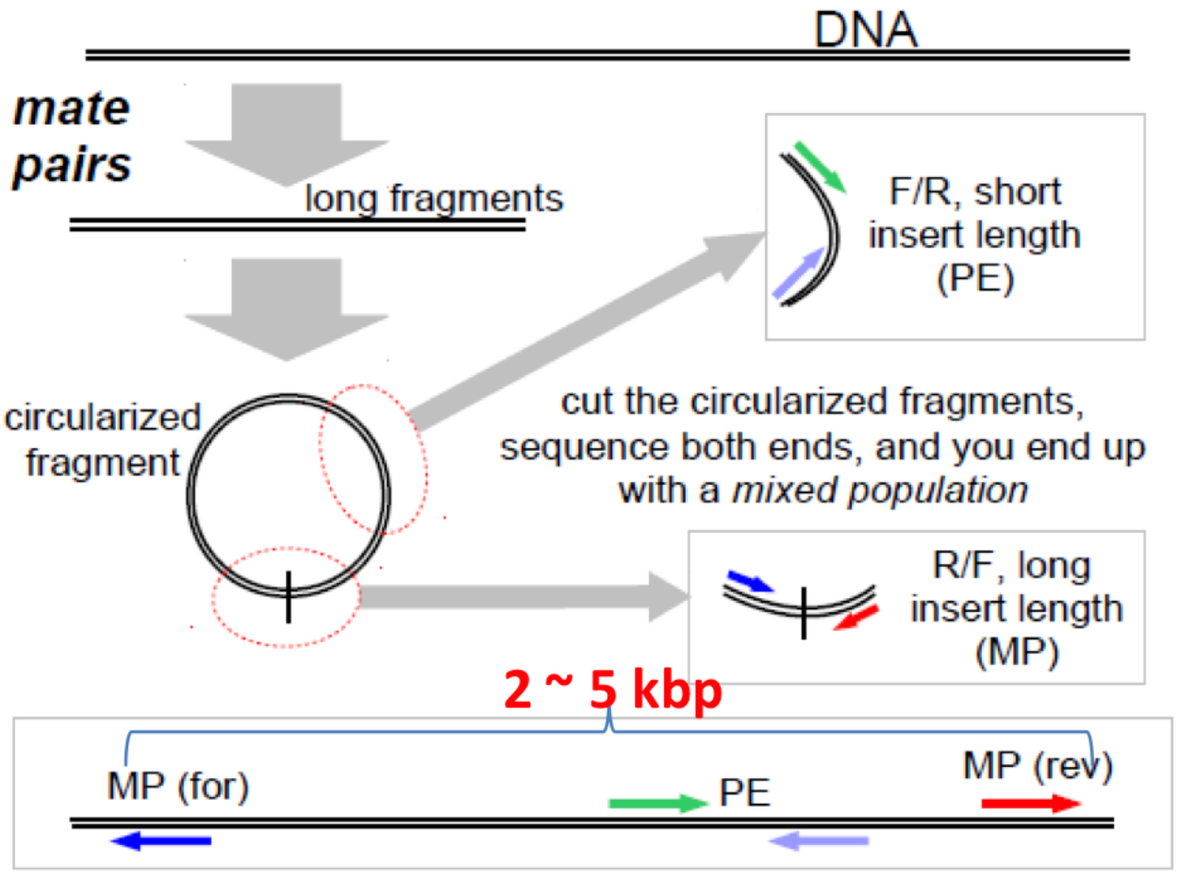
Extended Single Reads (SR) with Index

Sequence Read Data

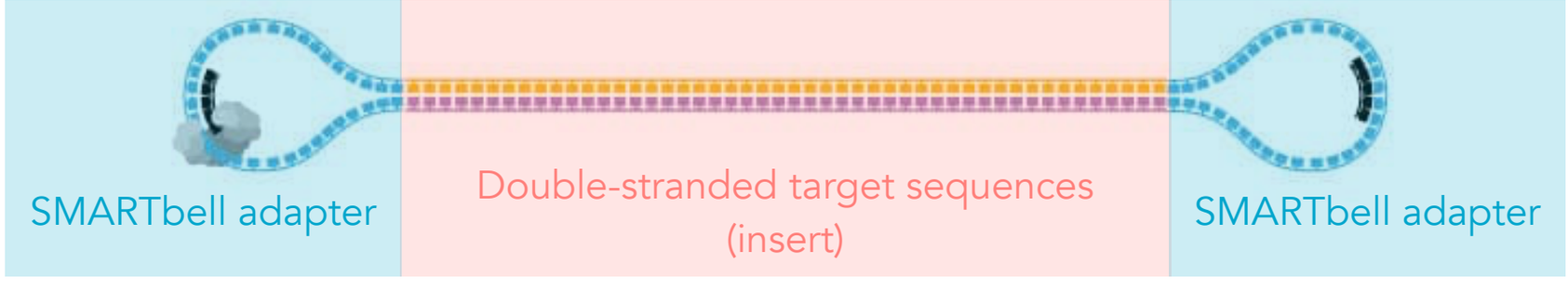
paired-end (PE)

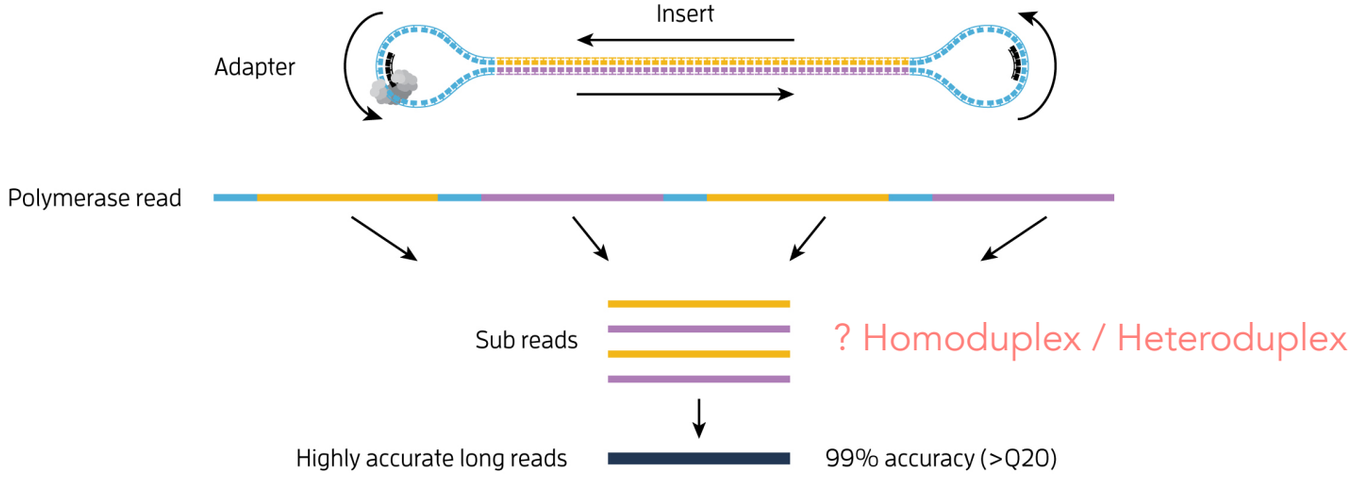


mate-pair (MP)

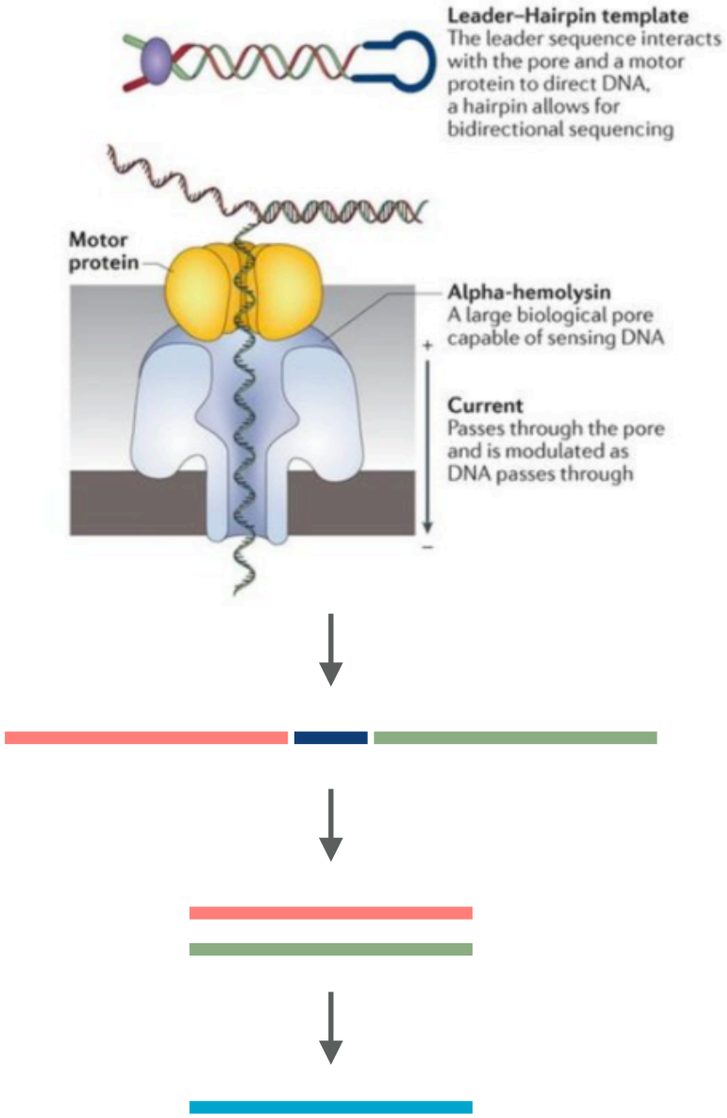


PacBio SMRTbell Library





ONT Sequencing



Data Submission

The European Nucleotide Archive (ENA) captures and presents information relating to experimental workflows that are based around nucleotide sequencing. A typical workflow includes the isolation and preparation of material for sequencing, a run of a sequencing machine in which sequencing data are produced and a subsequent bioinformatic analysis pipeline. ENA records this information in a data model that covers input information (sample, experimental setup, machine configuration), output machine data (sequence traces, reads and quality scores) and interpreted information (assembly, mapping, functional annotation).

Sequence Read Archive (SRA) makes biological sequence data available to the research community to enhance reproducibility and allow for new discoveries by comparing data sets. The SRA stores raw sequencing data and alignment information from high-throughput sequencing platforms, including Roche 454 GS System®, Illumina Genome Analyzer®, Applied Biosystems SOLiD System®, Helicos Heliscope®, Complete Genomics®, and Pacific Biosciences SMRT®.

Data Deposition Example from the Literature:

Mushegian *et al*. (2018) **Environmental sources of bacteria and genetic variation in behavior influence host-associated microbiota**. AEM doi:10.1128/AEM.01547-18.

Sequence data are deposited in the **European Nucleotide Archive of the EBI under accession number PRJEB30308** (http://www.ebi.ac.uk/ena/data/view/PRJEB30308). Data tables, OTUs sequences and code used for analysis can be found on **Github** at https://github.com/amusheg/Daphnia-microbiota-behavior and will be deposited in **Dryad** upon publication.

EMBL-EBI Services Research Training About us

ENA European Nucleotide Archive

Search

Examples: BN000065, histone

Advanced Sequence

Home | Search & Browse | Submit & Update | Software | About ENA | Support

Contact Helpdesk

Study: PRJEB30308

Microbiota associated with Daphnia exhibiting genetic variation in behavior

View: Project XML Study XML **Download:** Project XML Study XML

Name
Microbiota of browsing Daphnia

Submitting Centre
Universitaet Basel

Secondary accession(s)
ERP112744

Description
In many organisms, host-associated microbial communities are acquired horizontally after birth. This process is believed to be shaped by a combination of environmental and host genetic factors. We examined whether genetic variation in animal behavior could affect the composition of the animal's microbiota in different environments. The freshwater crustacean Daphnia magna is primarily planktonic, but exhibits variation in the degree to which it browses in benthic sediments. We performed an experiment with clonal lines of D. magna showing different levels of sediment-browsing intensity exposed to either bacteria-rich or bacteria-poor sediment or whose access to sediments was prevented. We find that the bacterial composition of the environment and genotype-specific browsing intensity together influence the composition of the Daphnia-associated bacterial community. Exposure to more diverse bacteria did not lead to a more diverse microbiome, but greater abundances of environment-specific bacteria were found associated with host genotypes that exhibited greater browsing behavior. Our results indicate that, although there is a great deal of variation between individuals, behavior can mediate genotype-by-environment interaction effects on microbiome composition.

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Download: 1 - 512 of 512 results in TEXT

Select columns

Showing results 1 - 10 of 512 results

Study accession	Sample accession	Secondary sample accession	Experiment accession	Run accession	Tax ID	Scientific name	Instrument model	Library layout	FASTQ files (FTP)	FASTQ files (Galaxy)	Submitted files (FTP)	Submitted files (Galaxy)	NCBI SRA file (FTP)	NCBI SRA file (Galaxy)	CRAM Index files (FTP)	CRAM Index files (Galaxy)
PRJEB30308	SAMEA5166093	ERS2973813	ERX2993334	ERR2990925	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166094	ERS2973814	ERX2993335	ERR2990926	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166095	ERS2973815	ERX2993336	ERR2990927	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166096	ERS2973816	ERX2993337	ERR2990928	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		

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NCBI Resources How To Sign in to NCBI

BioProject

BioProject PRJEB30308 Search

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Microbiota of browsing Daphnia

Accession: PRJEB30308 ID: 516850

Microbiota associated with Daphnia exhibiting genetic variation in behavior

In many organisms, host-associated microbial communities are acquired horizontally after birth. More...

Accession	PRJEB30308
Scope	Monoisolate
Submission	Registration date: 24-Jan-2019 **Universitaet Basel**

Project Data:

Resource Name	Number of Links
SEQUENCE DATA	
SRA Experiments	512
OTHER DATASETS	
BioSample	512

▾ SRA Data Details

Parameter	Value
Data volume, Gbases	22
Data volume, Mbytes	14805

Related information

BioSample

SRA

Recent activity Turn Off Clear

PRJEB30308 (1)

BioProject
Microbiota of browsing Daphnia

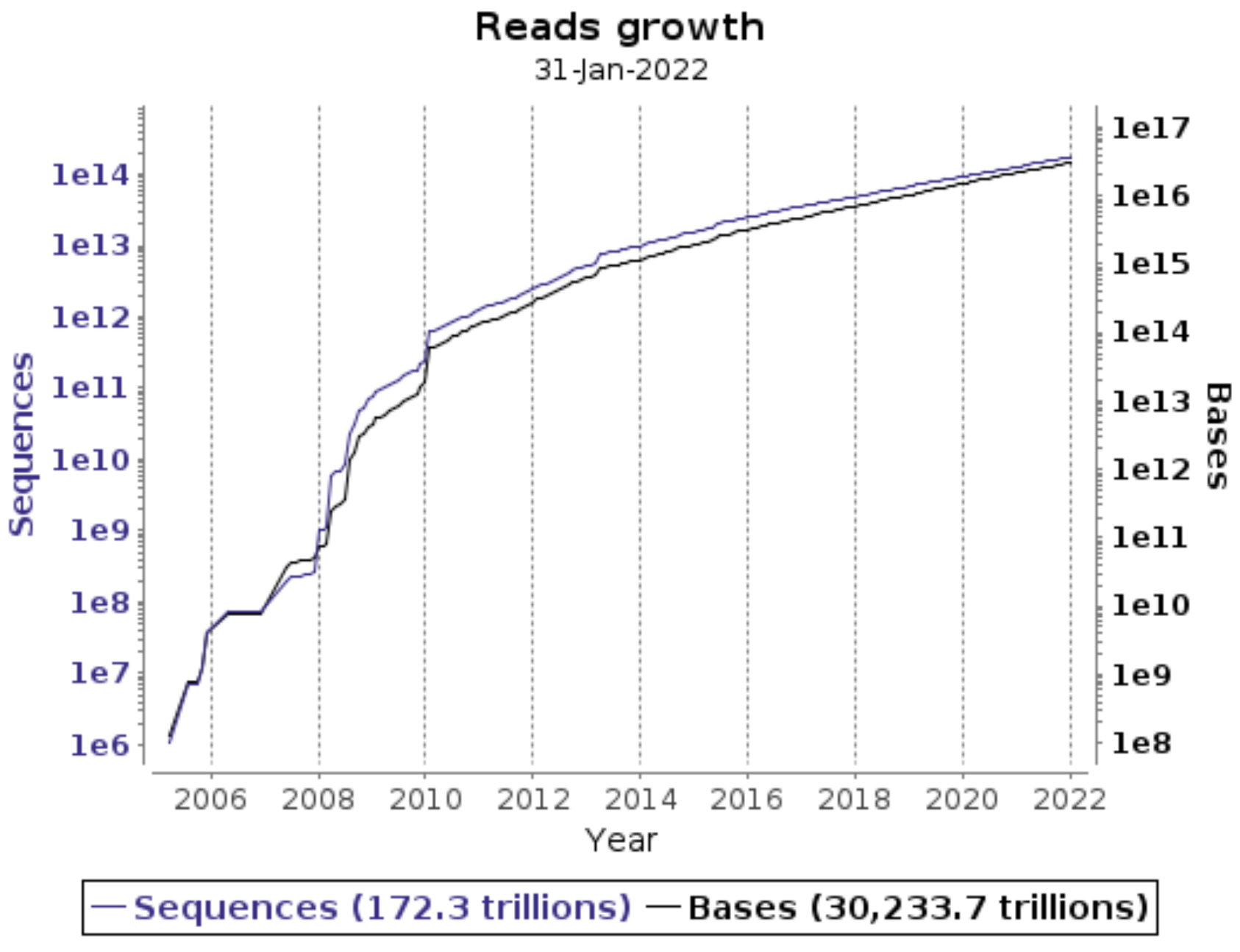
BioProject
The European Nucleotide Archive in 2017

A Benchmark Study on Error Assessment and Quality Control of CCS Reads Derived

Testing the potential of a ribosomal 16S marker for DNA metabarcoding of insects

See more...

ENA European Nucleotide Archive



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1

Choose the MPS technology according to your needs.

2

Keep your raw data safe and submit it as early as possible.

3

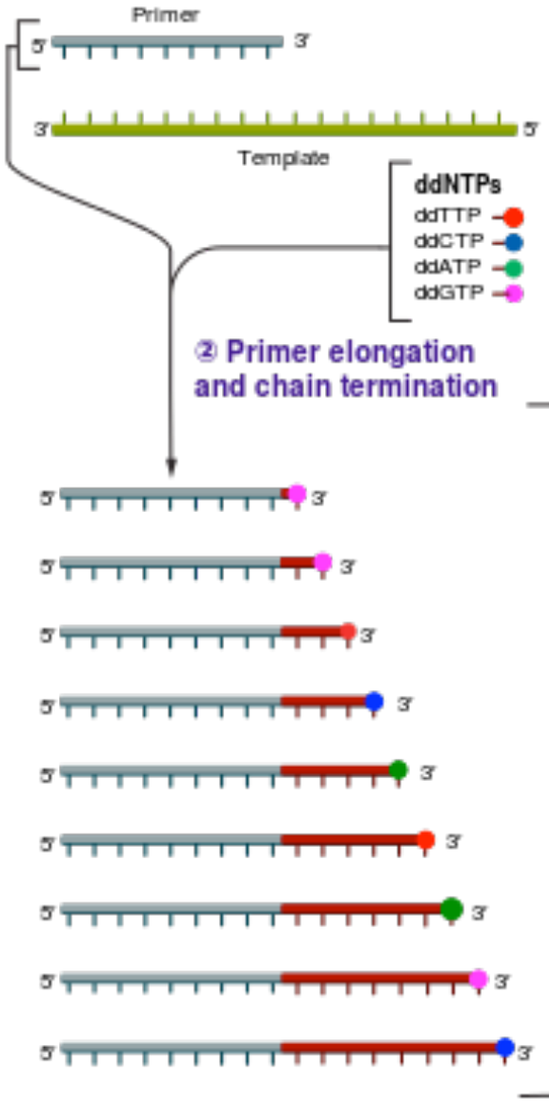
Keep your sequence files zipped.

EXTRAS

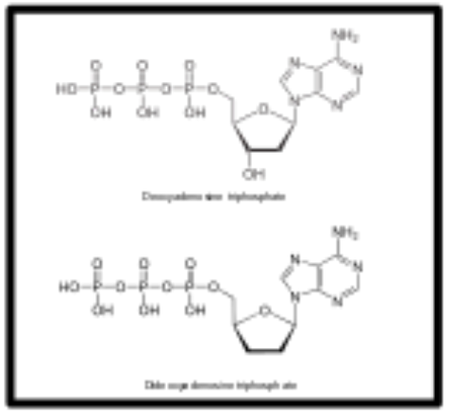
Sequencing Technologies Extended

Capillary sequencing

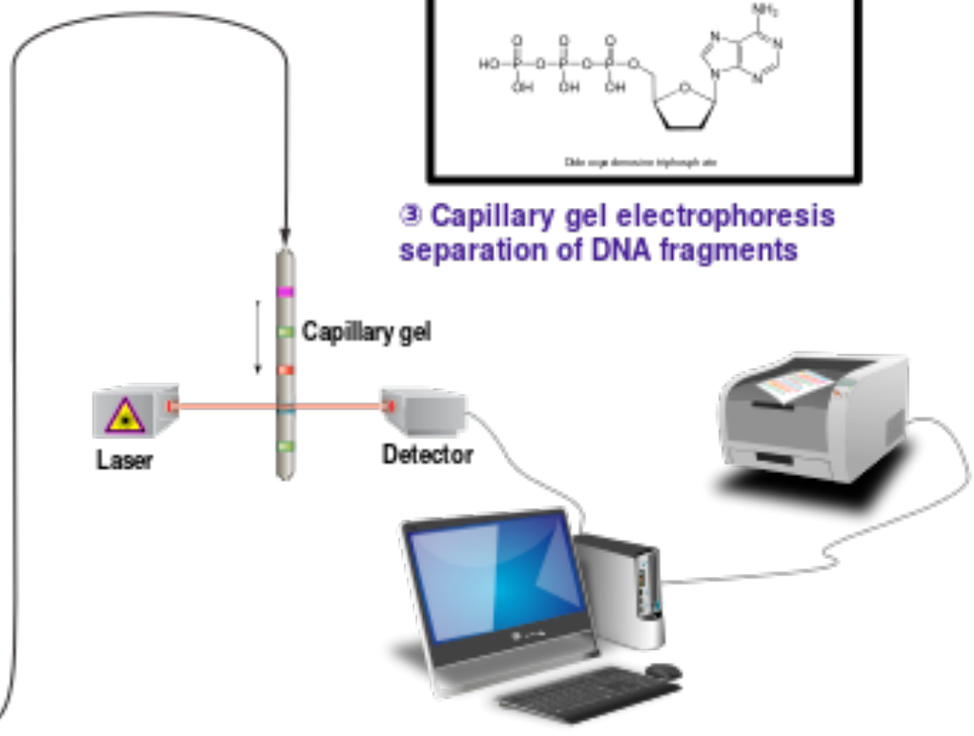
① Reaction mixture
- Primer and DNA template
- DNA polymerase
- ddNTPs with flourochromes
- dNTPs (dATP, dCTP, dGTP, and dTTP)

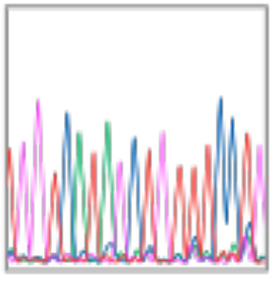


② Primer elongation and chain termination



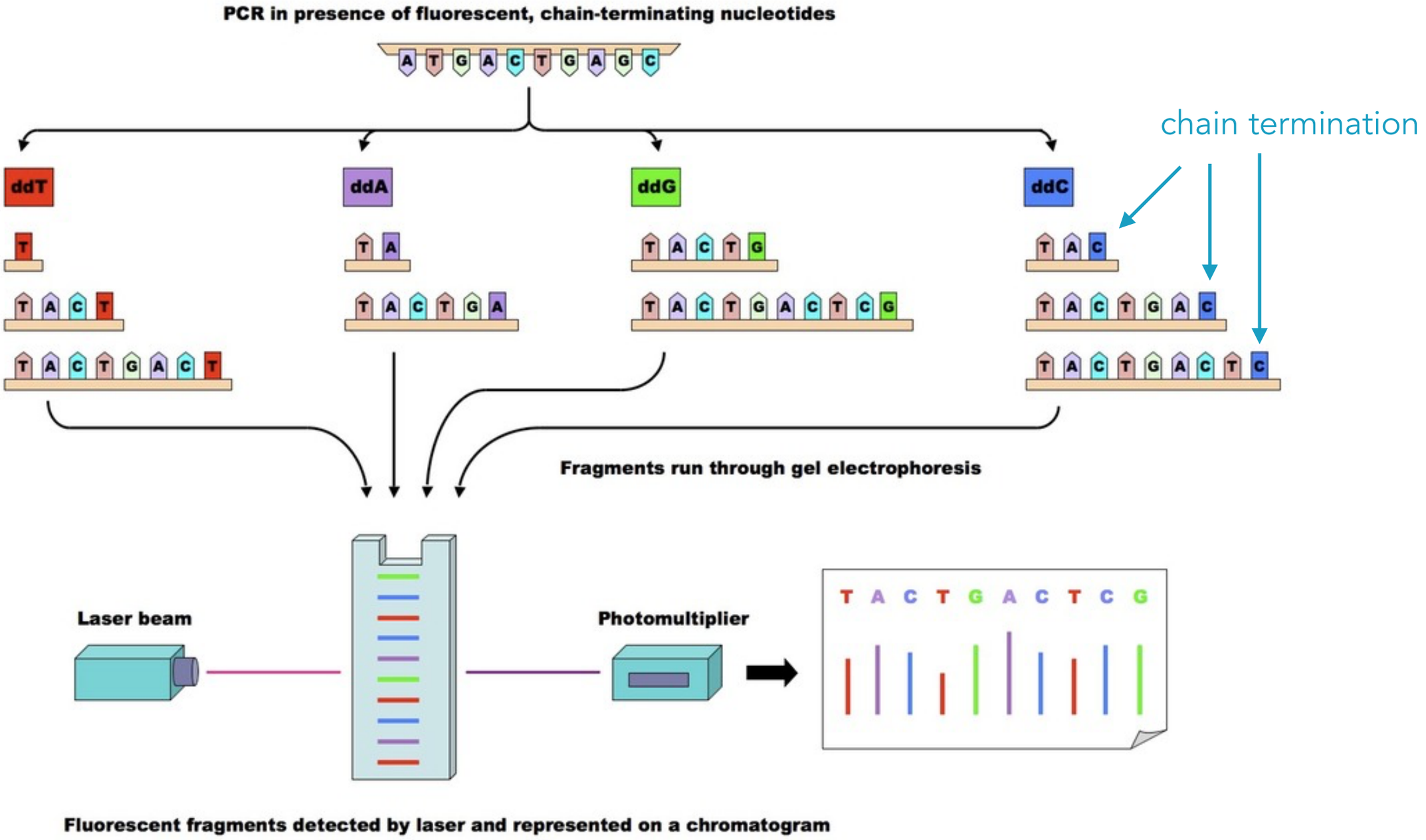
③ Capillary gel electrophoresis separation of DNA fragments





Chromatograph

④ Laser detection of flourochromes and computational sequence analysis



Fluorescent fragments detected by laser and represented on a chromatogram

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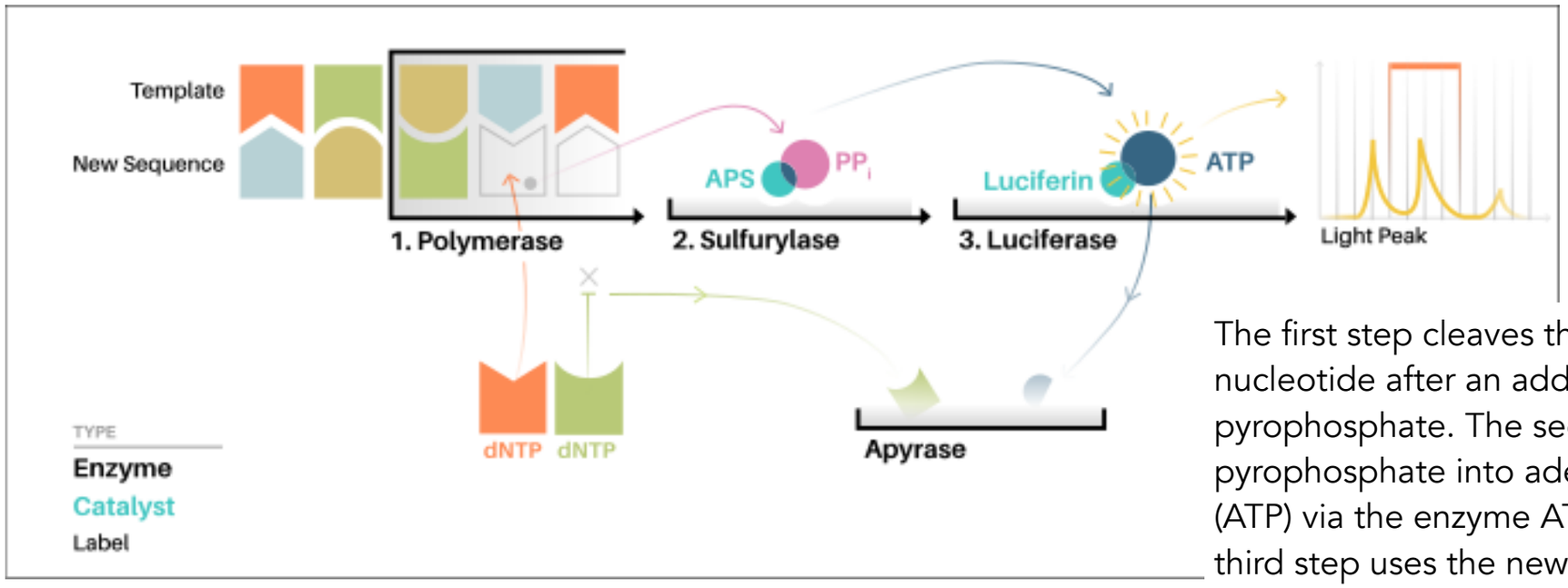
Pyrosequencing

GS Junior

Roche 454

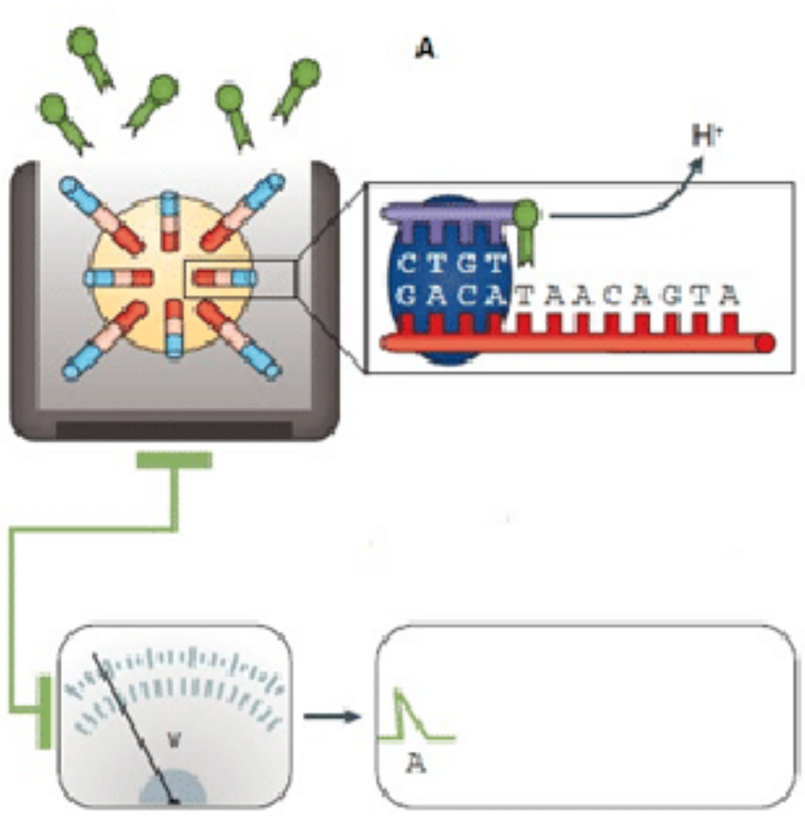
The **PyroMark** uses Pyrosequencing technology for real-time, sequence-based detection and quantification of sequence variants and epigenetic methylation. The PyroMark Q24 is highly suited for the analysis of CpG methylation, SNPs, insertion/deletions, STRs, variable gene copy number, as well as for microbial identification and resistance typing.

Pyrosequencing (pyrophosphate)



The first step cleaves the triphosphate nucleotide after an addition, releasing pyrophosphate. The second step converts pyrophosphate into adenosine triphosphate (ATP) via the enzyme ATP sulfurylase. The third step uses the newly synthesized ATP to catalyze the conversion of luciferin into oxyluciferin via the enzyme luciferase and this reaction generates a quanta of light that is captured from the picotiter plate by a charge- coupled camera.

Ion Torrent (semiconductor technology)



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MiniSeq System

1.8-7.5 Gb
8-25 million
2 x 150 bp
50

MiSeq Series

0.3-15 Gb
1-25 million
2 x 300 bp
384

NextSeq Series

20-120 Gb
130-400 million
2 x 150 bp
96

HiSeq Series

125-1500 Gb
2.5-5 billion
2 x 150 bp
12

HiSeq X Series

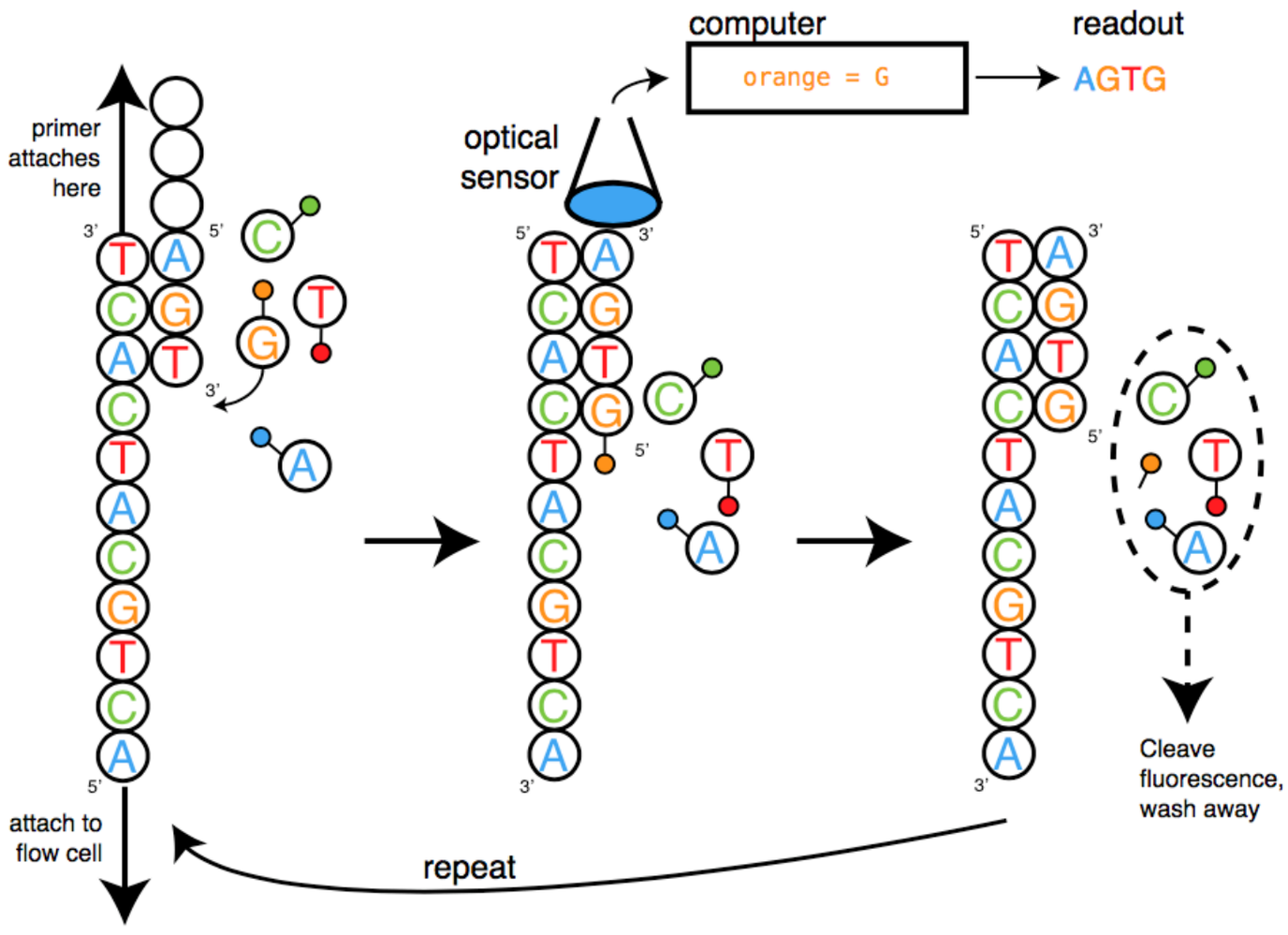
900-1800 Gb
3-6 billion
2 x 150 bp
16

NovaSeq Series

134-6000 Gb
Up to 20 billion
2 x 150 bp
48

http://www.illumina.com

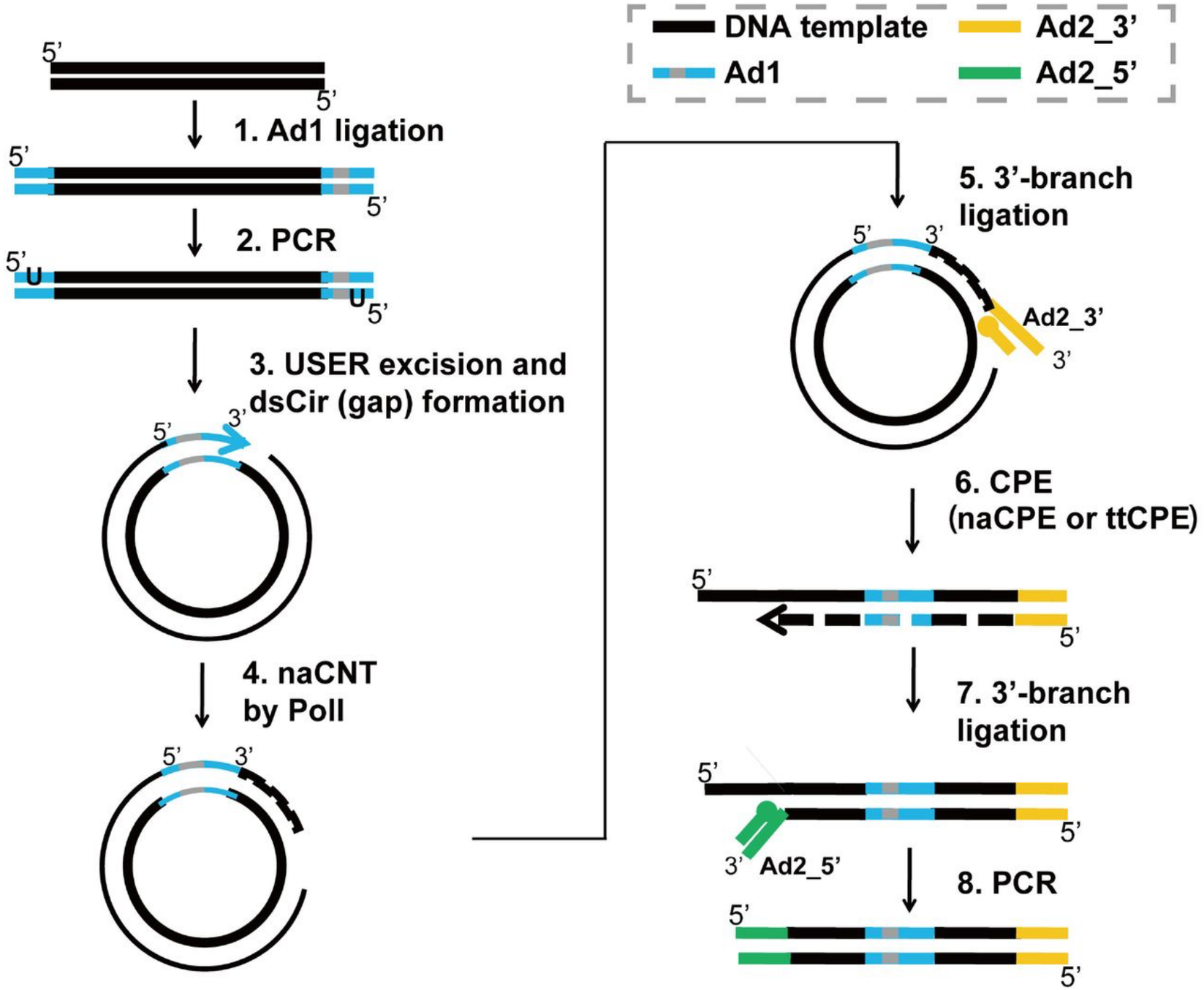
Sequencing by Synthesis (fluorescent)



Sequencing by Synthesis. dNTP fluorescence is translated to a base call.

	 Sequencers ⊕	 Sequencers ⊕	 Sequencers ⊕	 Sequencers ⊕
Product Model	DNBSEQ-T7	DNBSEQ-G400	DNBSEQ-G50	DNBSEQ-G400 FAST
Features	Ultra-high Throughput	Adaptive	Effective	Fast
Applications	Whole Genome Sequencing,Deep Exome Sequencing,Transcriptome Sequencing,and Targeted Panel Projects.	WGS, WES, Transcriptome sequencing and more	Small whole genome sequencing, targeted DNA/RNA panels, low-pass whole genome sequencing	Targeted DNA, RNA, Epigenetics and clinical applications
Flow Cell Type	FC	FCL & FCS	FCL & FCS	FCS
Lane/Flow Cell++	1 lane	4 lane & 2 lane	1 lane	2 lane
Operation Mode	Ultra-high Throughput	High Throughput	Medium Throughput	Medium Throughput
Max. Throughput / RUN	6Tb	1440Gb	150Gb	330G
Effective Reads / Flow Cell	5000M	1500-1800M	500M / 100M	550M
Average run time	PE150 within 24 hours	~38 hours	10-66 hours	12-37 hours
Min. Read Length	PE100	SE50	SE50	SE100
Max. Read Length	PE150	SE400	PE150	PE150

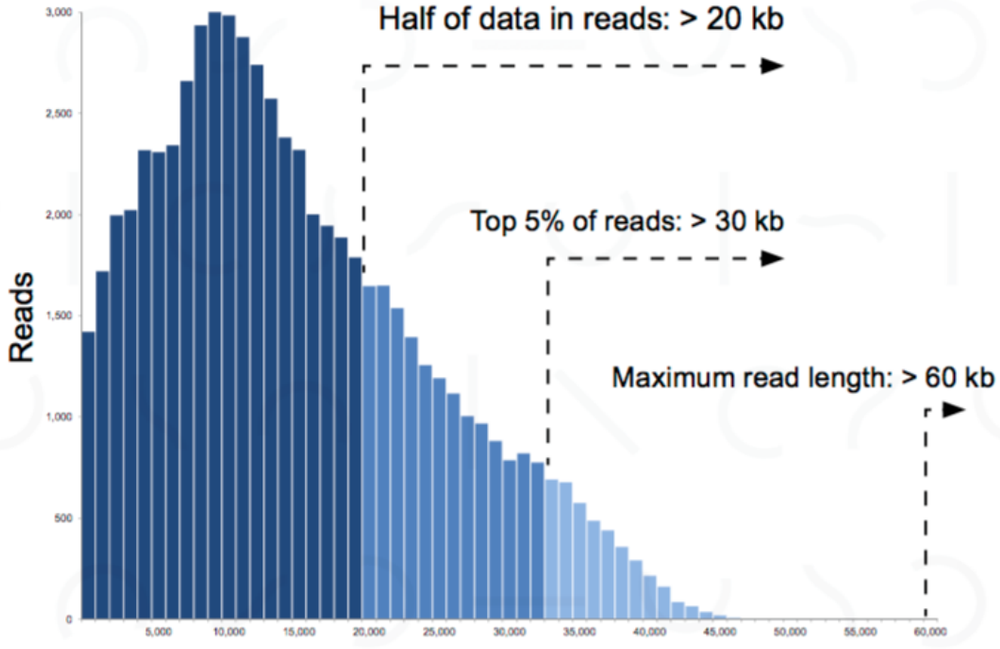
https://en.mgitech.cn


DNA template
Ad1
Ad2_3'
Ad2_5'
5'
5'
1. Ad1 ligation
5'
5'
2. PCR
5'
U
U
5'
3. USER excision and
dsCir (gap) formation
5'
3'
4. naCNT
by PolI
5'
3'
5. 3'-branch
ligation
5'
3'
Ad2_3'
3'
6. CPE
(naCPE or ttCPE)
5'
5'
7. 3'-branch
ligation
5'
5'
3'
Ad2_5'
8. PCR
5'
5'

PacBio RS II

Long Read Lengths

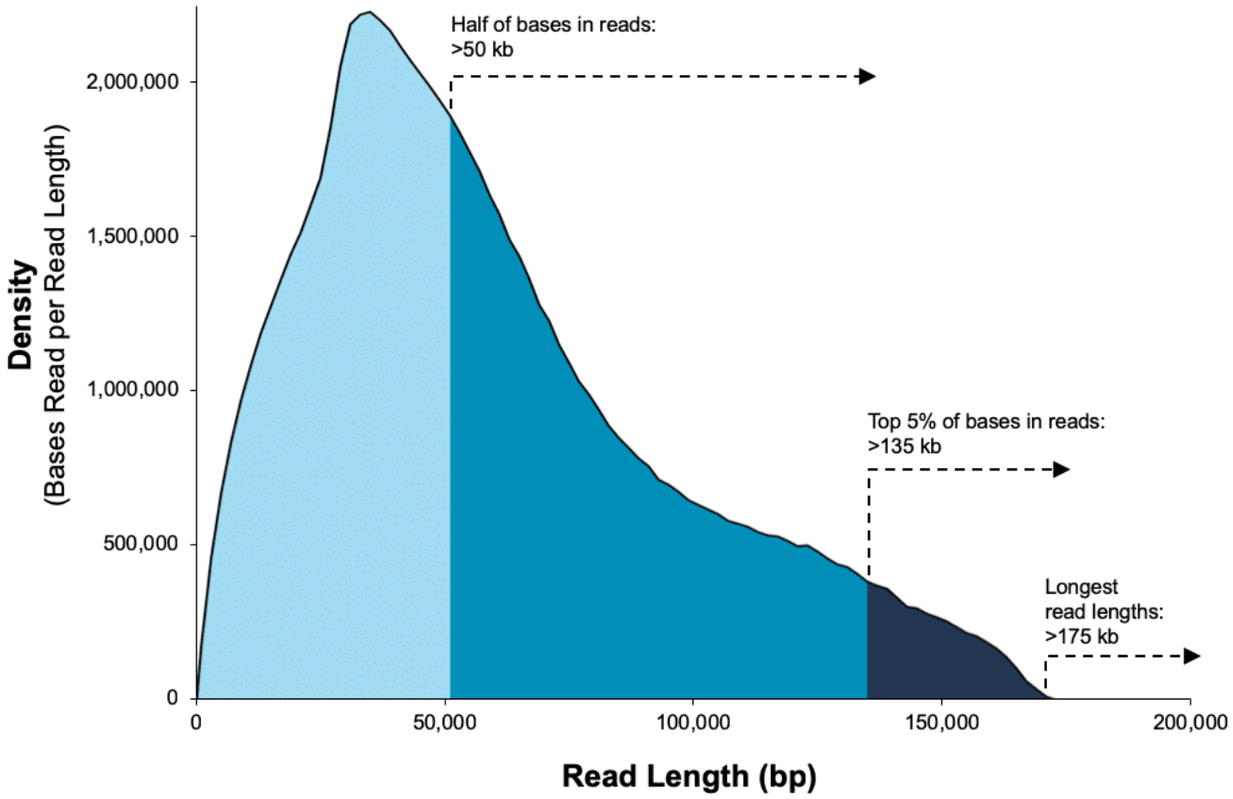
Read lengths > 20 kb
Data per SMRT Cell: 750 Mb – 1.25 Gb



Read-length data shown above is from a 20 kb size-selected human library run on a PacBio RS II (6-hour movie, P6-C4 chemistry). The PacBio RS II SMRT Cells generate ~55,000 reads. The Sequel System generates ~370,000 reads per SMRT Cell.

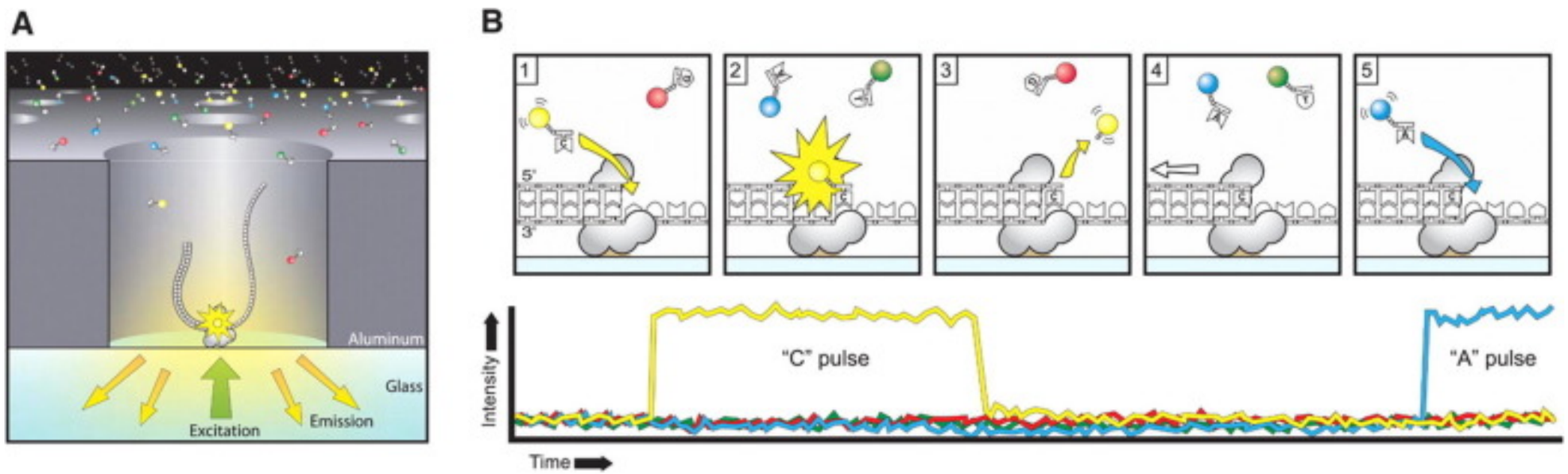
http://www.pacb.com

Sequel



Data from a 35 kb size-selected *E. coli* library using the SMRTbell Express Template Prep Kit 2.0 on a Sequel II System (1.0 Chemistry, Sequel II System Software v7.0, 15-hour movie)*.

PacBio (fluorophore)



Oxford NANOPORE Technologies

SmidgION

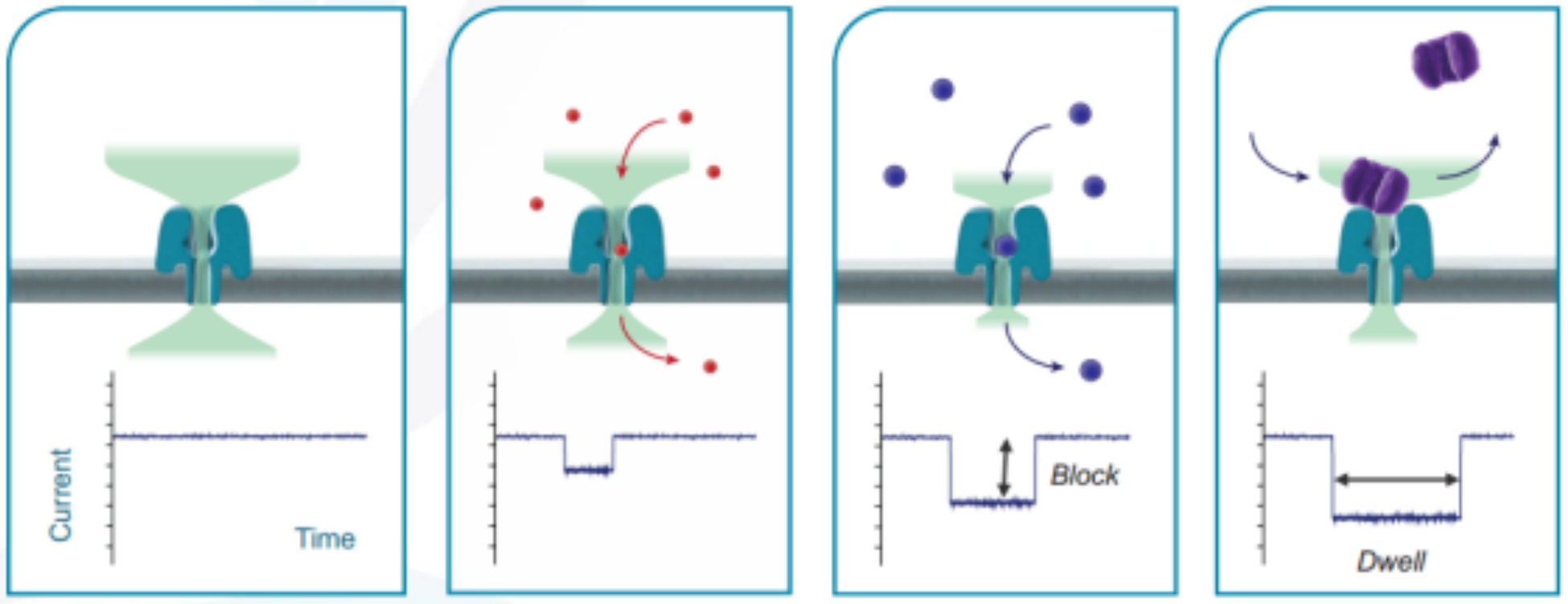
Flongle

MinION

GridION

PromethION

https://www.nanoporetech.com


Current
Time
Block
Dwell

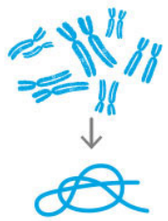
Helicos BioSciences Corporation

Genetic Diversity Centre Zurich

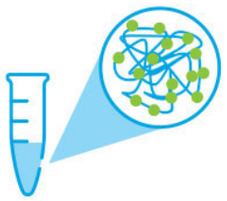
Genetic Diversity Centre Zurich

Customer Sample
- Blood
- Cells
- Tissue
- Microbes

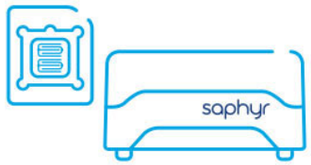
Isolate High Molecular Weight DNA



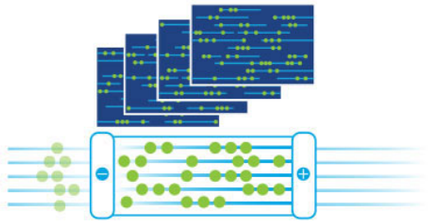
Label Specific Sequences Across the Entire Genome



Transfer Labeled DNA into Cartridge for Scanning

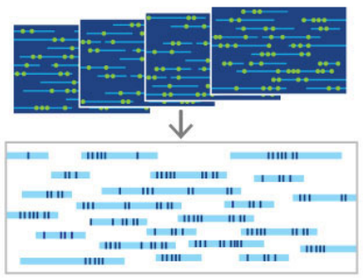


Load, Linearize & Image Labeled DNA in Repeated Cycling to Scan Whole Genome



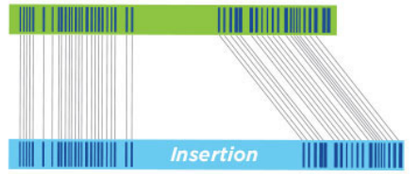
High-throughput, High-resolution Imaging of Megabase Length Molecules

Algorithms Convert Images into Molecules



Assembly Algorithms Align Molecules *de novo* to Construct Consensus Genome Maps

Cross-Mapping Across Multiple Samples or to a Reference



- Automated SV Detection
- Scaffolding