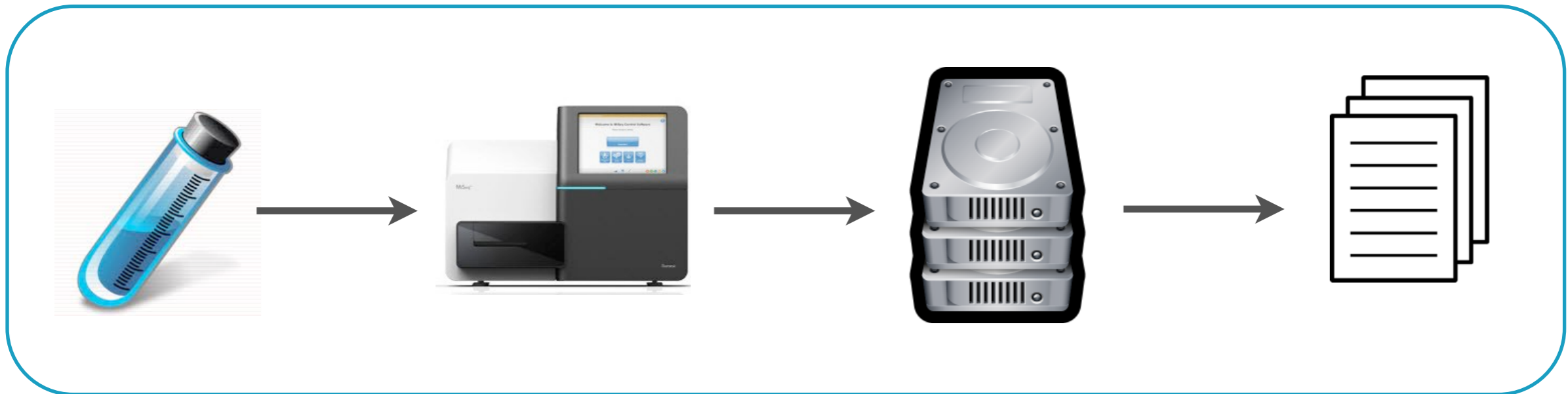
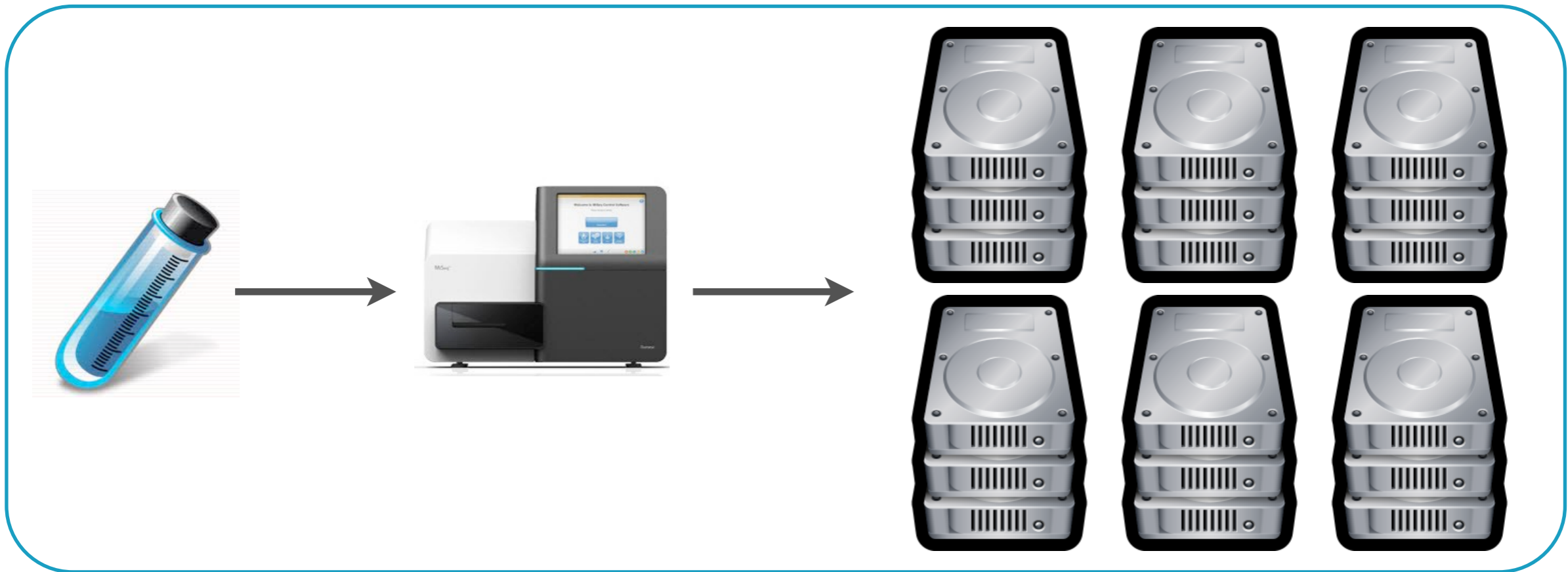


Next (Next) Generation Sequencing **Hype**



Next (Next) Generation Sequencing **Reality**



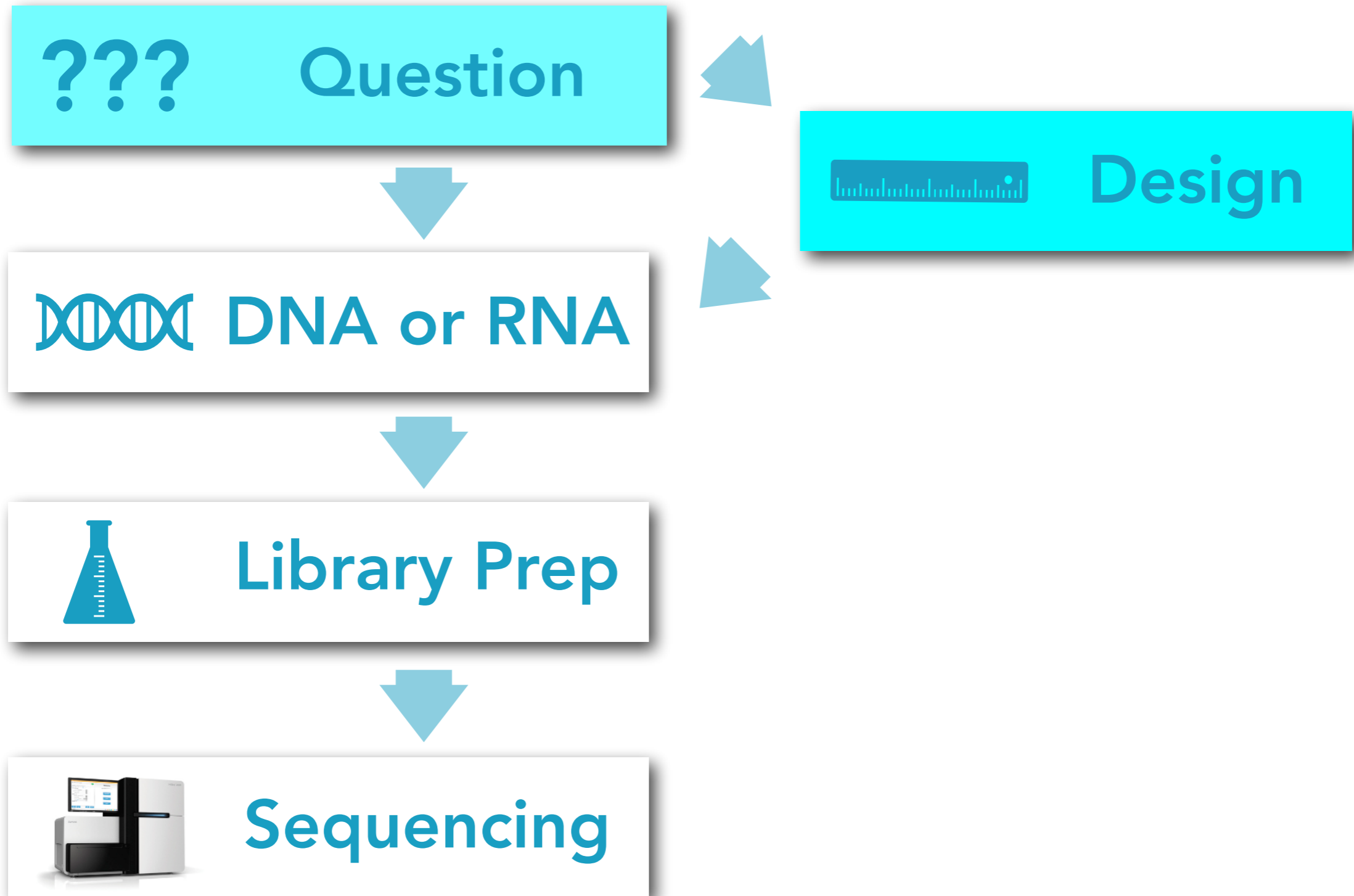


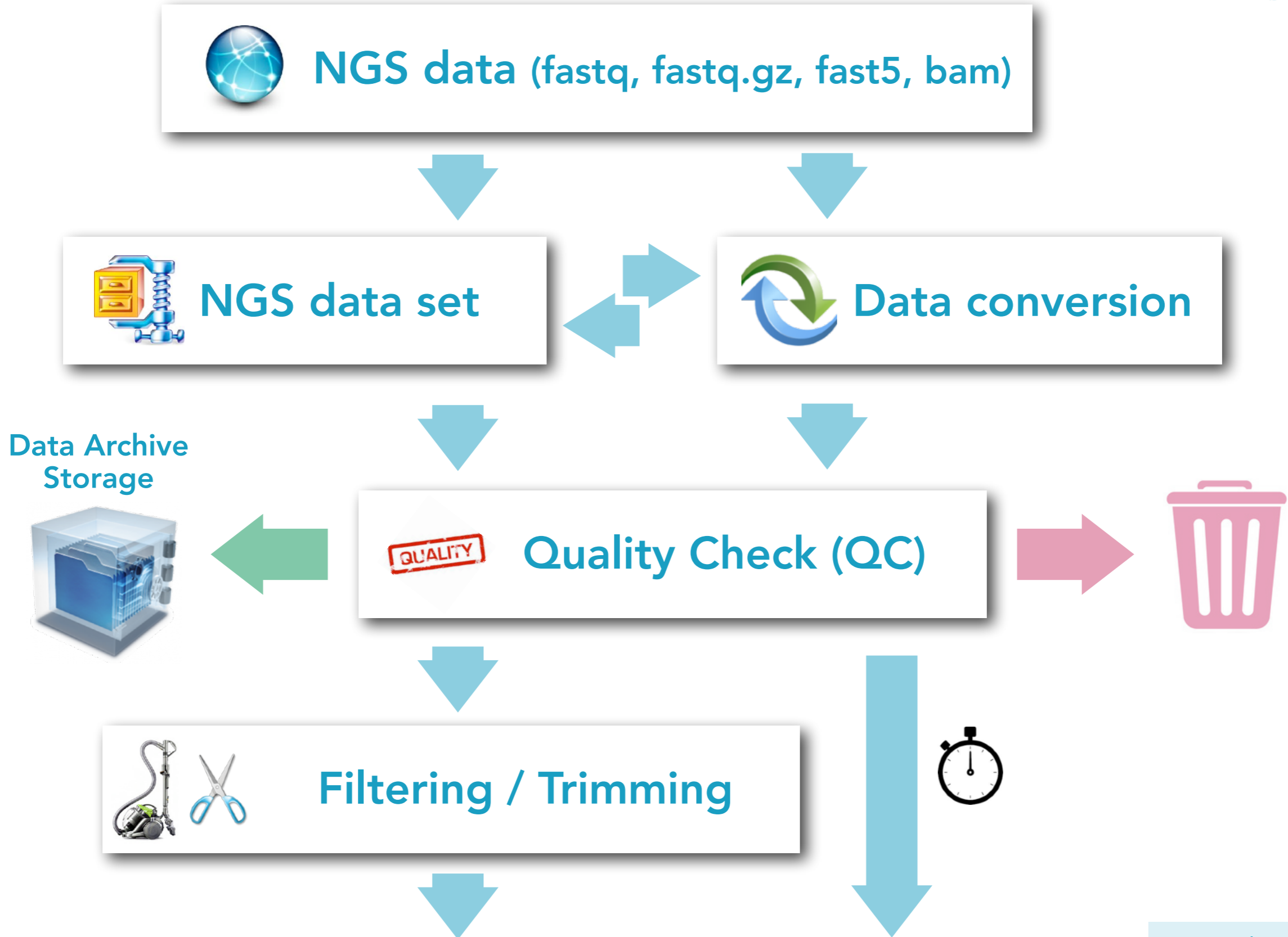
Actually, that's the coffee machine...this is the next-gen sequencer.

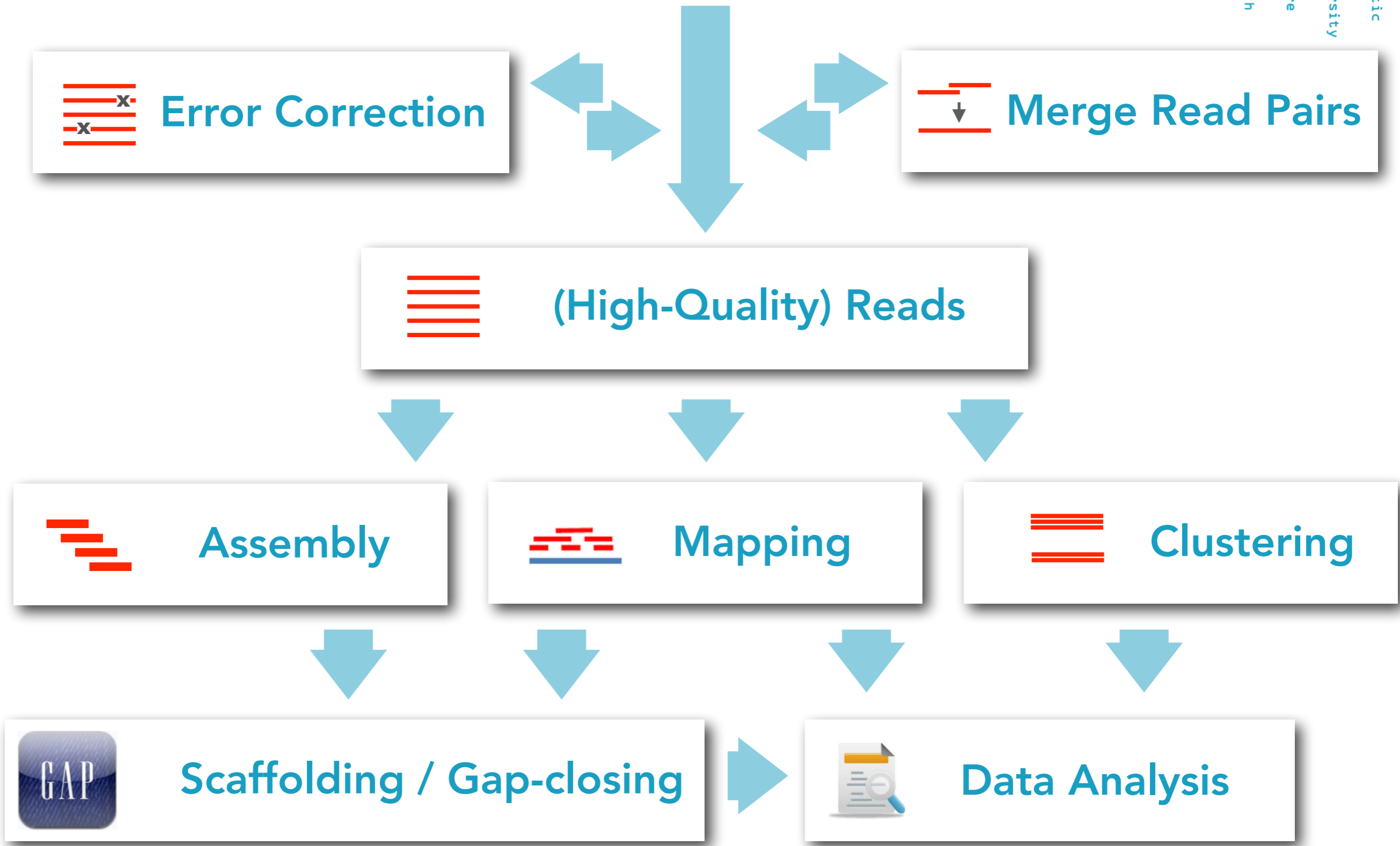
The **First Law of Technology** says we invariably **overestimate** the **short-term impact** of a truly transformational discovery, while **underestimating** its **longer-term effects**.

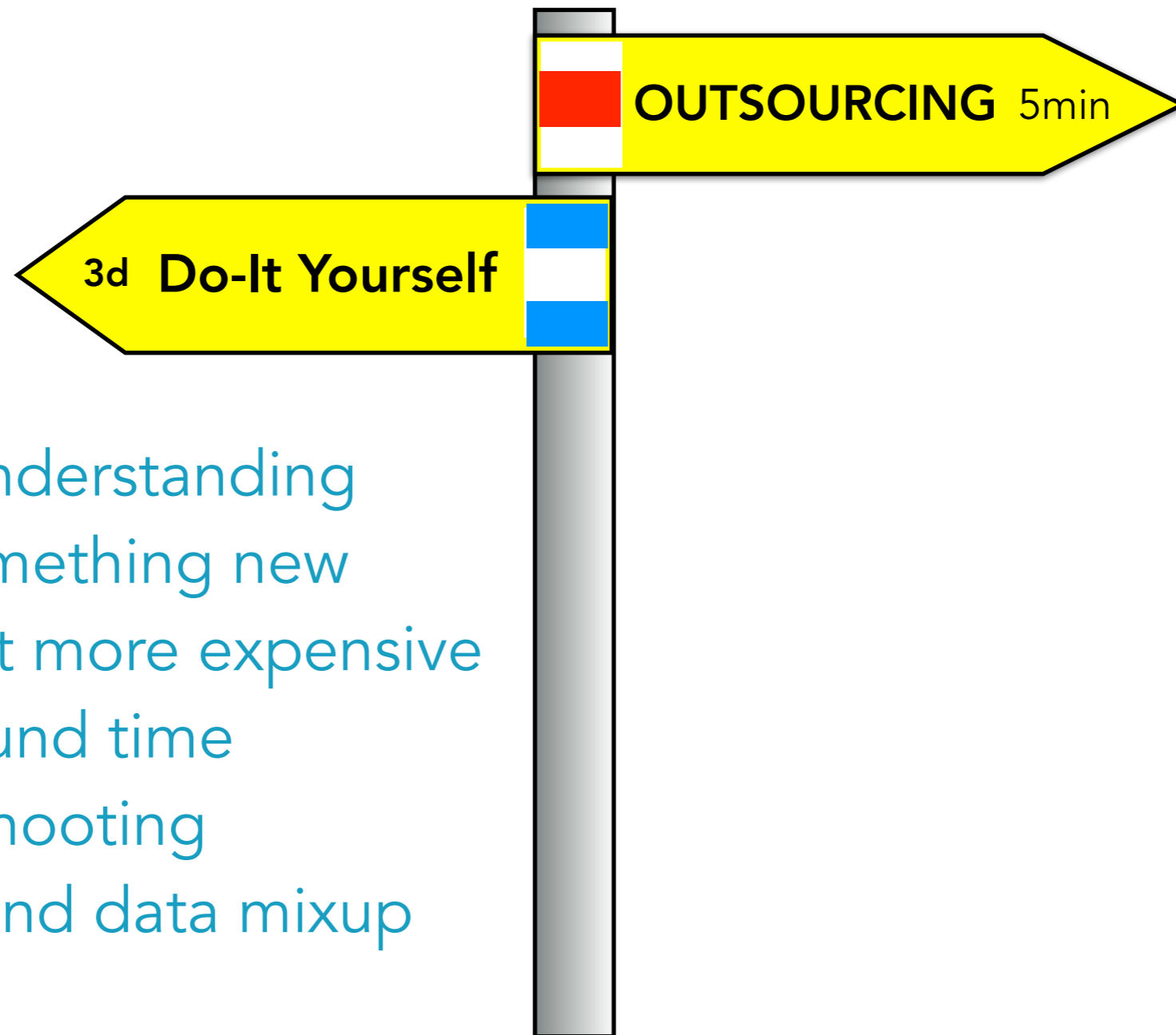
<https://www.scientificamerican.com/>

(A-)Typical Workflow



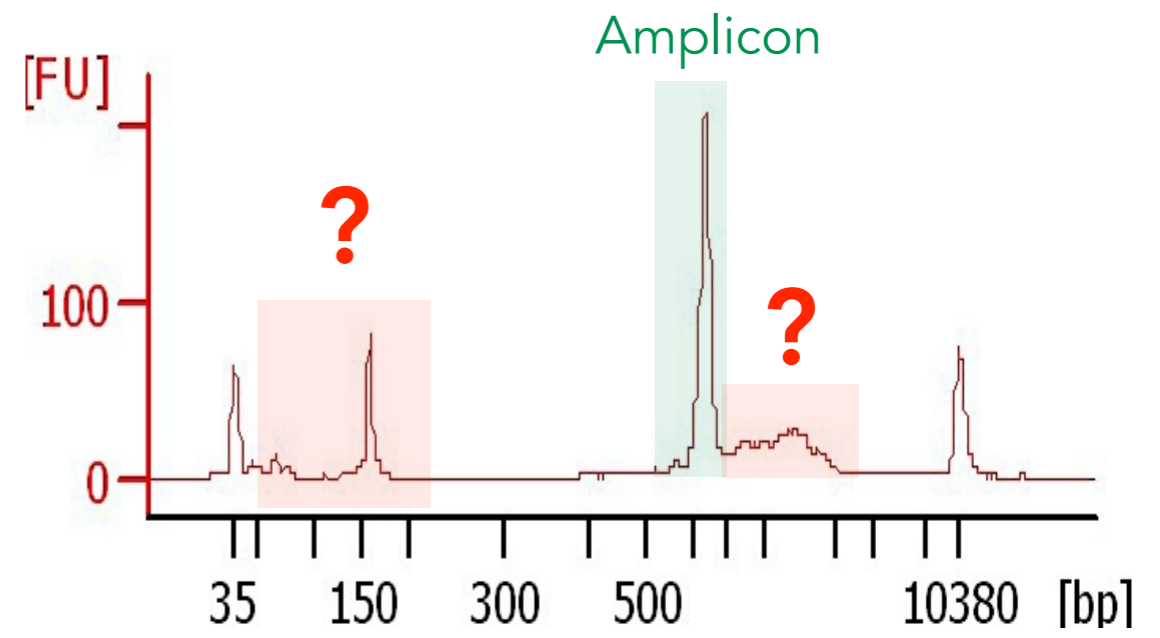
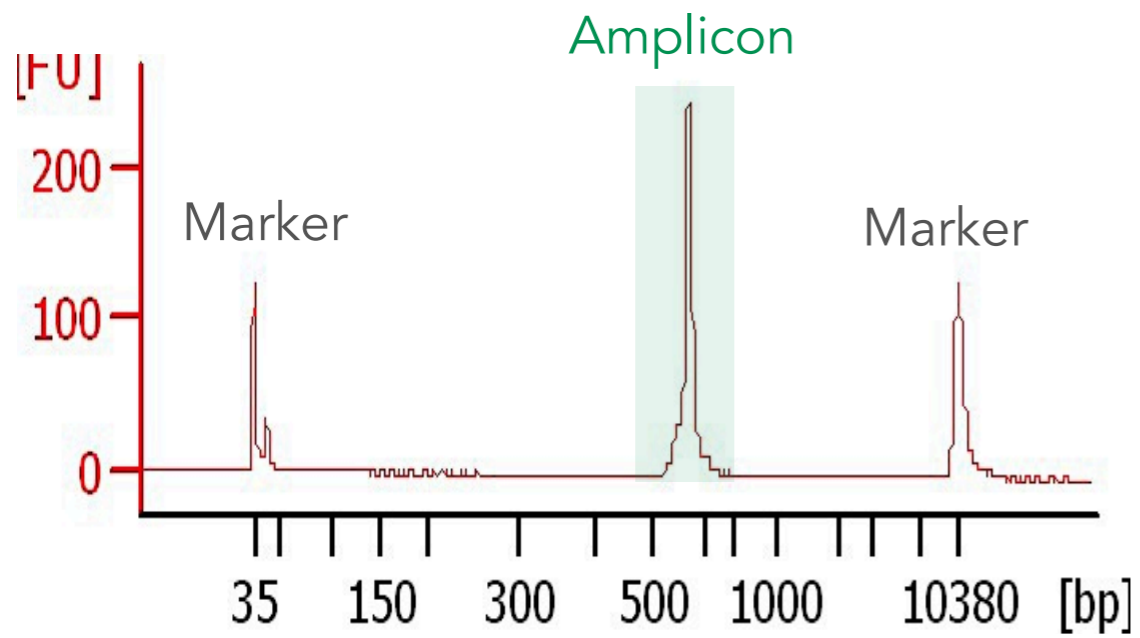






- better understanding
- learn something new
- often not more expensive
- turn-around time
- troubleshooting
- understand data mixup

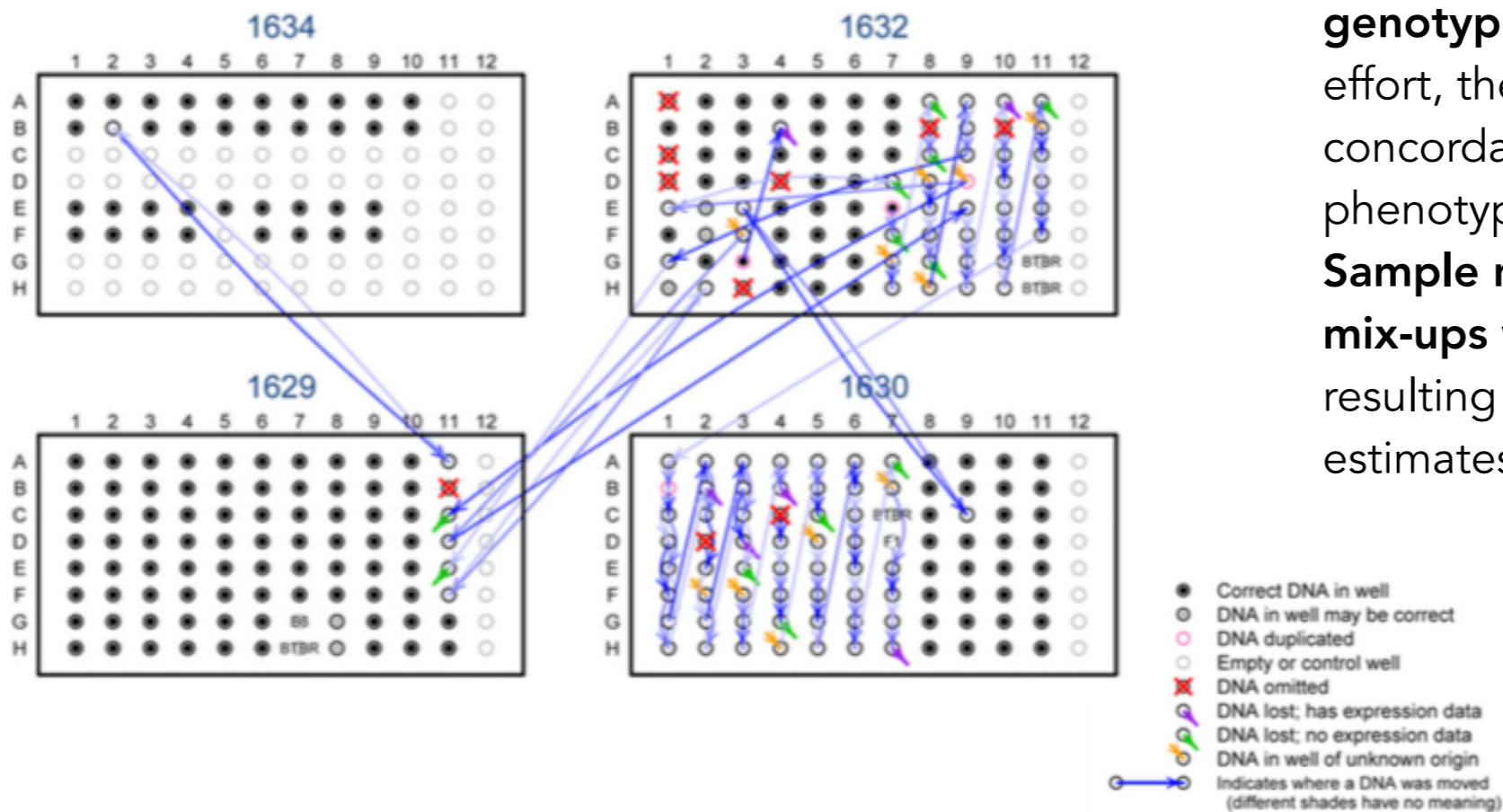




Identification and Correction of Sample Mix-Ups in Expression Genetic Data: A Case Study

Karl W. Broman,^{*2} Mark P. Keller,[†] Aimee Teo Broman,^{*} Christina Kendziorski,^{*} Brian S. Yandell,^{‡,§} Saunak Sen,^{**1} and Alan D. Attie[†]

^{*}Department of Biostatistics and Medical Informatics, [†]Department of Biochemistry, [‡]Department of Statistics, and [§]Department of Horticulture, University of Wisconsin, Madison, Wisconsin 53706, and ^{**}Department of Epidemiology and Biostatistics, University of California, San Francisco, California 94107



“To map the genetic loci influencing a complex phenotype, one seeks to establish an **association between genotype and phenotype**. In such an effort, the maintenance of the concordance between genotyped and phenotyped samples and data is critical. **Sample mislabeling and other sample mix-ups will weaken associations**, resulting in reduced power and biased estimates of locus effects.”

Sequencing Technologies



... the all-in-one NGS platform does not exist (yet)!



1 Sanger (chain termination)

Roche 454 Pyrosequencing (pyrophosphate)

2 Ion Torrent (semiconductor technology)

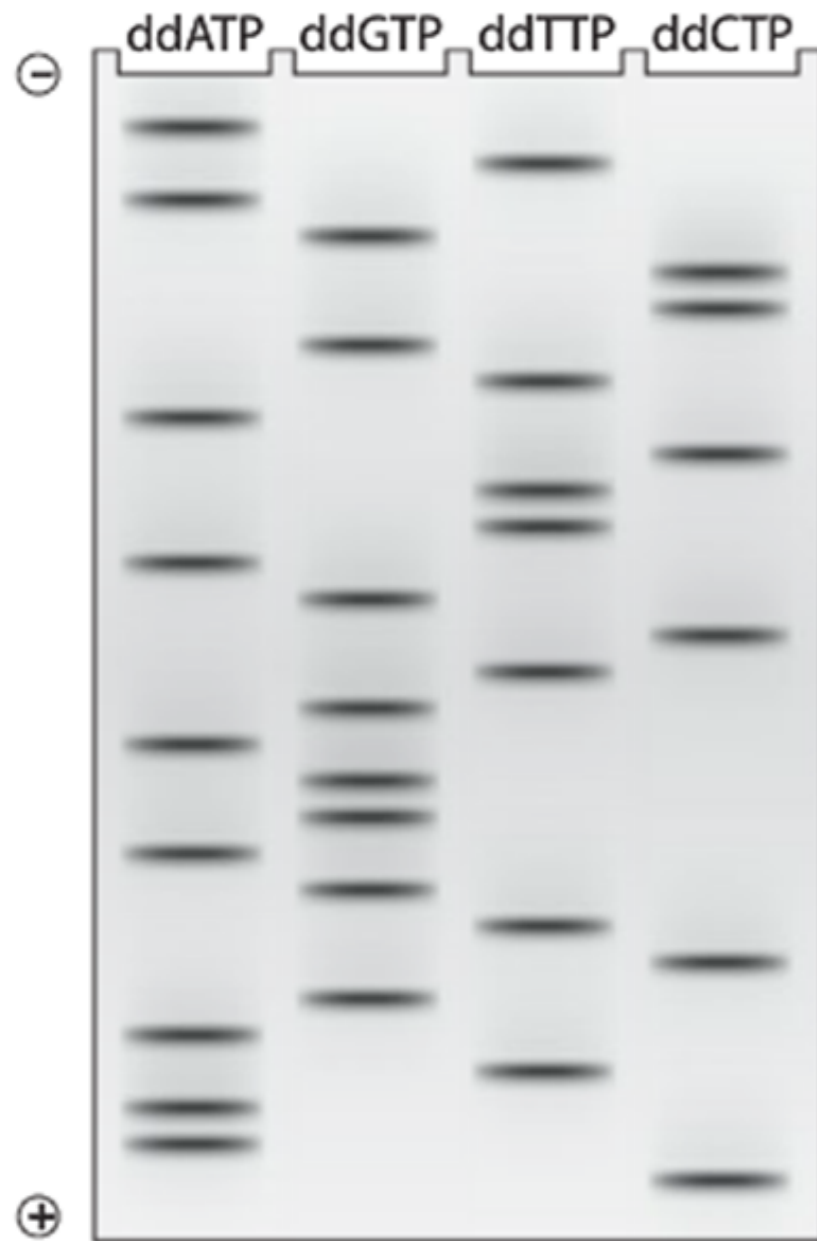
Illumina Sequencing by Synthesis (fluorescent)

PacBio (fluorophore)

3 **Nanopore** (ionic current)

Helicos - SeqLL (fluorescent)

4 Bionano - Saphyr (third-generation optical mapping)



The Nobel Prize in Chemistry 1980
Paul Berg, Walter Gilbert, Frederick Sanger



Paul Berg

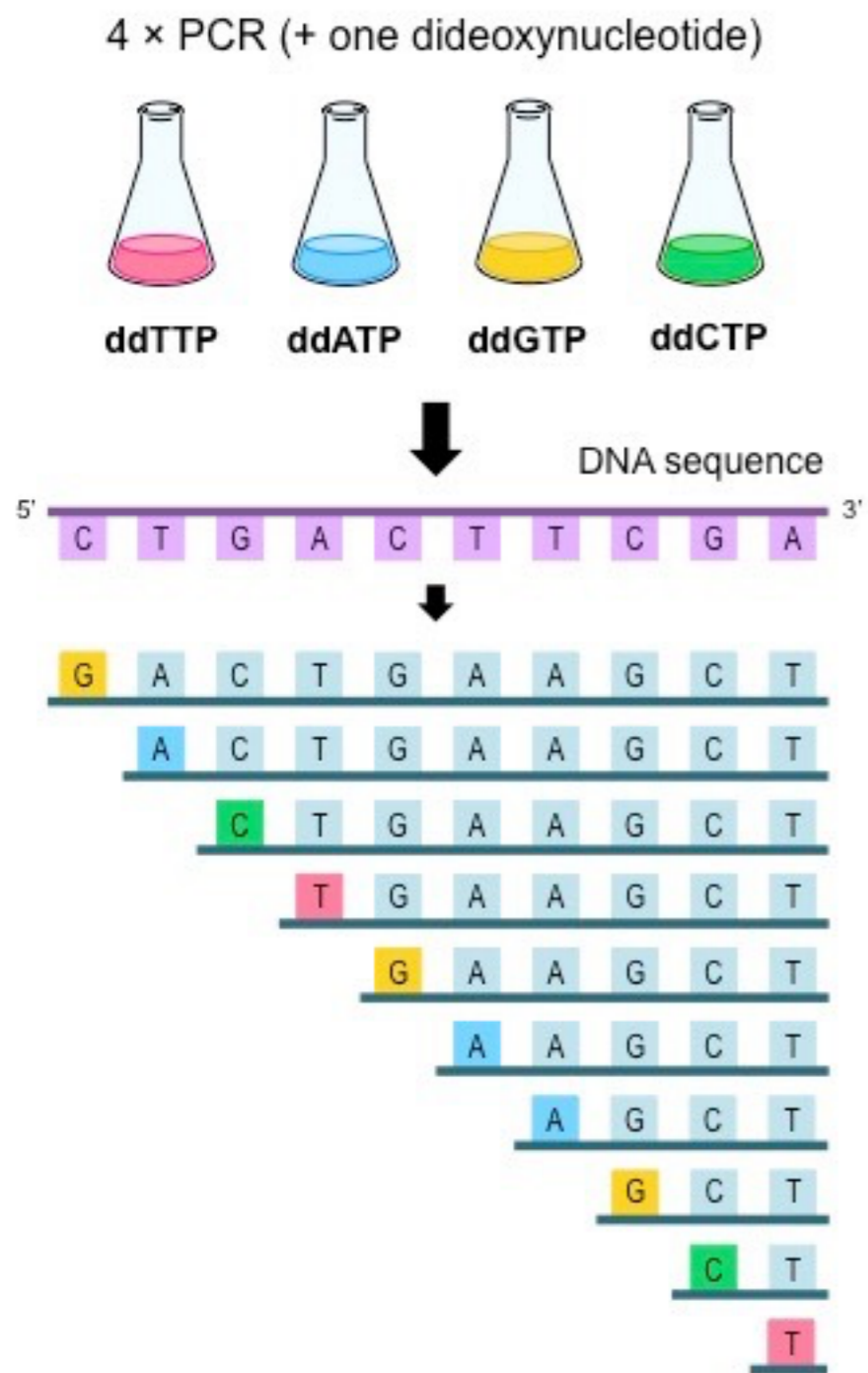


Walter Gilbert



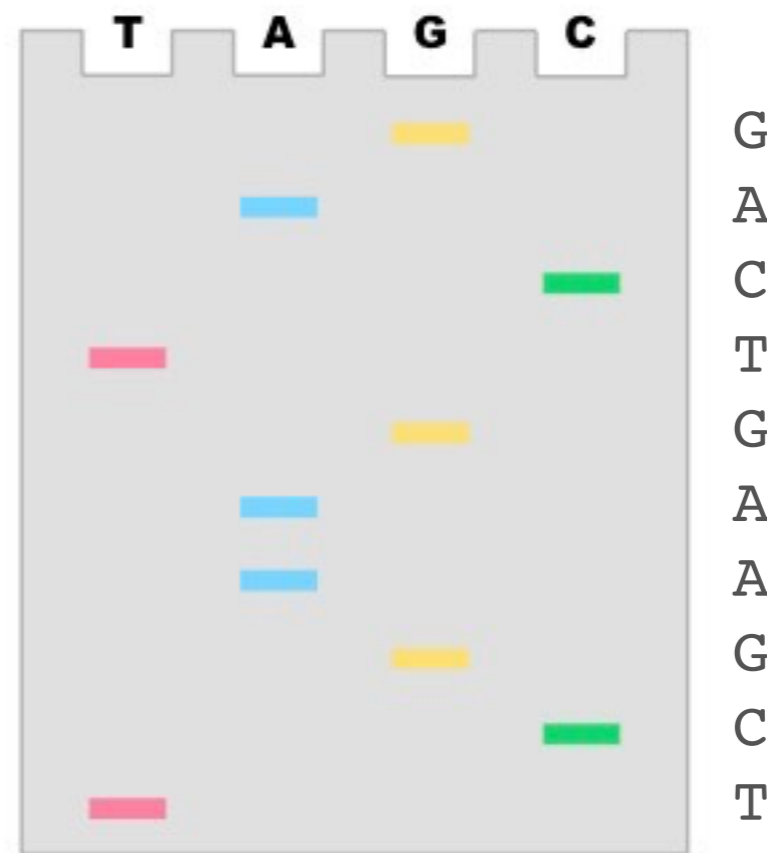
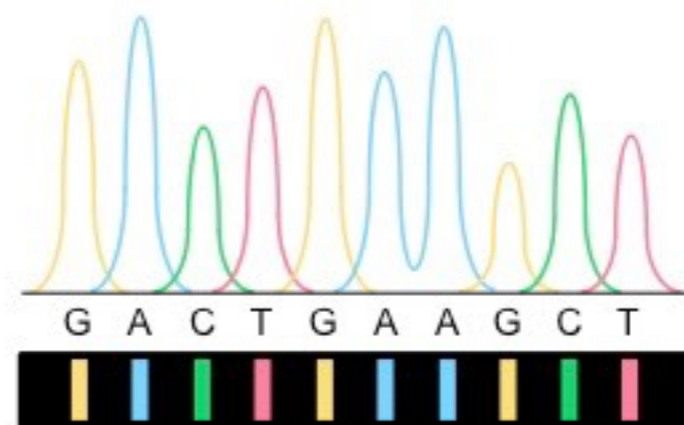
Frederick Sanger

The Nobel Prize in Chemistry 1980 was divided, one half awarded to Paul Berg "for his fundamental studies of the biochemistry of nucleic acids, with particular regard to recombinant-DNA", the other half jointly to Walter Gilbert and Frederick Sanger "for their contributions concerning the determination of base sequences in nucleic acids".



Use a sequencing machine

Separate with a gel





MiniSeq System

1.8-7.5 Gb
8-25 million
2 x 150 bp
50



MiSeq Series

0.3-15 Gb
1-25 million
2 x 300 bp
384



NextSeq Series

20-120 Gb
130-400 million
2 x 150 bp
96



HiSeq Series

125-1500 Gb
2.5-5 billion
2 x 150 bp
12



HiSeq X Series

900-1800 Gb
3-6 billion
2 x 150 bp
16



NovaSeq Series

134-6000 Gb
Up to 20 billion
2 x 150 bp
48

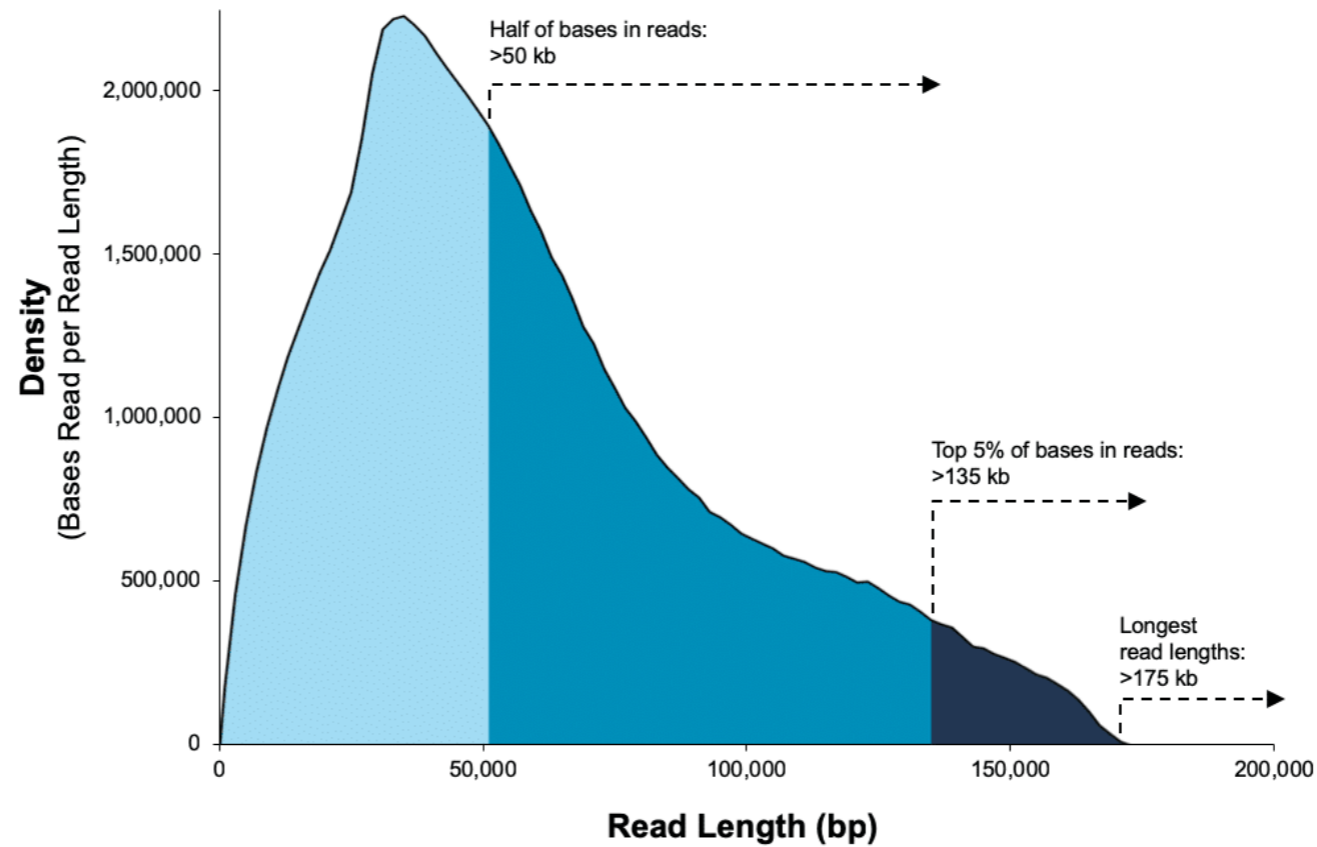
<http://www.illumina.com>



<https://www.pacb.com>



Sequel



Data from a 35 kb size-selected *E. coli* library using the SMRTbell Express Template Prep Kit 2.0 on a Sequel II System (1.0 Chemistry, Sequel II System Software v7.0, 15-hour movie)*.



SmidgION



Flongle



MinION



GridION



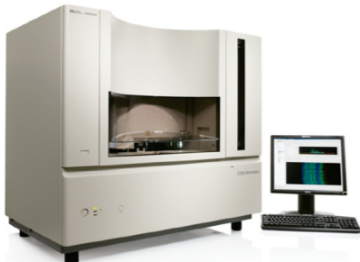
PromethION

<https://www.nanoporetech.com>

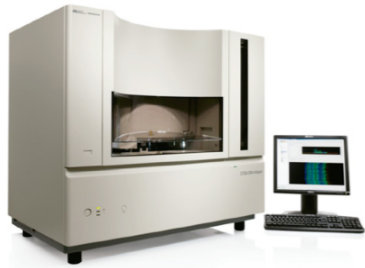




MinION Mk1C



- ▶ Research question
- ▶ Budgeted (including storage and analysis)
- ▶ Read / sequence length
- ▶ Number of reads / coverage
- ▶ Possible contaminants
- ▶ Quality and quantity of template
- ▶ Number of samples
- ▶ Availability



Few but good
sequences



High coverage or
many samples but
shorter reads



Longer sequences
but not so many
samples



Fast results, long
reads but higher
error rate



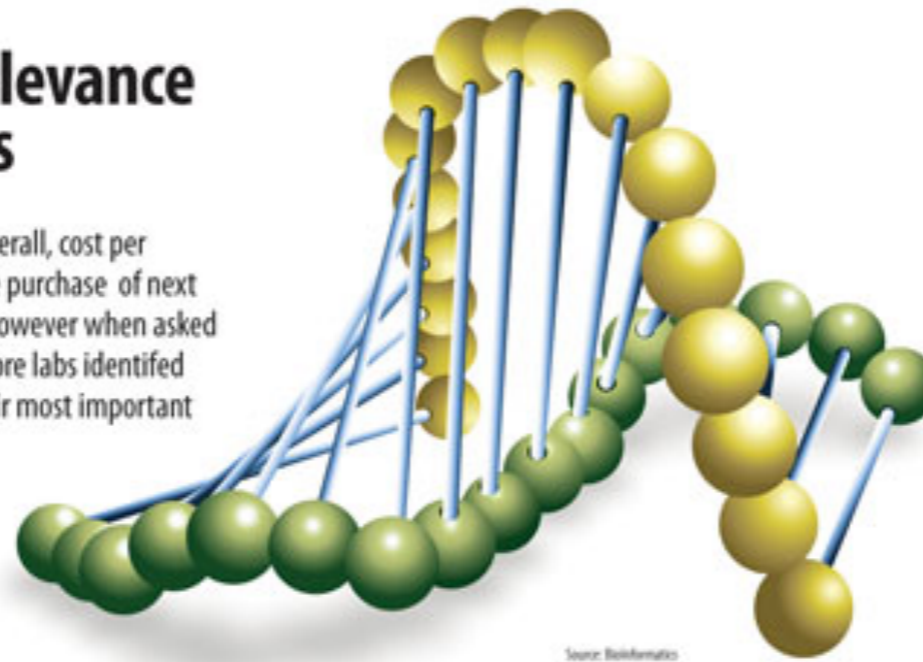
Structural variants
and smaller sample
size



NGS: Cost and Relevance Are Key to Buyers

Recent market research shows that, overall, cost per base was the most cited concern in the purchase of next generation sequencing instruments. However when asked to identify their top three concerns, more labs identified "Appropriate to My Application" as their most important criteria.

The 10 Most Critical Platform Attributes as Defined by Purchasers

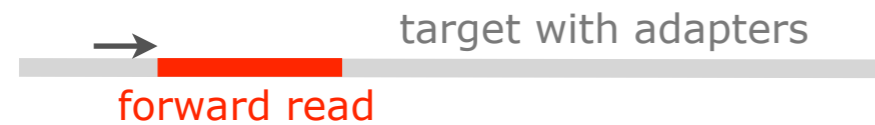


Source: Bioinformatics

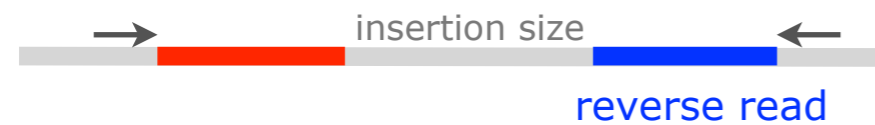
1. Cost per base	43%	6. Read length	24%
2. Sequencing data quality	34%	7. Instrument cost	18%
3. Appropriate for my application	32%	8. Number of reads	17%
4. Reproducibility/accuracy	31%	9. Available software analysis tools	16%
5. Amount of DNA/RNA needed per experiment	25%	10. Instrument reliability	16%

Sequencing Data

Illumina Sequence Read Data



Single Reads (SR)



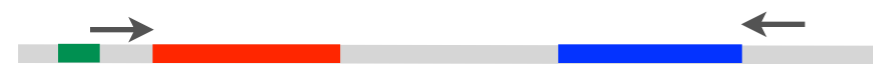
Paired-End (PE) Reads



Overlapping Paired-End (PE) Reads



Single Reads (SR) with Index



Paired-End (PE) Reads with Index



Paired-End (PE) Reads with Dual Indexing

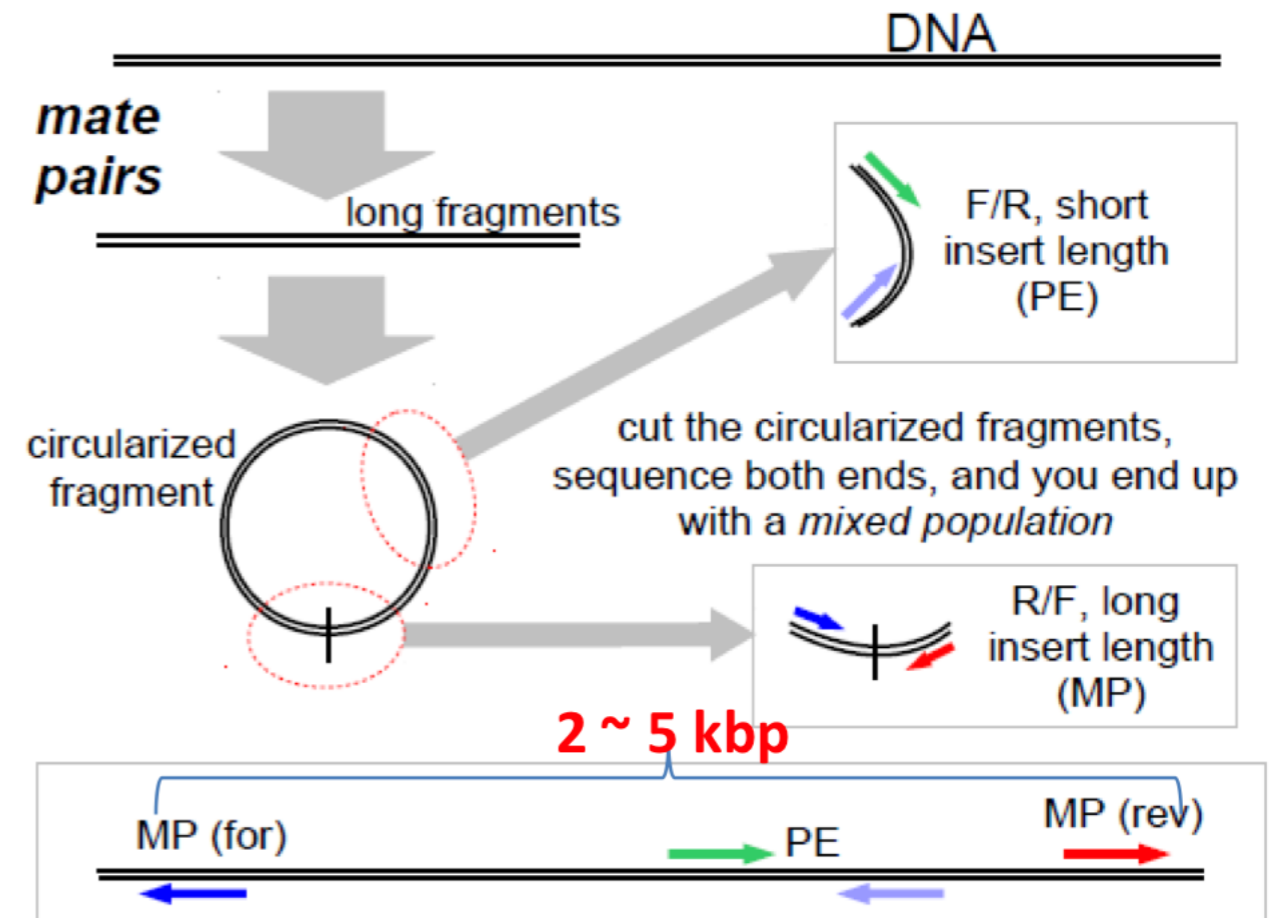
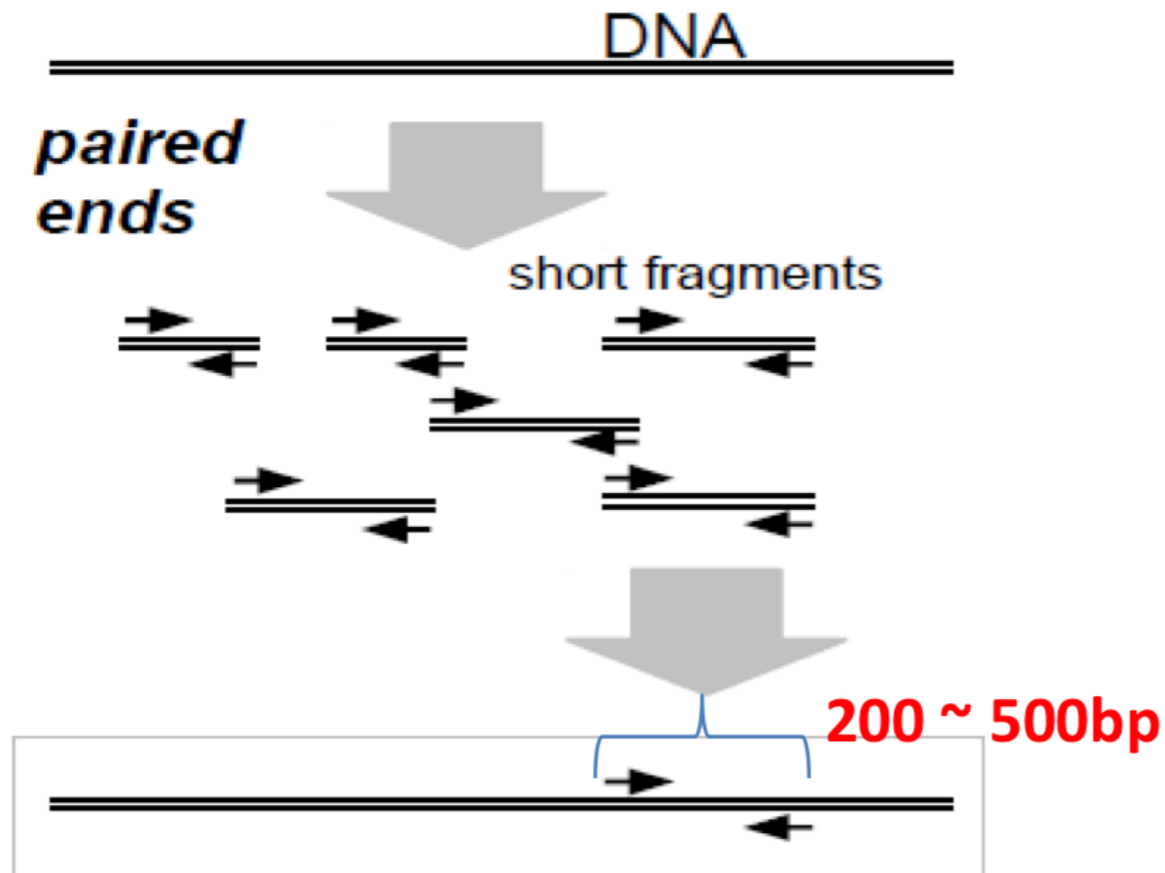


Extended Single Reads (SR) with Index

Illumina Sequence Read Data

paired-end (PE)

mate-pair (MP)



PacBio SMRTbell Library



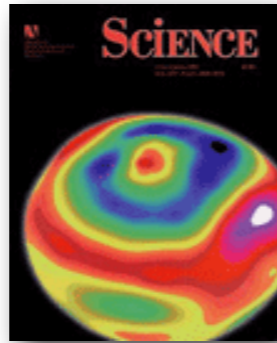
1995



1996



1997



1998



2000



2000



2001



2002



2002



2002



2002



2004



2004



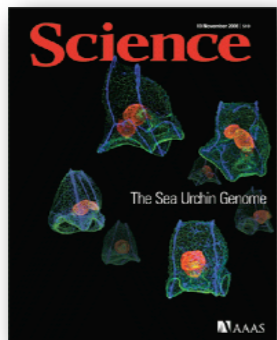
2005



2005



2006



2006



2007



2007



2008



2008



2009



2009



2010



2011



2011

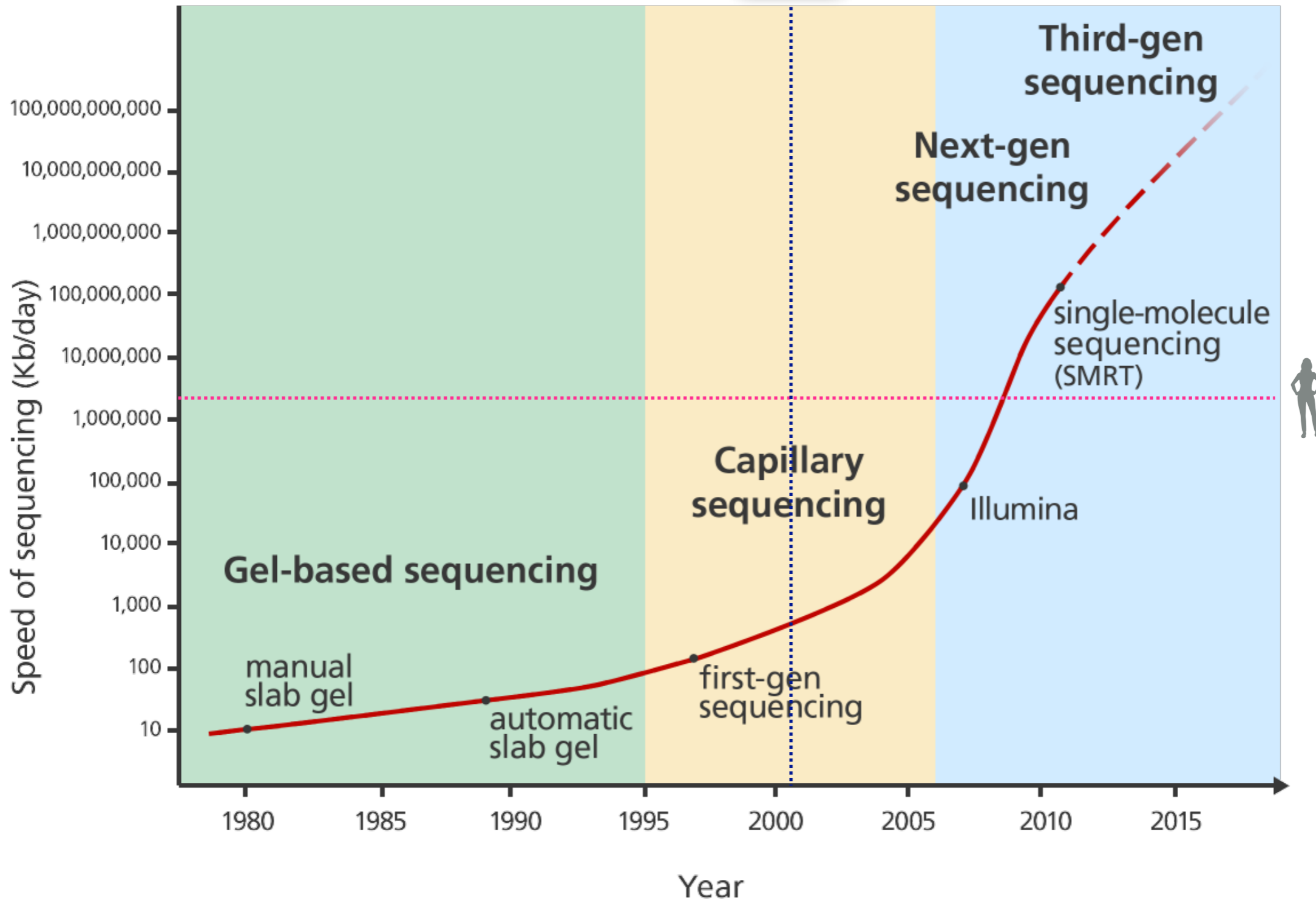


2011



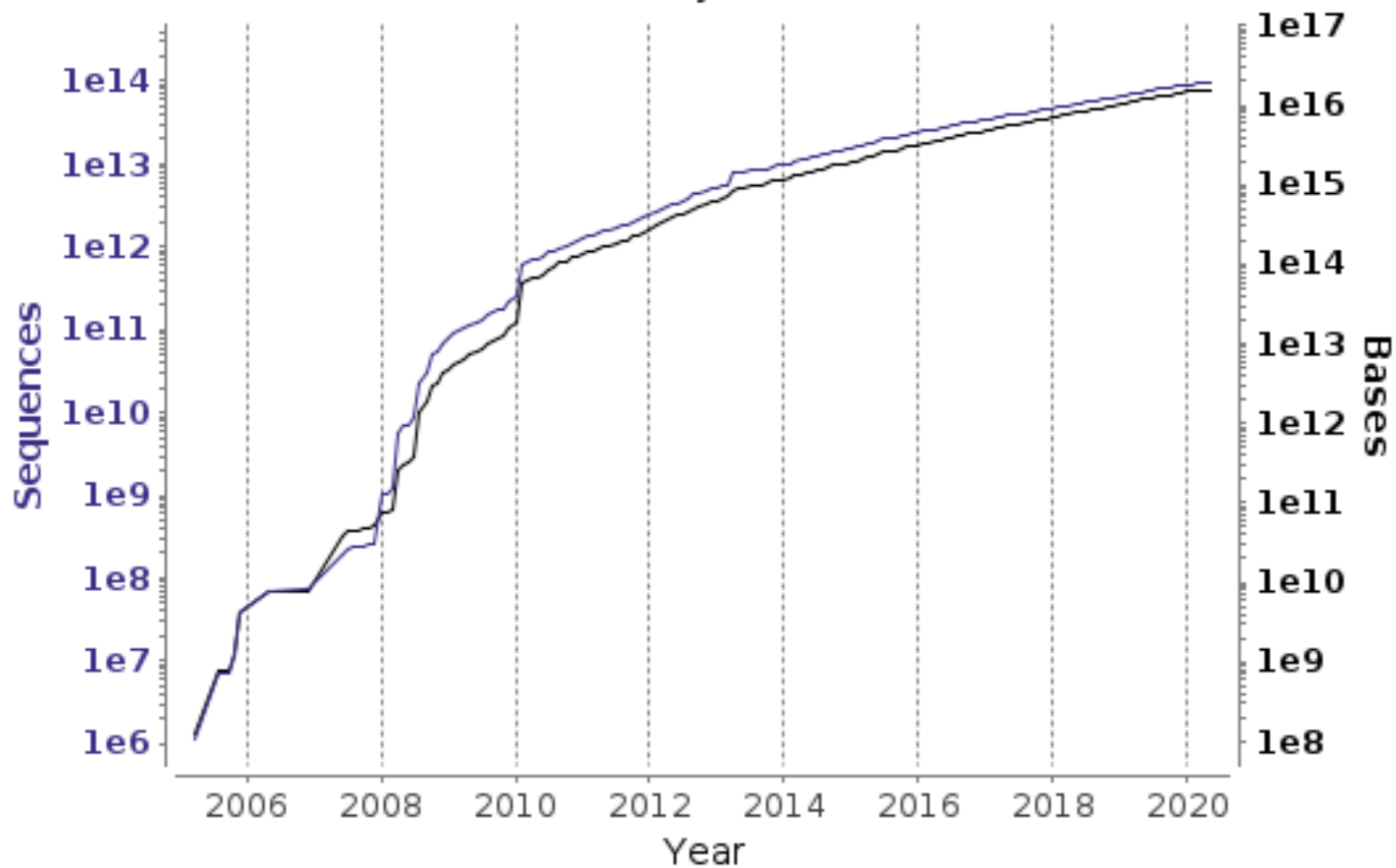
2012





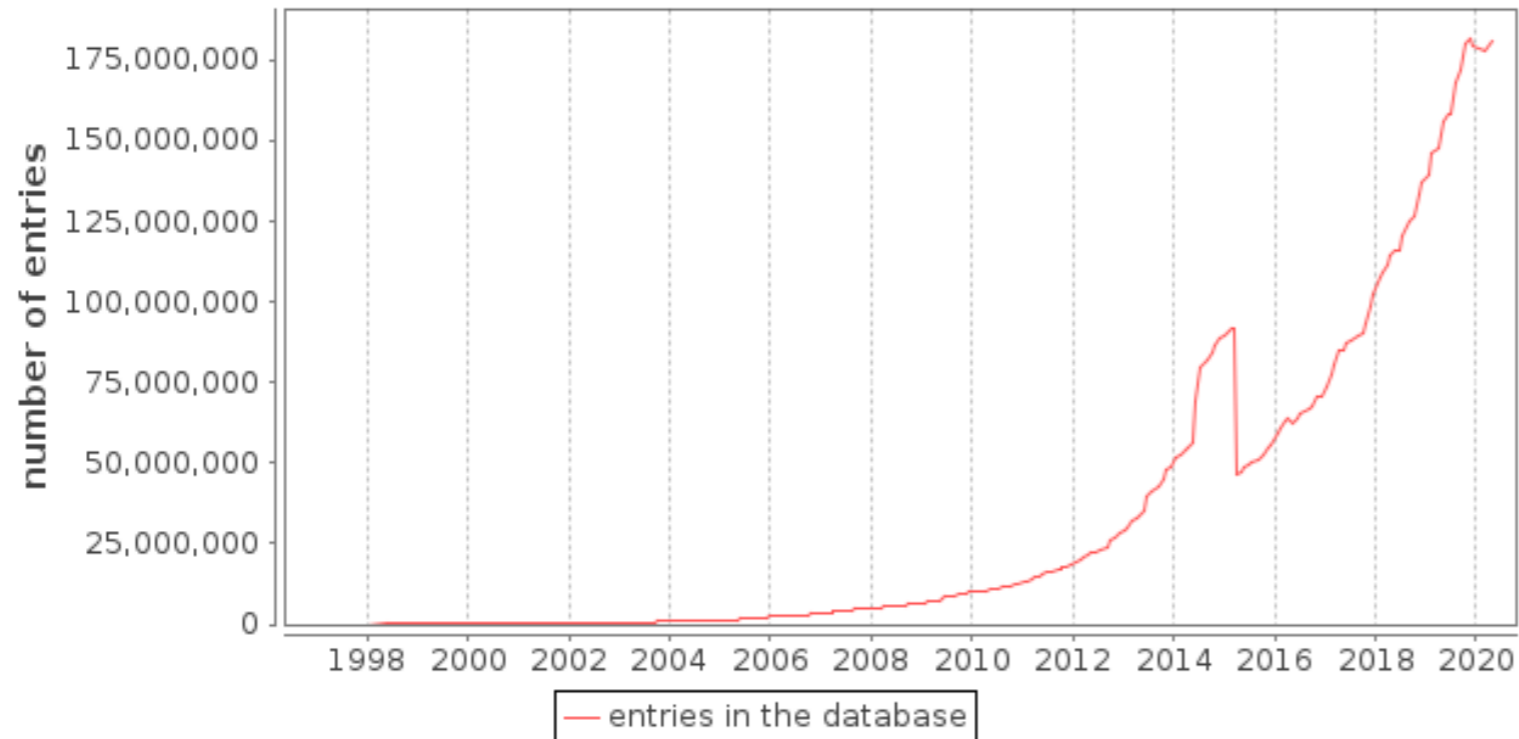


Reads growth
18-May-2020



— Sequences (95.7 trillions) — Bases (15,775.5 trillions)

Number of entries in UniProtKB/TrEMBL over time



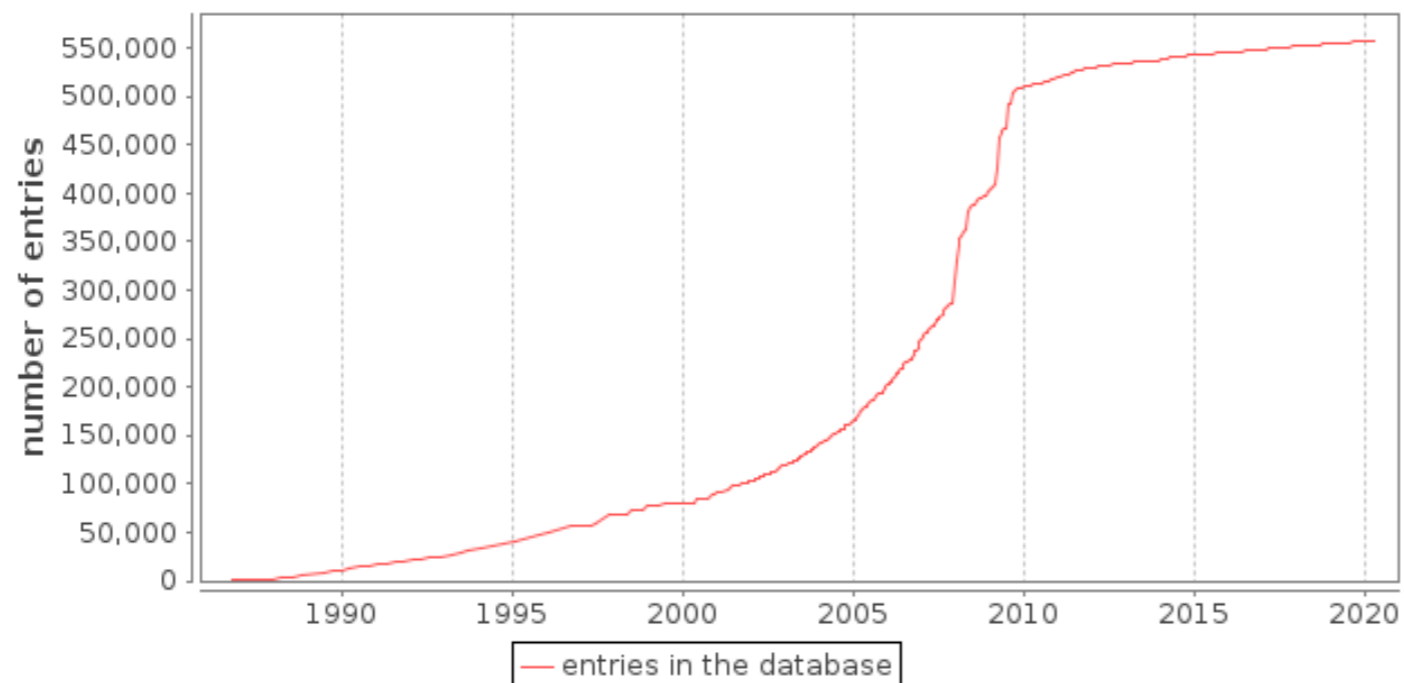
The **UniProt Knowledgebase (UniProtKB)** is the central hub for the collection of functional information on proteins, with accurate, consistent and rich annotation.

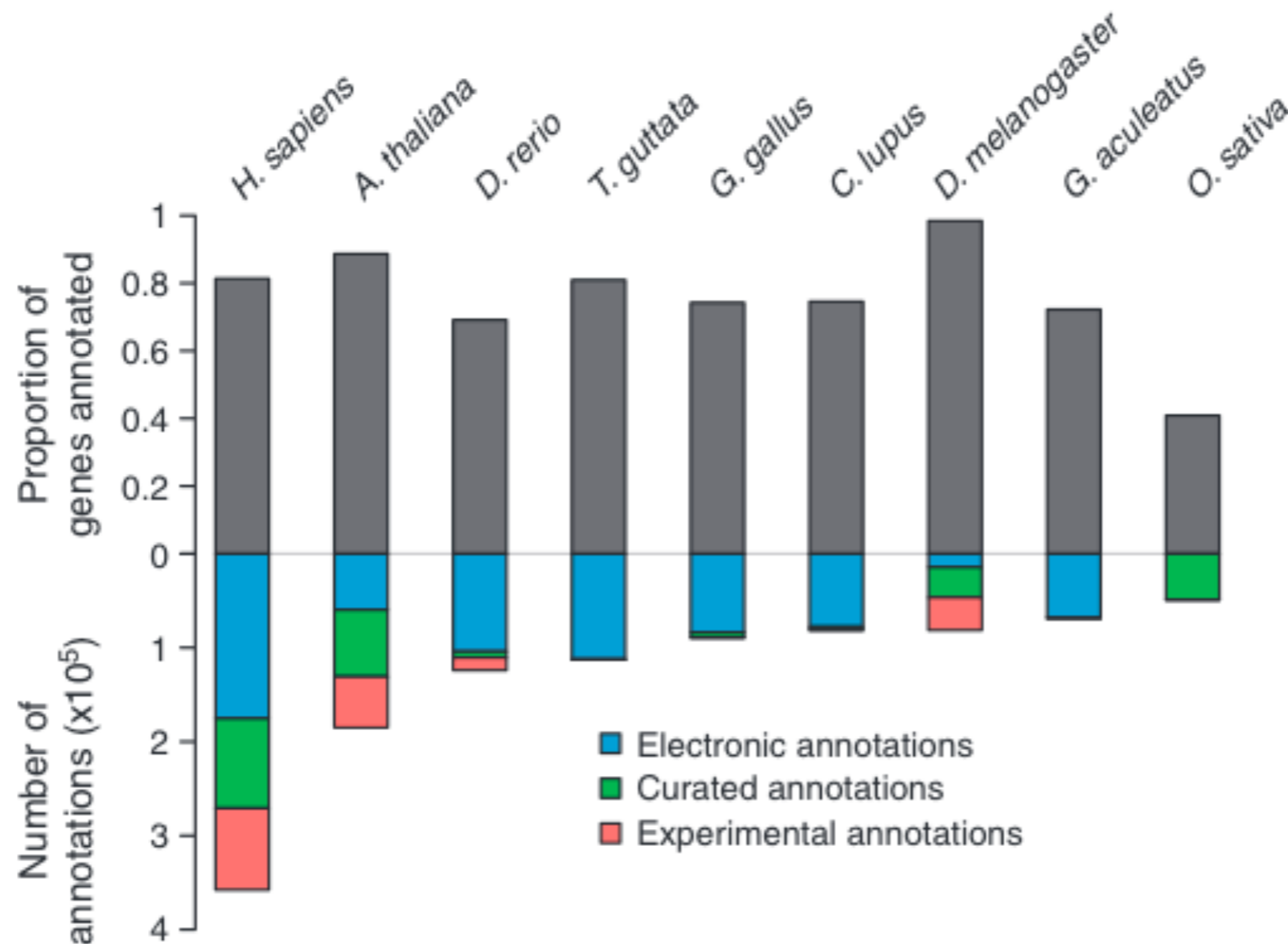
UniProtKB consists of two sections:

Reviewed (Swiss-Prot) - Manually annotated
Records with information extracted from literature and curator-evaluated computational analysis.

Unreviewed (TrEMBL) - Computationally analyzed
Records that await full manual annotation.

Number of entries in UniProtKB/Swiss-Prot over time





The proportion of annotated genes and their types of annotations for nine sequenced genomes (as of February 2013). Humans (*Homo sapiens*) and *Arabidopsis thaliana* have the highest number of annotations for animals and plants, respectively. They also have the most experimentally derived annotations. Most other species, except *Drosophila melanogaster*, are annotated mostly electronically.

Primmer et al. (2013) Mol Ecol

Data Submission



The European Nucleotide Archive (ENA) captures and presents information relating to experimental workflows that are based around nucleotide sequencing. A typical workflow includes the isolation and preparation of material for sequencing, a run of a sequencing machine in which sequencing data are produced and a subsequent bioinformatic analysis pipeline. ENA records this information in a data model that covers input information (sample, experimental setup, machine configuration), output machine data (sequence traces, reads and quality scores) and interpreted information (assembly, mapping, functional annotation).



Sequence Read Archive (SRA) makes biological sequence data available to the research community to enhance reproducibility and allow for new discoveries by comparing data sets. The SRA stores raw sequencing data and alignment information from high-throughput sequencing platforms, including Roche 454 GS System®, Illumina Genome Analyzer®, Applied Biosystems SOLiD System®, Helicos Heliscope®, Complete Genomics®, and Pacific Biosciences SMRT®.

Data Deposition Example from the Literature:

Mushegian *et al.* (2018) **Environmental sources of bacteria and genetic variation in behavior influence host-associated microbiota**. AEM doi:10.1128/AEM.01547-18.

Sequence data are deposited in the **European Nucleotide Archive of the EBI under accession number PRJEB30308** (<http://www.ebi.ac.uk/ena/data/view/PRJEB30308>). Data tables, OTUs sequences and code used for analysis can be found on **Github** at <https://github.com/amusheg/Daphnia-microbiota-behavior> and will be deposited in **Dryad** upon publication.

EMBL-EBI

[Services](#)
[Research](#)
[Training](#)
[About us](#)

Examples: [BN000065](#), [histone](#)
[Advanced Sequence](#)

Home
Search & Browse
Submit & Update
Software
About ENA
Support

Study: PRJEB30308

Microbiota associated with Daphnia exhibiting genetic variation in behavior

View: [Project XML](#) [Study XML](#)

Download: [Project XML](#) [Study XML](#)

Name	Submitting Centre
Microbiota of browsing Daphnia	Universitaet Basel
Secondary accession(s)	
ERP112744	

Description

In many organisms, host-associated microbial communities are acquired horizontally after birth. This process is believed to be shaped by a combination of environmental and host genetic factors. We examined whether genetic variation in animal behavior could affect the composition of the animal's microbiota in different environments. The freshwater crustacean Daphnia magna is primarily planktonic, but exhibits variation in the degree to which it browses in benthic sediments. We performed an experiment with clonal lines of D. magna showing different levels of sediment-browsing intensity exposed to either bacteria-rich or bacteria-poor sediment or whose access to sediments was prevented. We find that the bacterial composition of the environment and genotype-specific browsing intensity together influence the composition of the Daphnia-associated bacterial community. Exposure to more diverse bacteria did not lead to a more diverse microbiome, but greater abundances of environment-specific bacteria were found associated with host genotypes that exhibited greater browsing behavior. Our results indicate that, although there is a great deal of variation between individuals, behavior can mediate genotype-by-environment interaction effects on microbiome composition.

Navigation
Read Files
Portal
Attributes

Bulk Download Files ⚠ (If the downloader app doesn't open, please try using Firefox to launch it.)

Download: - of 512 results in [TEXT](#)

[Select columns](#)

Showing results 1 - 10 of 512 results

Study accession	Sample accession	Secondary sample accession	Experiment accession	Run accession	Tax ID	Scientific name	Instrument model	Library layout	FASTQ files (FTP)	FASTQ files (Galaxy)	Submitted files (FTP)	Submitted files (Galaxy)	NCBI SRA file (FTP)	NCBI SRA file (Galaxy)	CRAM Index files (FTP)	CRAM Index files (Galaxy)
PRJEB30308	SAMEA5166093	ERS2973813	ERX2993334	ERR2990925	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166094	ERS2973814	ERX2993335	ERR2990926	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166095	ERS2973815	ERX2993336	ERR2990927	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		
PRJEB30308	SAMEA5166096	ERS2973816	ERX2993337	ERR2990928	1869227	bacterium	Illumina MiSeq	PAIRED	File 1 File 2	File 1 File 2	Fastq file 1 Fastq file 2	Fastq file 1 Fastq file 2	File 1	File 1		

NCBI Resources How To Sign in to NCBI

BioProject Help

[Create alert](#) [Advanced](#) [Browse by Project attributes](#)

Display Settings: ▾

Send to: ▾

Microbiota of browsing Daphnia

Accession: PRJEB30308 ID: 516850

Microbiota associated with Daphnia exhibiting genetic variation in behavior

In many organisms, host-associated microbial communities are acquired horizontally after birth. [More...](#)

Accession	PRJEB30308
Scope	Monoisolate
Submission	Registration date: 24-Jan-2019 Universitaet Basel

Project Data:

Resource Name	Number of Links
SEQUENCE DATA	
SRA Experiments	512
OTHER DATASETS	
BioSample	512

▾ SRA Data Details

Parameter	Value
Data volume, Gbases	22
Data volume, Mbytes	14805

Related information

[BioSample](#)

[SRA](#)

Recent activity

[Turn Off](#) [Clear](#)

PRJEB30308 (1)

[BioProject](#)

[Microbiota of browsing Daphnia](#)

[BioProject](#)

[The European Nucleotide Archive in 2017](#)

[A Benchmark Study on Error Assessment and Quality Control of CCS Reads Derived](#)

[Testing the potential of a ribosomal 16S marker for DNA metabarcoding of insects](#)

[See more...](#)

1

Choose the NGS technology according to your needs.

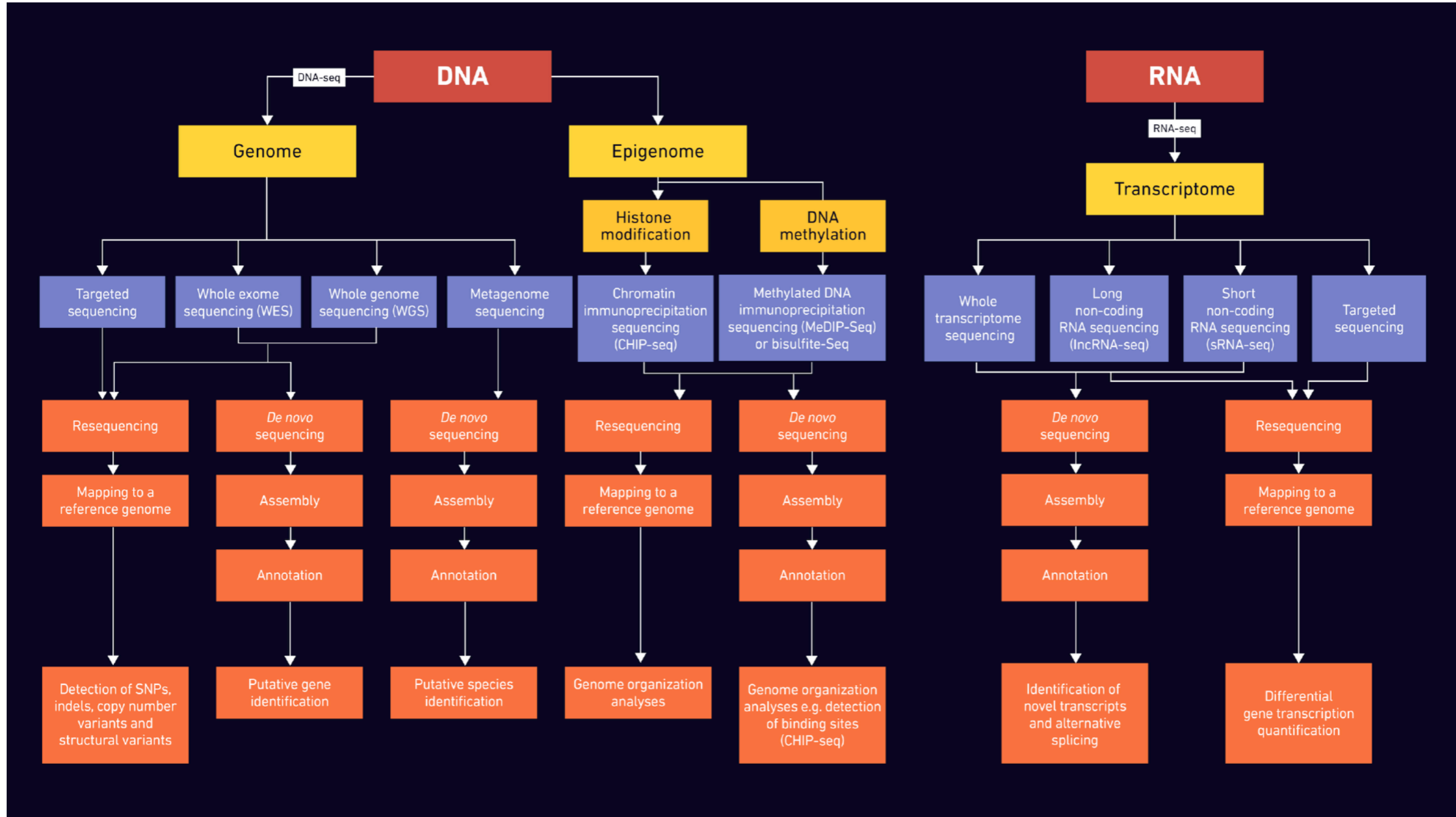
2

Keep your raw data safe and submit it as early as possible.

3

Coping one file (archive) is safer than coping multiple files.

NGS Applications



Flow diagram indicating possible sequencing strategies for different sample types.

www.technologynetworks.com

Table 1: A table of advantages and disadvantages for short vs long read sequencing.

	Advantages	Limitations
Short-read sequencing	<ul style="list-style-type: none"> · Higher sequence fidelity · Cheap · Can sequence fragmented DNA 	<ul style="list-style-type: none"> · Not able to resolve structural variants, phasing alleles or distinguish highly homologous genomic regions · Unable to provide coverage of some repetitive regions
Long-read sequencing	<ul style="list-style-type: none"> · Able to sequence genetic regions that are difficult to characterize with short-read seq due to repeat sequences · Able to resolve structural rearrangements or homologous regions · Able to read through an entire RNA transcript to determine the specific isoform · Assists <i>de novo</i> genome assembly 	<ul style="list-style-type: none"> · Lower per read accuracy · Bioinformatic challenges, caused by coverage biases, high error rates in base allocation, scalability and limited availability of appropriate pipelines

www.technologynetworks.com

EXTRAS

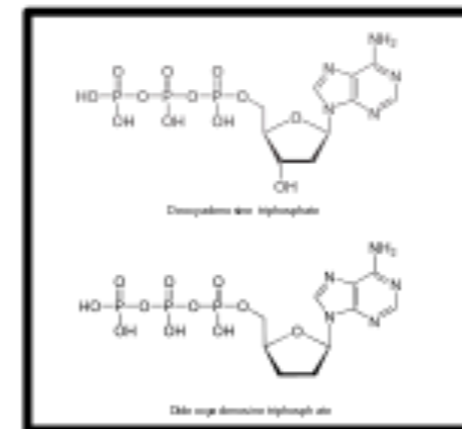
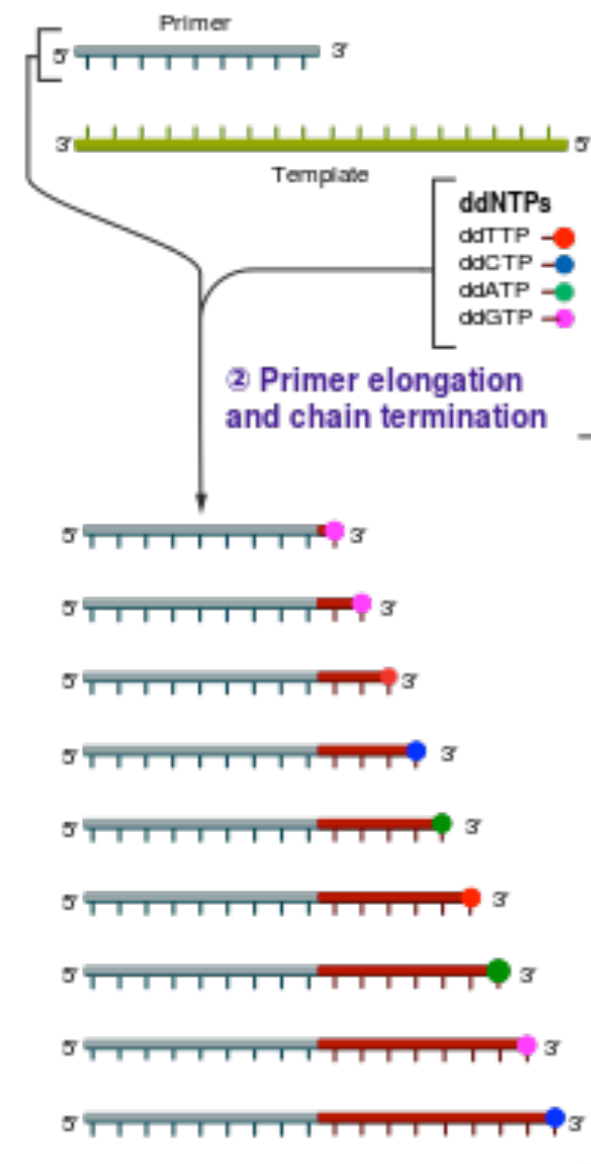
Sequencing Technologies

Extended

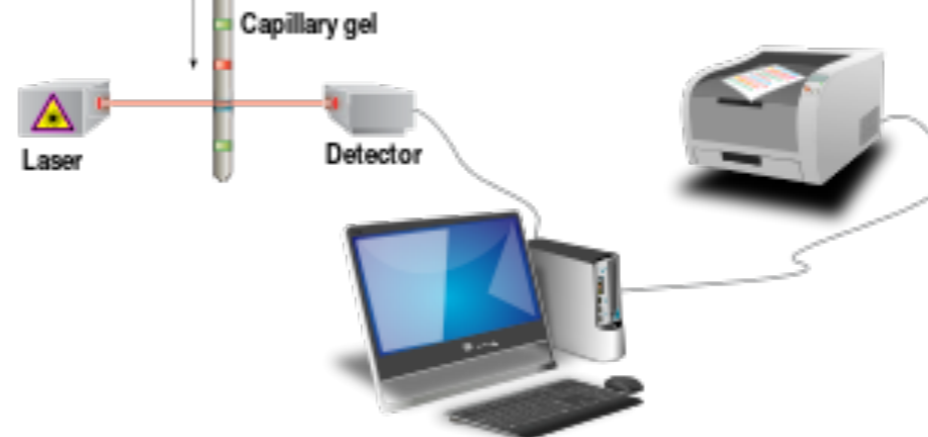
Capillary sequencing

① Reaction mixture

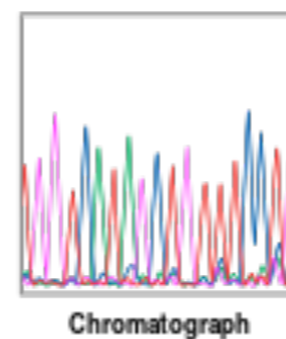
- ▶ Primer and DNA template
- ▶ DNA polymerase
- ▶ ddNTPs with flouochromes
- ▶ dNTPs (dATP, dCTP, dGTP, and dTTP)



③ Capillary gel electrophoresis separation of DNA fragments



④ Laser detection of flouochromes and computational sequence analysis



Pyrosequencing



GS Junior

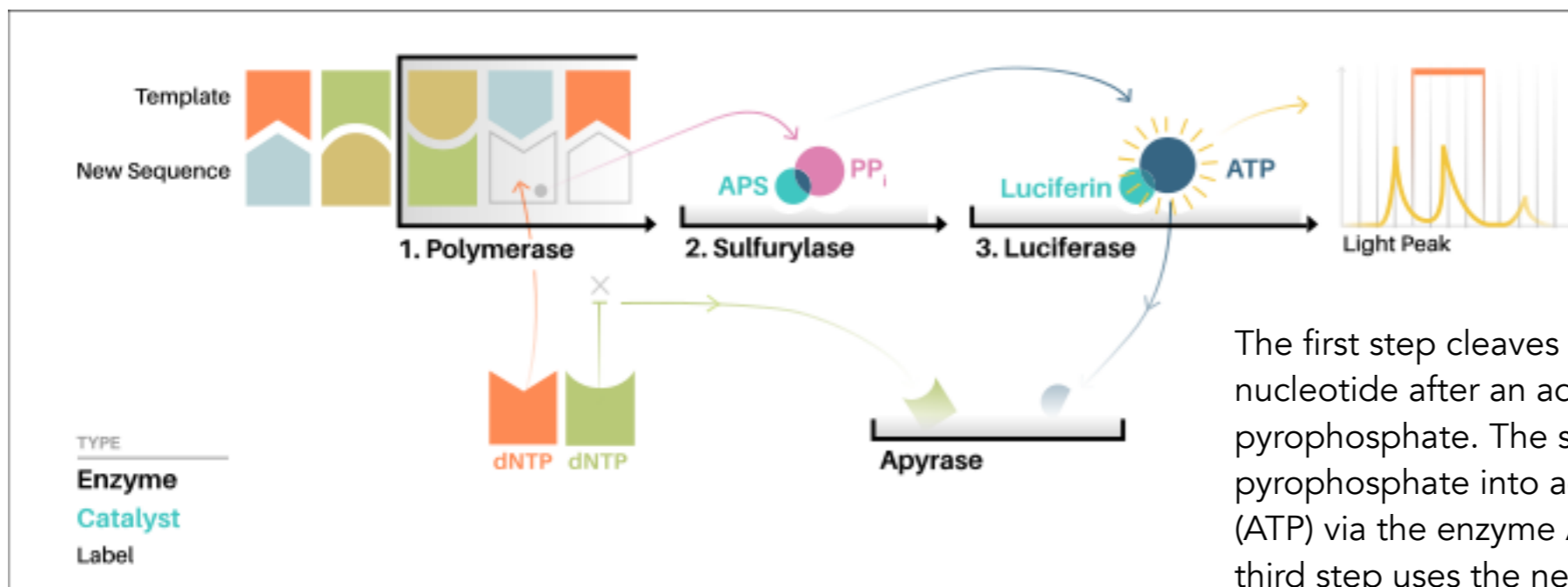


Roche 454



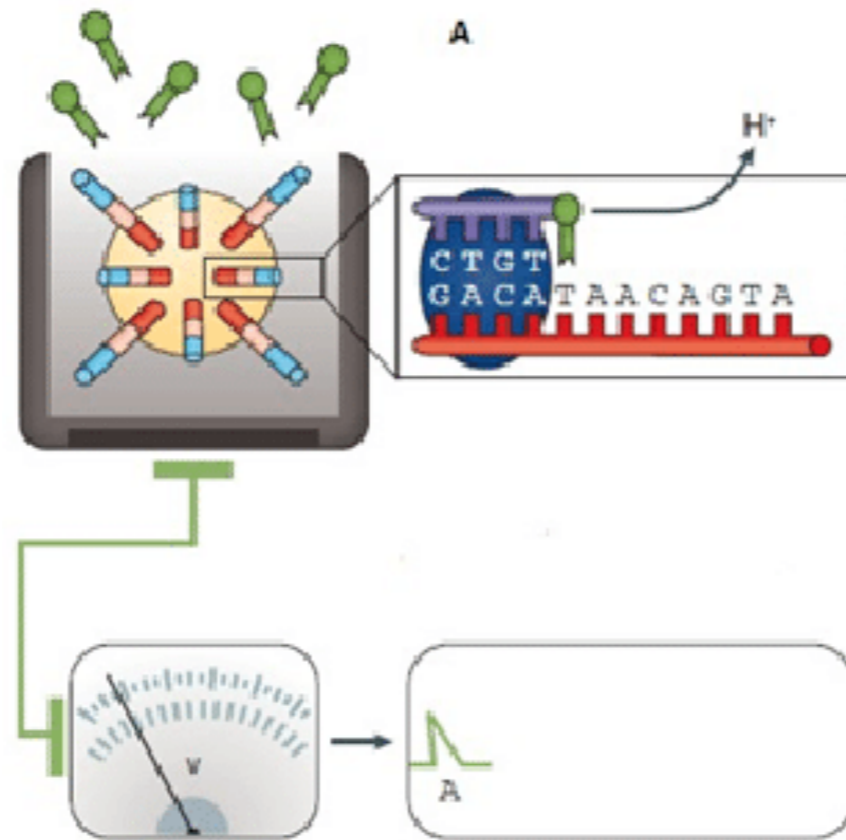
The **PyroMark** uses Pyrosequencing technology for real-time, sequence-based detection and quantification of sequence variants and epigenetic methylation. The PyroMark Q24 is highly suited for the analysis of CpG methylation, SNPs, insertion/deletions, STRs, variable gene copy number, as well as for microbial identification and resistance typing.

Pyrosequencing (pyrophosphate)



The first step cleaves the triphosphate nucleotide after an addition, releasing pyrophosphate. The second step converts pyrophosphate into adenosine triphosphate (ATP) via the enzyme ATP sulfurylase. The third step uses the newly synthesized ATP to catalyze the conversion of luciferin into oxyluciferin via the enzyme luciferase and this reaction generates a quanta of light that is captured from the picotiter plate by a charge- coupled camera.

Ion Torrent (semiconductor technology)





MiniSeq System

1.8-7.5 Gb
8-25 million
2 x 150 bp
50



MiSeq Series

0.3-15 Gb
1-25 million
2 x 300 bp
384



NextSeq Series

20-120 Gb
130-400 million
2 x 150 bp
96



HiSeq Series

125-1500 Gb
2.5-5 billion
2 x 150 bp
12



HiSeq X Series

900-1800 Gb
3-6 billion
2 x 150 bp
16

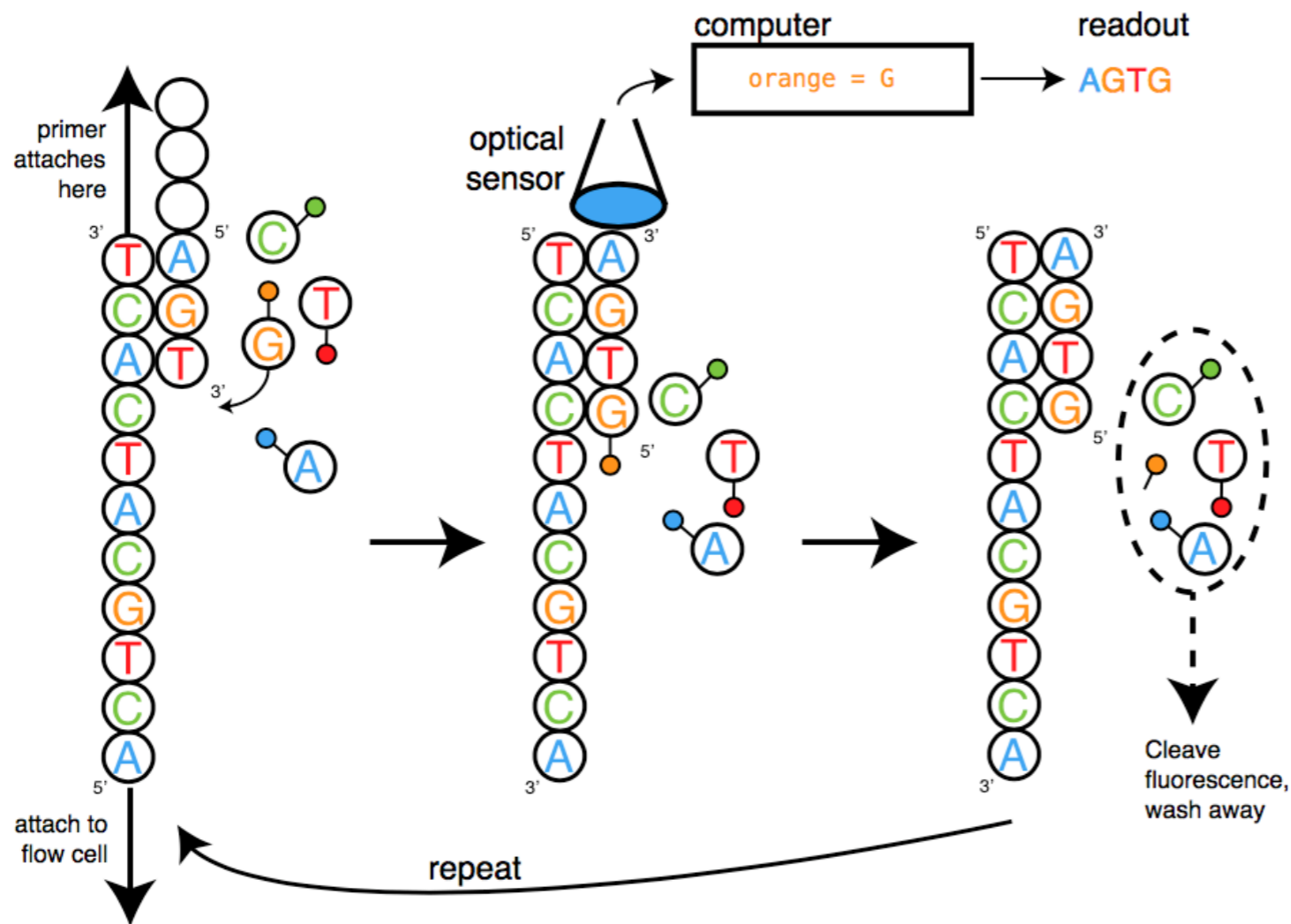


NovaSeq Series

134-6000 Gb
Up to 20 billion
2 x 150 bp
48

<http://www.illumina.com>

Sequencing by Synthesis (fluorescent)



Sequencing by Synthesis. dNTP fluorescence is translated to a base call.



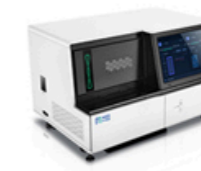
Sequencers +







Sequencers +



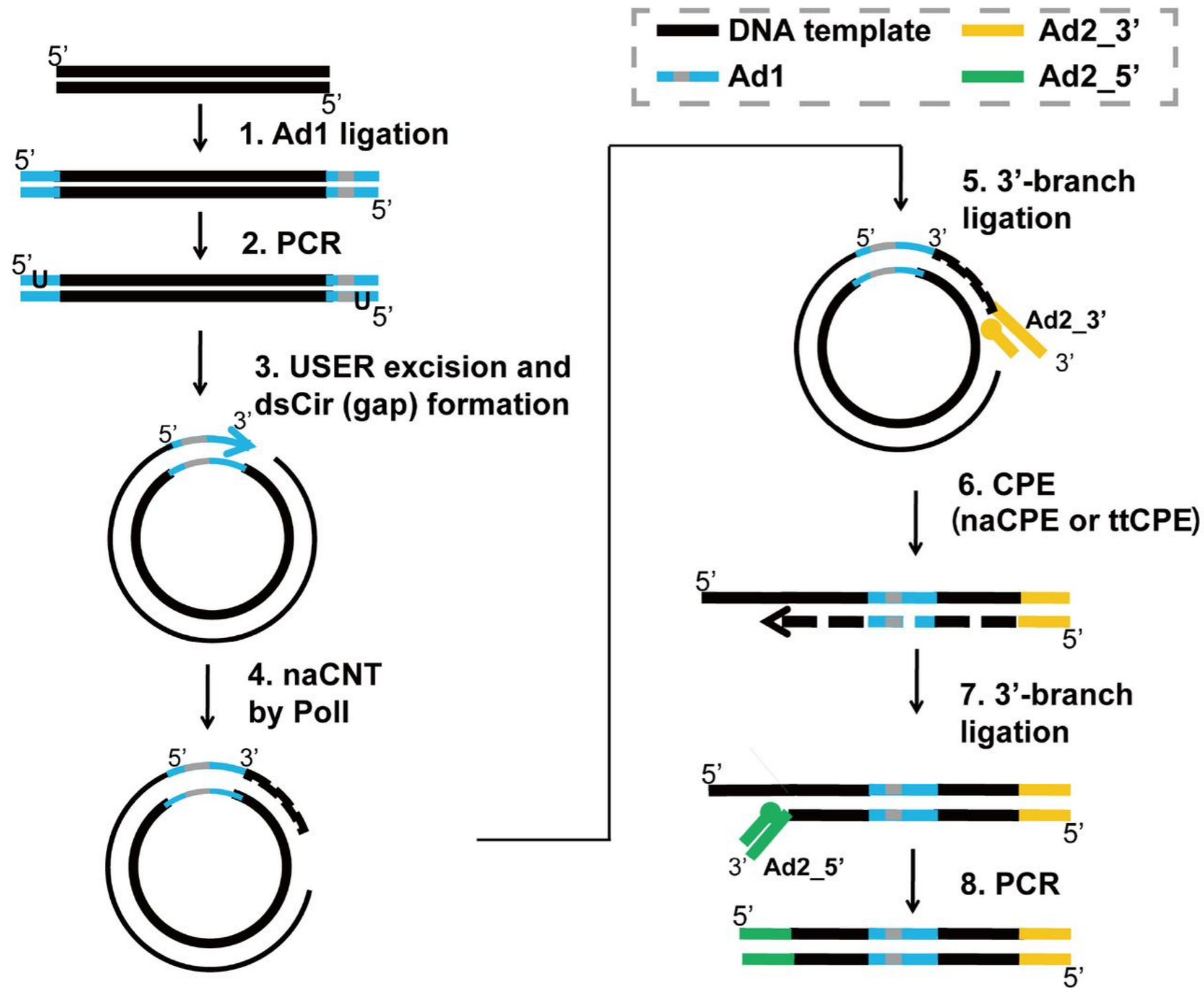
Sequencers +



Sequencers +

				
Product Model	DNBSEQ-T7	DNBSEQ-G400	DNBSEQ-G50	DNBSEQ-G400 FAST
Features	Ultra-high Throughput	Adaptive	Effective	Fast
Applications	Whole Genome Sequencing, Deep Exome Sequencing, Transcriptome Sequencing, and Targeted Panel Projects.	WGS, WES, Transcriptome sequencing and more	Small whole genome sequencing, targeted DNA/RNA panels, low-pass whole genome sequencing	Targeted DNA, RNA, Epigenetics and clinical applications
Flow Cell Type	FC	FCL & FCS	FCL & FCS	FCS
Lane/Flow Cell++	1 lane	4 lane & 2 lane	1 lane	2 lane
Operation Mode	Ultra-high Throughput	High Throughput	Medium Throughput	Medium Throughput
Max. Throughput / RUN	6Tb	1440Gb	150Gb	330G
Effective Reads / Flow Cell	5000M	1500-1800M	500M / 100M	550M
Average run time	PE150 within 24 hours	~38 hours	10-66 hours	12-37 hours
Min. Read Length	PE100	SE50	SE50	SE100
Max. Read Length	PE150	SE400	PE150	PE150

<https://en.mgitech.cn>

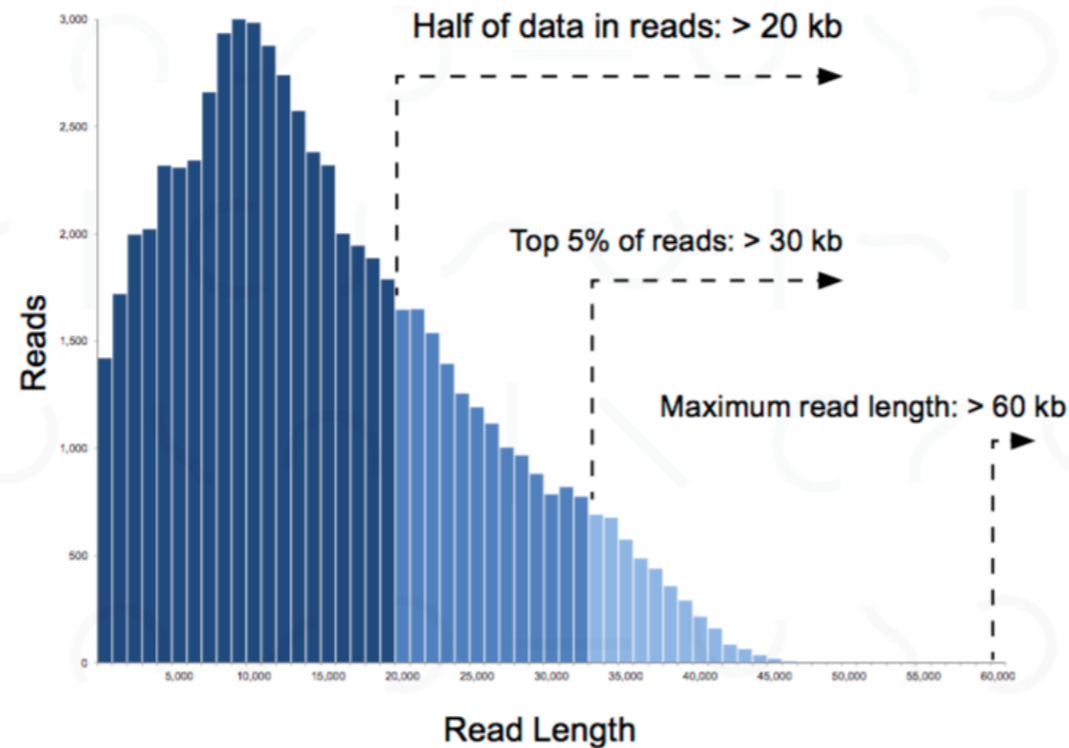




PacBio RS II

Long Read Lengths

Read lengths > 20 kb
Data per SMRT Cell: 750 Mb - 1.25 Gb

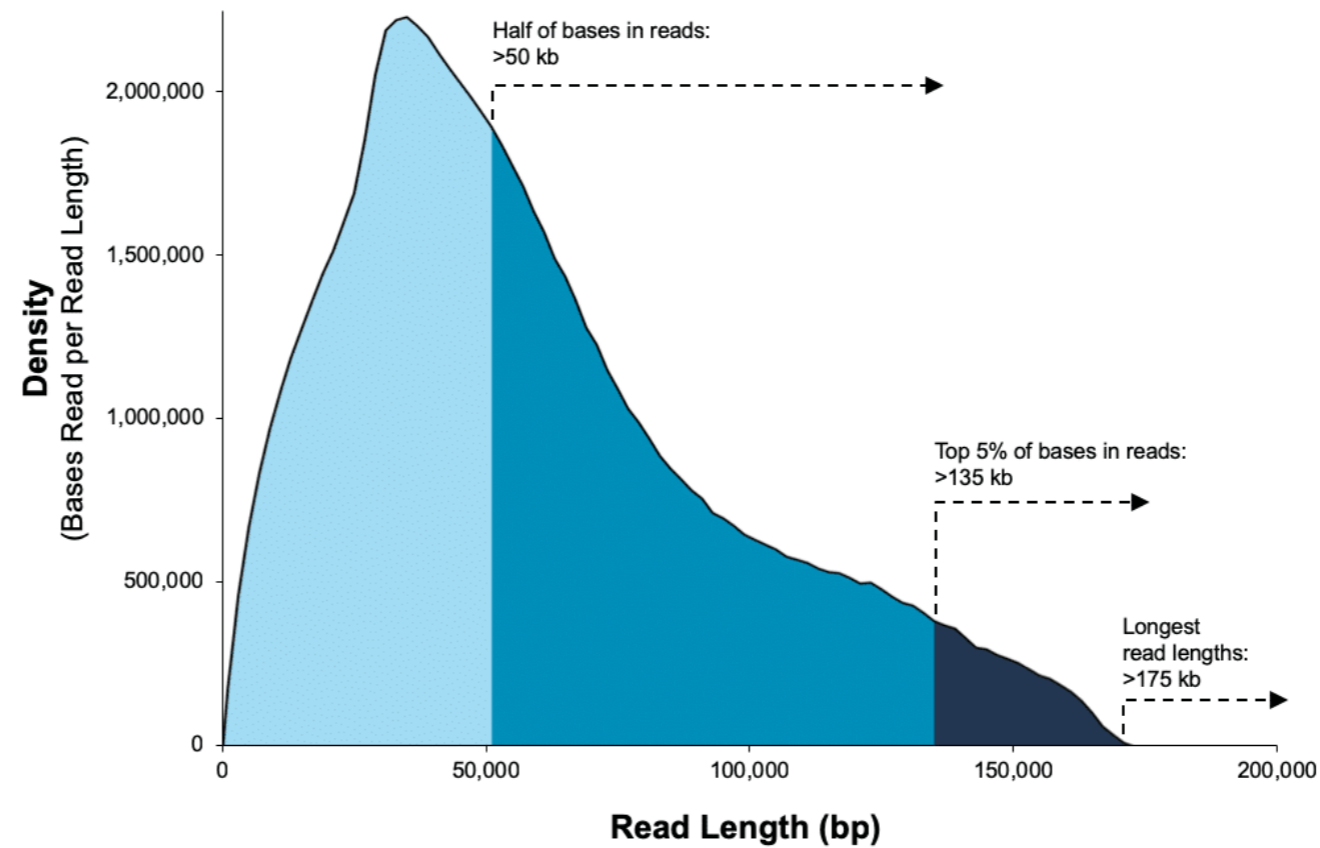


Read-length data shown above is from a 20 kb size-selected human library run on a PacBio RS II (6-hour movie, P6-C4 chemistry). The PacBio RS II SMRT Cells generate ~55,000 reads. The Sequel System generates ~370,000 reads per SMRT Cell.



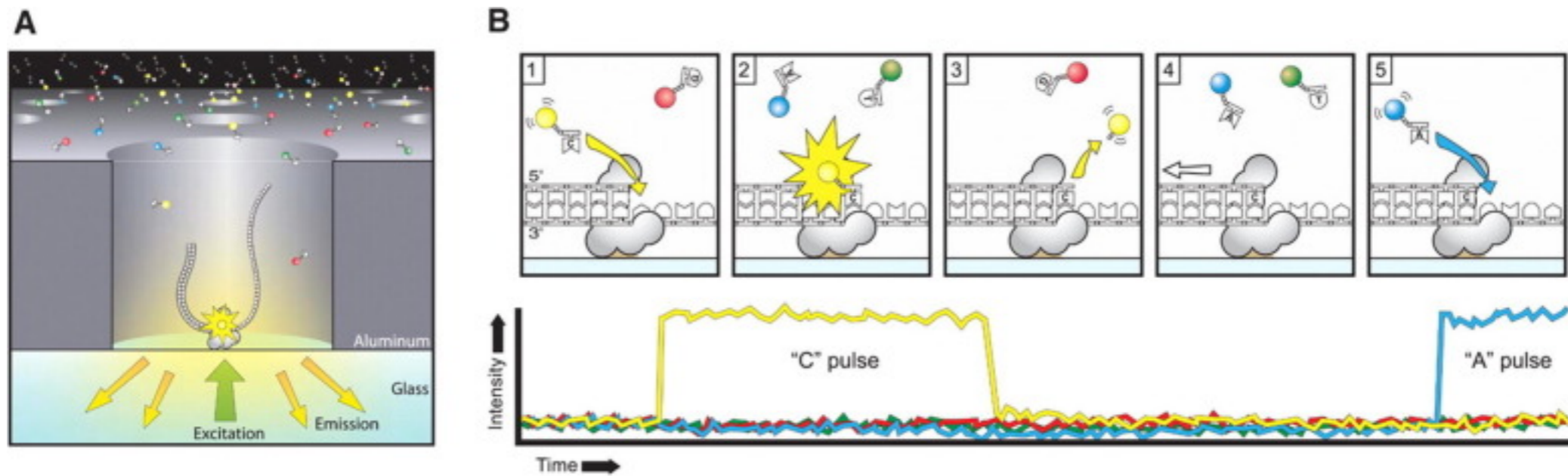


Sequel



Data from a 35 kb size-selected *E. coli* library using the SMRTbell Express Template Prep Kit 2.0 on a Sequel II System (1.0 Chemistry, Sequel II System Software v7.0, 15-hour movie)*.

PacBio (fluorophore)





SmidgION



Flongle



MinION

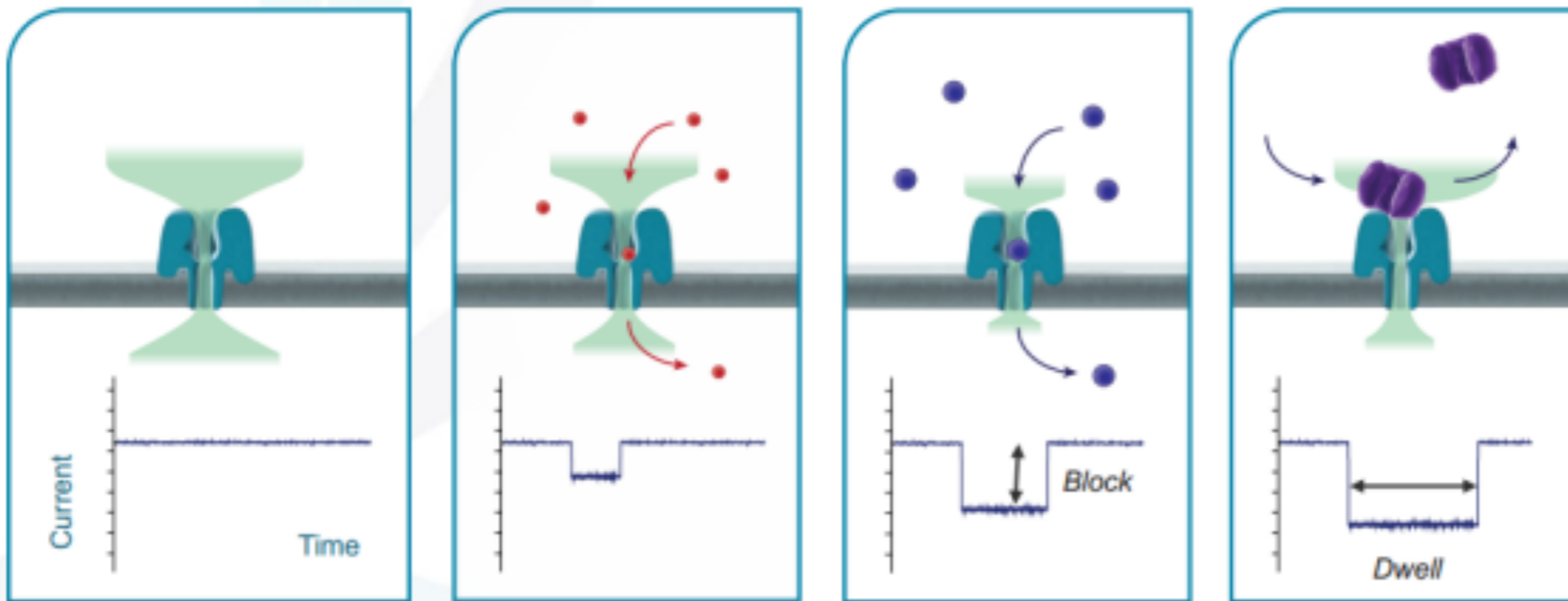


GridION



PromethION

<https://www.nanoporetech.com>

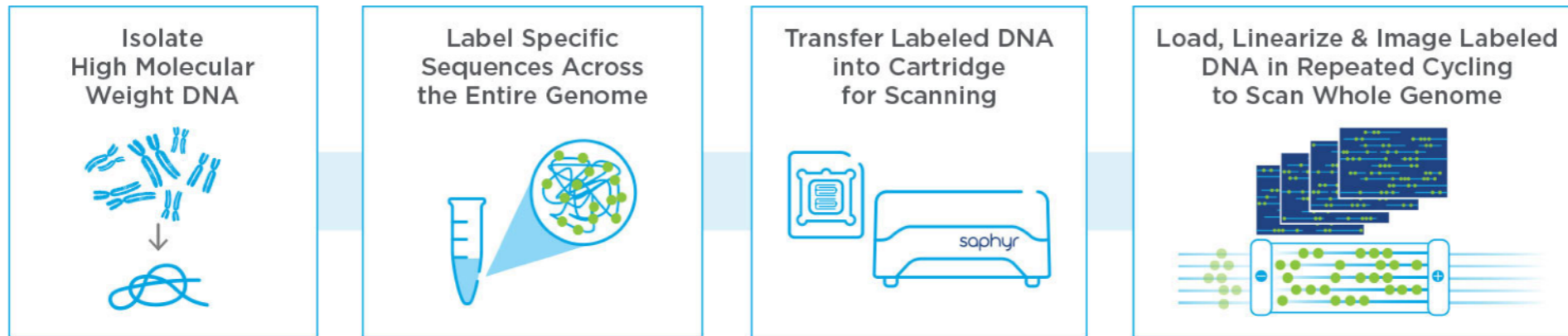






Customer Sample

- Blood
- Tissue
- Cells
- Microbes



High-throughput, High-resolution Imaging of Megabase Length Molecules

