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GD Diversity
Centre
Zurich
```

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          GCATGCATGCATGCATGCATGCATGCATGCAT
        ATGCATGCATGCATGCATGCATCCATGCATGCATGCA
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     *TGCATGCATGCA
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TGC

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Biocomputing with R

Niklaus Zemp 17 June 2025

Genetic Diversity Centre (GDC)

Bioinformatics

ETH Zurich



ATCHICATE CONTROLLED EXAMPLE EXAMPLE

Write an R script (R code) that creates a text file containing the results of 10 rolls of a pair of dice.

Rprojects Use Al Tools R-packages

Comments set.seed() pip

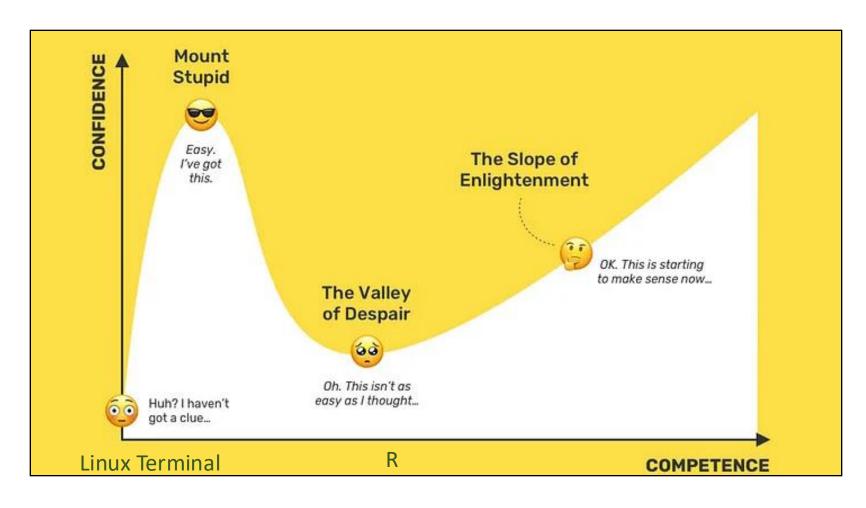
loops own functions tidyverse rMarkdown

Good scripting involves writing code that is not only functional but also maintainable, readable, efficient and flexible.



ning-Kruger effect



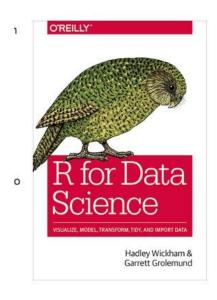




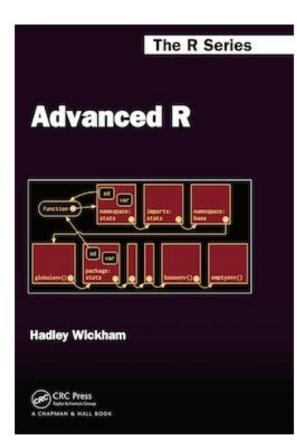




Many many tutorials, forum, YouTube videos posts and books available









TREMEDIA CONTROL CONTR

Currently, the CRAN package repository features 22507 available packages.

Table of available packages, sorted by date of publication

Table of available packages, sorted by name





About Bioconductor

Bioconductor uses the R statistical programming language, and is open source and open development. It has two releases each year, and an active user community.



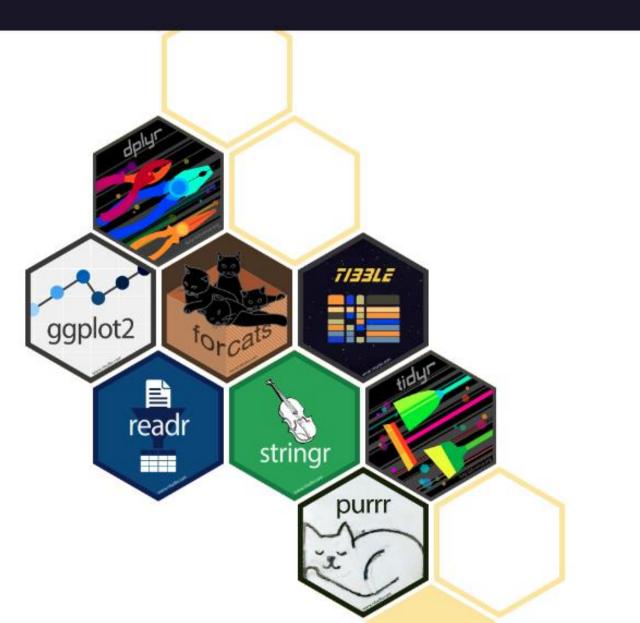
Packages

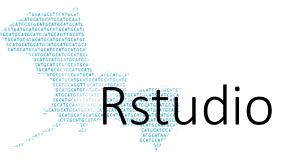
Discover <u>2341</u> software packages available in Bioconductor release 3.21.

Bioconductor 3.21 release announcement

Tidyverse







enetic Diversity Centre (GDC) ourse Webpage

quirements

nux 1 - Local Terminal

านx 2 - Remote Terminal

ocomputing

ocomputing with R

ocomputing on a HPC cluster

set working directory

file.edit("Demo.R")

producible Research

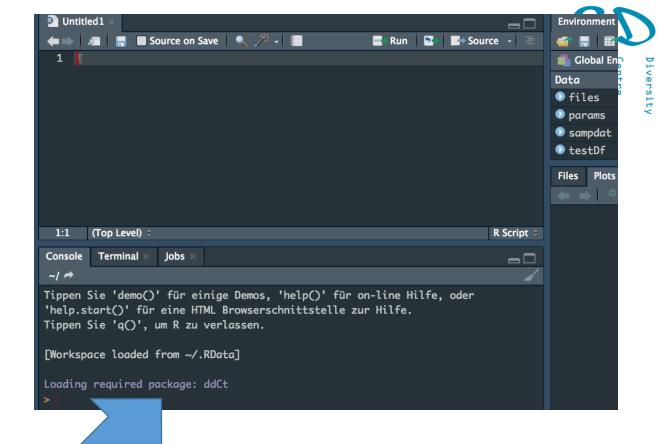
PS (NGS)

Biocomputing with R



```
### generate folder
dir.create("Demo")
setwd("Demo")

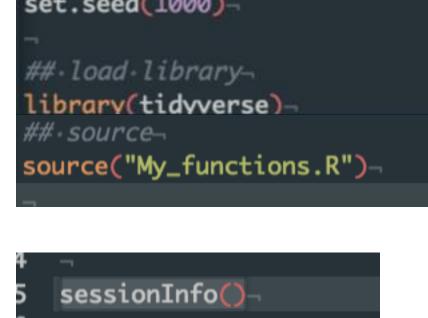
### download scripts for Demo
utils::download.file("https://www.gdc-docs.ethz.ch/GeneticDiversityAnalysis/GDA
utils::download.file("https://www.gdc-docs.ethz.ch/GeneticDiversityAnalysis/GDA
### open R scripts
file.edit("My_functions.R")
```



Copy-paste



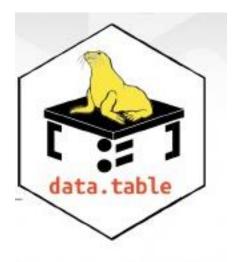
```
##·set·working·directory-
setwd("~/Desktop")-
#.Prepare.workplace.----
##install-library-
install.packages("tidyverse")-
##-remove-all-variables-
rm(list = ls())-
## set seed
set.seed(1000)-
##-load-library-
library(tidyverse)
##·source-
source("My_functions.R")-
```





Affection of the control of the cont

```
dat <- read.csv("http://gdc-web.ethz.ch/gdc-analysis-course/2019/data/Students19.txt", header = T, sep="\t")
dat <- read_tsv("http://gdc-web.ethz.ch/gdc-analysis-course/2019/data/Students19.txt")</pre>
```





data.table::fread

The state of the s

- Data frames
- Lists
- Vectors
- Matrixes

as.logical	TRUE, FALSE, TRUE	Boolean values (TRUE or FALSE).
as.numeric	1, 0, 1	Integers or floating point numbers.
as.character	'1', '0', '1'	Character strings. Generally preferred to factors.
as.factor	'1', '0', '1', levels: '1', '0'	Character strings with preset levels. Needed for some statistical models.

```
str(iris)
```

```
## 'data.frame': 150 obs. of 5 variables:
## $ Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
## $ Sepal.Width : num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
## $ Petal.Length: num 1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
## $ Petal.Width : num 0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
## $ Species : Factor w/ 3 levels "setosa", "versicolor",..: 1 1 1 1 1 1 1 1 1 1 1 ...
```



ilaale |

tibble

as_tibble(iris)

```
## # A tibble: 150 x 5
      Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##
             <dbl>
                          <dbl>
                                       <dbl>
                                                    <dbl> <fct>
               5.1
                            3.5
                                         1.4
                                                      0.2 setosa
               4.9
                                                      0.2 setosa
               4.7
                            3.2
                                         1.3
                                                     0.2 setosa
                            3.1
               4.6
                                         1.5
                                                      0.2 setosa
                            3.6
                                         1.4
                                                     0.2 setosa
               5.4
                            3.9
                                         1.7
                                                      0.4 setosa
               4.6
                            3.4
                                         1.4
                                                     0.3 setosa
                            3.4
                                         1.5
                                                      0.2 setosa
                            2.9
                                         1.4
                                                      0.2 setosa
## 10
                            3.1
                                         1.5
                                                      0.1 setosa
## # ... with 140 more rows
```



Manipulation ATTENDED TO THE PROPERTY OF THE

```
iris.df <- data.frame(iris$Sepal.Length, iris$Sepal.Width, iris$Species)</pre>
```

Replace patterns

```
iris.df$Species2 <- gsub("setosa", "Setosa", iris.df$iris.Species)
```

subset

```
iris.df.sub <- subset(iris.df, iris.df$iris.Species == "setosa")</pre>
```

order table

```
iris.df.orderd <- iris.df[order(iris.df$iris.Sepal.Length), ]</pre>
```



Manipulation

```
iris.tb <- as_tibble(iris) %>%
    select(Species, Sepal.Length, Sepal.Width, Petal.Length) %>%
    dplyr::filter(Species != "setosa") %>%
    mutate(Species2 = gsub("versicolor", "Versicolor", Species)) %>%
    arrange(., Sepal.Length)
```







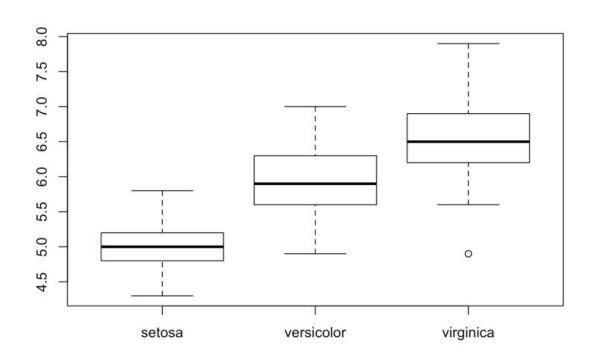


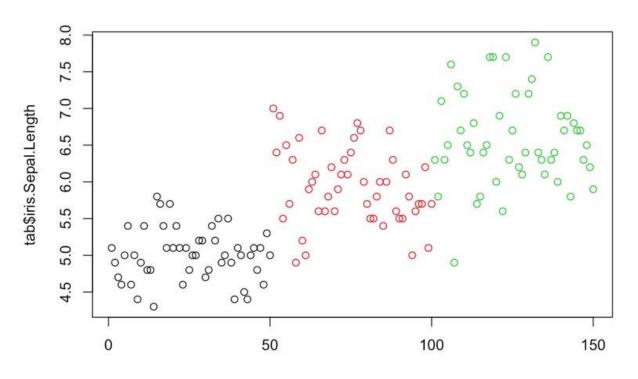


REGISTATION OF THE PROPERTY OF

boxplot(tab\$iris.Sepal.Length~tab\$iris.Species)

plot(tab\$iris.Sepal.Length, col=tab\$iris.Species)

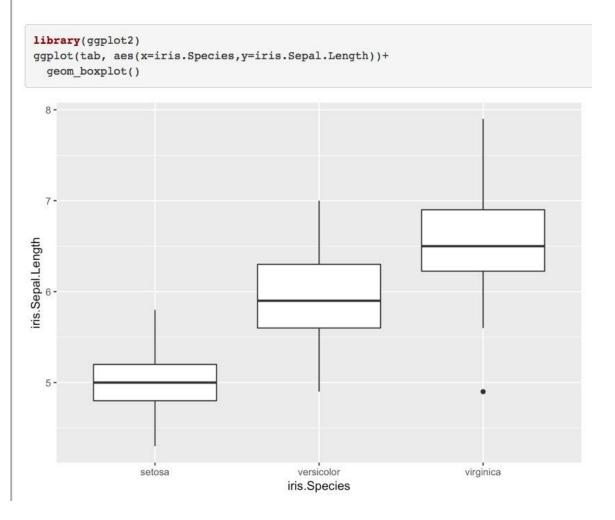


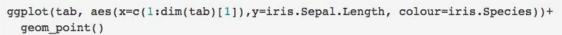


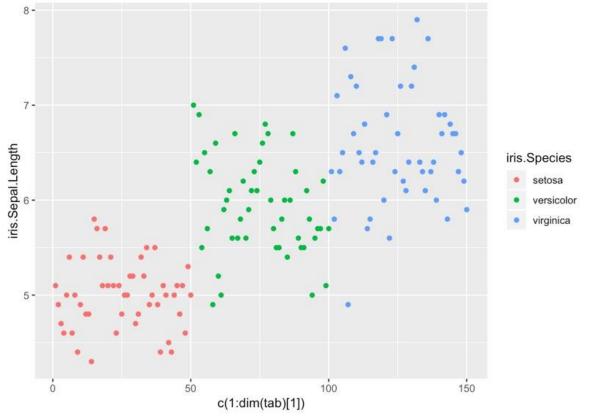


alization-ggplot2





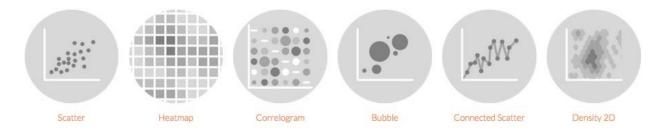




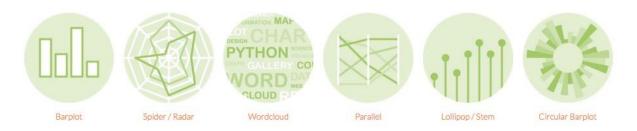
Distribution



Correlation



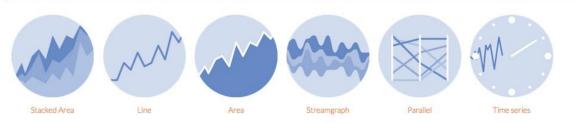
Rankings



Part of a whole



Evolution



Maps



Flow



Other

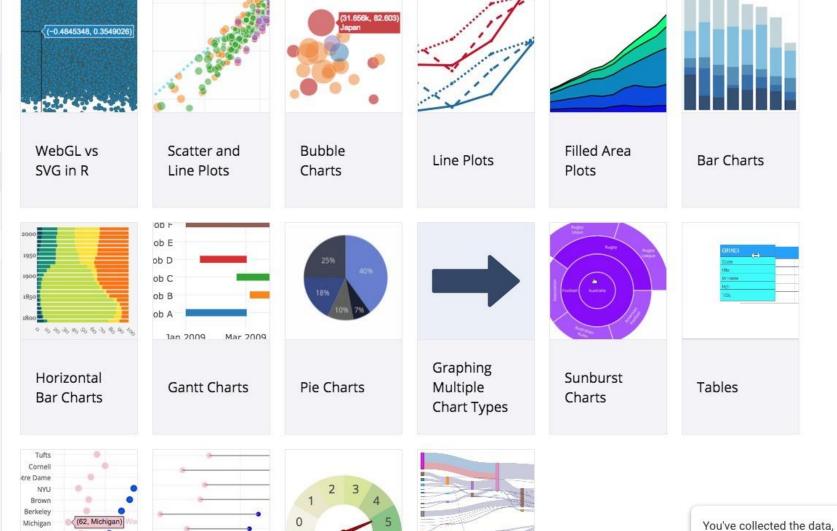


https://www.r-graph-gallery.com/

SoCal

Dot Plots

Dumbbell



Sankey

Gauge Charts

agonized over the right mor and now you've GOT to con management.....

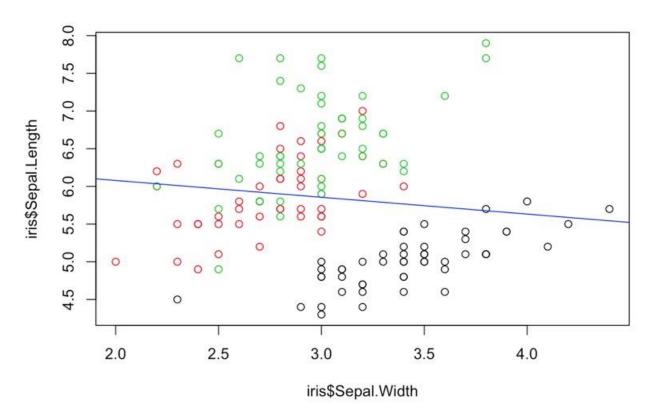


Scale-Location

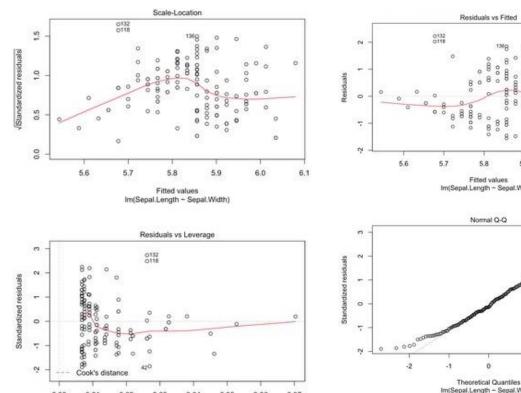
The strict using R-Linear models

m1<-lm(Sepal.Length~Sepal.Width,data=iris)

plot(iris\$Sepal.Length~iris\$Sepal.Width, col=iris\$Species)
abline(m1\$coefficients, col="blue")



plot(ml)





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save specific objects to a file

```
save(iris,file="iris.RData")
```

load object

```
load("iris.RData")
```

save workspace

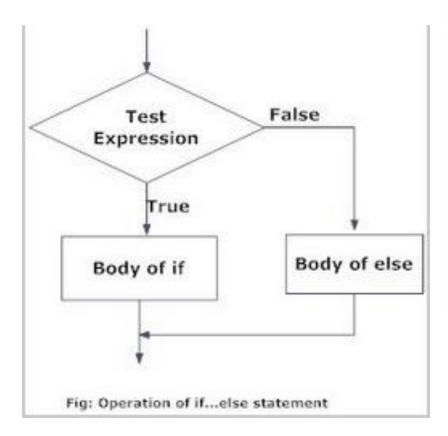
```
save.image(file='image.RData')
```

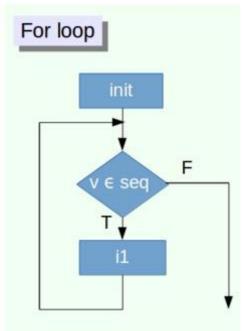
load workspace

```
load("image.RData")
```



Programming in R







Hands-On Programming with R

https://rstudio-education.github.io/hopr/



tomized functions

```
Myfunction <- function(variables) {</pre>
```

Function1
Function2
value/plot
}

```
ploting <- function(n_samples) {
  dat <- rnorm(n_samples, 100, 5)
  plot(dat)
}

ploting(10000)</pre>
```

Arterior de la constitución de l

Shiny from R Studio

Iris k-means clustering 1 show below ui.R function(input, output, session) { X Variable # Combine the selected variables into a new data frame Sepal.Length selectedData <- reactive({ iris[, c(input\$xcol, input\$ycol)] Y Variable Sepal.Width clusters <- reactive({ kmeans(selectedData(), input\$clusters) Cluster count 3 output\$plot1 <- renderPlot({ palette(c("#E41A1C", "#377EB8", "#4DAF4A", "#984EA3", 2.5 "#FF7F00", "#FFFF33", "#A65628", "#F781BF", "#999999")) par(mar = c(5.1, 4.1, 0, 1))2.0 plot(selectedData(), col = clusters()\$cluster, 4.5 6.5 8.0 5.0 5.5 7.0 7.5 pch = 20, cex = 3) Sepal.Length points(clusters()\$centers, pch = 4, cex = 4, lwd = 4)

https://shiny.rstudio.com/gallery/kmeans-example.html

Customized packages

Back to Gallery







Write an R script (R code) that creates a text file containing the results of 10 rolls of a pair of dice.

Take home message

- Fastly developing
- Many resources for biologists
- Also suitable for programming
- Al tools



Bioinformatics and computational biology, often referred to as "bioinformatics" or "biocomputing," is the application of computer science and technology to the management, analysis, and interpretation of biological data. This can include DNA sequences, protein structures, and genetic and metabolic pathways, as well as data from high-throughput experiments such as genome-wide association studies, transcriptomics, and proteomics.

R is a popular programming language and environment for data analysis that is widely used in bioinformatics and other fields. There are many libraries and packages available in R that are specifically designed for bioinformatics, such as Bioconductor, which provides tools for the analysis of genomic data, and rOpenSci, which provides tools for working with various bioinformatics data resources, such as the National Center for Biotechnology Information (NCBI) databases.

There are many R packages for bioinformatics and computational biology for tasks like sequence alignment, assembly, annotation, gene expression analysis, functional analysis, and more. Some popular bioinformatics R package include:

Biostrings: for working with DNA and RNA sequences

BSgenome: for working with genome assemblies

Bioconductor: It's a collection of R packages for the analysis of genomic data.

ShortRead: for quality control and preprocessing of high-throughput sequencing data

edgeR, DESeq2: for identifying differentially expressed genes from RNA-seq data

GAGE, GOstats, topGO for functional analysis of gene sets

R can also be used to visualize bioinformatics data using packages like ggplot2, heatmap3, and ComplexHeatmap for heatmap visualization, igraph for network visualization, and rgl for 3D visualization of molecular structures.

Overall, R provides a powerful and flexible platform for bioinformatics and computational biology research, with a wide range of tools and resources available for managing, analyzing, and interpreting large and complex biological data sets.